



Genetic polymorphism at the *KIR* gene locus: determination of gene, genotype, and haplotype frequencies in the Xinjiang Han population

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ABSTRACT. The aim of this study was to explore the genetic polymorphism, genotype, and haplotype characteristics of the *KIR* locus in the Xinjiang Han population in order to establish a foundation for future analysis of the relationship between *KIR* genes and disease. *KIR* genes were detected by sequence-specific primer-polymerase chain reaction in 184 randomly selected, healthy individuals from the Han population in Xinjiang, China. Standard genotype and haplotype analyses were conducted using Hsu's standards classified for analysis. Sixteen *KIR* genes were detected: *3DL3*, *2DL4*, *3DL2*, and *3DL2* (100%); *2DL1* and *2DP1* (99.46%); *2DL3* (98.91%); and so on. The *2DS2* gene frequency was the lowest at 21.74%. Twenty-one genotypes were detected: AJ (2, 2) was relatively common (42.39%), followed by AH (5, 2), AE (2, 8) and H (2, 4), with frequencies of 17.39, 11.96, and 8.15%, respectively. In addition, six novel genotypes were identified in 11 Han individuals as well as in other populations in China, which could not be classified for analysis. These results indicated that the Xinjiang Han population shares *KIR* gene,

genotype, and haplotype frequency distributions with the Chinese Han population, but also has unique genotypes and haplotypes.

Key words: Killer cell immunoglobulin-like receptor (KIR); Polymerase chain reaction-sequence specific primers; Genetic polymorphism; Genotype; Haplotype

INTRODUCTION

Killer cell immunoglobulin-like receptors (KIRs) are a group of specifically recognized human major histocompatibility antigen (MHC-I) molecule receptors expressed on the surface of natural killer (NK) cells and T cells. *KIR* has been shown to be the next most highly diverse immune regulatory gene family after the discovery of the human leukocyte antigen (HLA) cluster, and KIR receptors have important immunomodulatory effects on NK and T cells. The KIR-encoding locus is on chromosome 19q13.4, and the family group includes 18 genes, namely *KIR1D*, *KIR2DL1-5*, *KIR2DS1-5*, *KIR3DL1-3*, *KIR3DS1*, *Xv*, *X*, and *KIR2DP1*. *Xv*, *X*, and *KIR2DP1* are pseudogenes, and *KIR2DL4*, *KIR3DL2*, and *KIR3DL3* are framework genes (Zhang et al., 2003a). Studies have found that the *KIR* genes are genetically polymorphic and that there is a richly diverse KIR receptor library in different individuals that are expressed in different NK cell clones. The NK cell KIR expression pattern directly regulates their ability to clear foreign and harmful substances, and the *KIR* genotype of an individual is the most critical factor to determine its expression profile, which plays an important role in the body's natural and acquired immunities. Therefore, *KIR* genotyping assists with the analysis of its function in certain diseases (Zhang et al., 2003b). Previous studies (Khakoo and Carrington, 2006) have shown that *KIR* locus polymorphism is associated with certain infectious and autoimmune diseases, cancer susceptibility, and pregnancy. The regulatory effects of KIR in hematopoietic stem cell transplantation have also been of great concern in the clinical field. This study focused on the multi-ethnic region of Xinjiang to determine the distribution characteristics of genes within the polymorphic *KIR* locus in the Xinjiang Han population. We utilized the sequence-specific primer polymerase chain reaction (SSP-PCR) to perform *KIR* genotyping and genotype and haplotype analysis of 184 cases of Han volunteers from Xinjiang. The results from this study will lay the foundation for further studies of the association between the *KIR* gene polymorphism and certain diseases in different groups of people.

MATERIAL AND METHODS

Study design

This study was a prospective analysis. DNA was analyzed *in vitro* using molecular biology detection methods. Study participants consisted of unrelated individuals recruited at random from the Xinjiang Han population. Our study was carried out from November 2009 to September 2011, and analyses were performed in the kidney transplant tissue typing room of the blood purification center in No. 474 People's Liberation Army Hospital.

General information

Study subjects consisted of kidney transplant HLA-matching donors and recipients

processed through the No. 474 People's Liberation Army Hospital. All subjects gave their written informed consent. A 4-mL venous whole blood sample was collected from donors and recipients and kept in ethylenediaminetetraacetic acid anticoagulant. Genomic DNA was obtained from 84 unrelated Han individuals from the Xinjiang region. DNA extraction was in strict accordance with the instructions of the PROTRANS DNA rapid extraction kit (Beijing San Taihua Biotechnology Company, Beijing, China). The A_{260}/A_{280} ratio of the extracted DNA was from 1.8 to 2.0. The genomic DNA samples were stored at -20°C for subsequent analysis.

Reagents and instruments

The *KIR* SSP-PCR kit was provided by Tianjin Xiu Peng Biotechnology Development Co., Ltd. (Tianjin, China). The internal reference control was the conservative fragment of the human growth hormone gene, and the amplification product of the internal reference was 588 bp Taq DNA polymerase (Promega, Shanghai China). The DNA extraction kit was the German PROTRANS DNA extraction kit (Beijing San Taihua Biotechnology Company). The PCR amplification instrument was from BIO-RAD Laboratories (MyCycler™ Thermal Cycler, USA). The electrophoresis instruments such as the DDY-BC and WD-9413B gel imaging analyzers were supplied by the Beijing Liuyi Instrument Company (Beijing, China).

Detection and genotyping of *KIR* genes

The SSP-PCR genotyping method for *KIR* genes was adopted in all samples collected. *KIR2DL1-5*, *KIR2DS1-5*, *KIR3DL1-3*, *KIR3DS1*, and the two pseudogenes *KIR2DPI* (aka *KIRZ*) and *KIR3DPI* (divided into *KIRX* and *KIRXv*) were detected from the *KIR* genetic locus. SSP-PCR amplification primers were designed based on the *KIR* gene sequence and genotyping literature design strategies, to provide a reference for these strategies or the program used for primer development, and a total of 32 pairs were synthesized by Tianjin Xiu Peng Super Biotechnology Co., Ltd. The human *KIR* genotype kit primer (locus and size) is shown in Table 1. Amplifications were performed in the BIO-RAD PCR amplification instrument. The PCR volume was 10 μL per well, and there were 16 wells per subject. The system contained 160 μL dNTP buffer, 20 μL genomic DNA, and 1.2 μL 5 U/ μL Taq DNA polymerase. The reaction conditions were as follows: 96°C for 2 min, 96°C for 20 s and 68°C for 60 s, for a total of 5 cycles; 96°C for 20 s, 64°C for 50 s, and 72°C for 45 s, for a total of 10 cycles; and 96°C for 20 s, 61°C for 50 s, and 72°C for 45 s, for a total of 18 cycles. Final extension was at 72°C for 5 min. The PCR products were loaded on a 2.5% agarose gel. Electrophoresis was performed at 160 V for 12 min, and bands were visualized using the WD-9413B ultraviolet gel imaging analyzer.

Genotype and haplotype analysis

We adopted the 36 *KIR* genotypes A to AJ, first defined in Caucasians by Hsu et

al. (2002). Genotypes and the designated 23 common haplotypes were determined using standard typing methods. The *KIR* haplotype combinations were projected according to the genotypes. When the same genotype had 2 potential haplotype combinations, it was calculated according to the common haplotype literature (Hsu et al., 2002). Of the 23 common haplotypes in the literature, haplotype 1 and haplotype 2 were type A, and the other 21 haplotypes were type B. The comprehensive classification criteria were that all the haplotypes contained *3DL3*, *2DL4*, and *3DL2*, and one haplotype contained *2DL2* or *2DL3*. One haplotype contained *3DP1* or *3DP1v*. Haplotype B carried one or more of the genes *KIR2DL5*, *KIR2DS1*, *KIR2DS2*, *KIR2DS3*, *KIR2DS5*, and *KIR3DS1*, but haplotype A did not include these genes.

Table 1. Human *KIR* genotype kit primers [locus and size (in bp)].

H	G	F	E	D	C	B	A	
2DL1, 145 bp	2DL2, 146 bp	2DL3, 153 bp	2DL4, 135 bp	2DL5, 145 bp	2DS1, 96 bp	2DS2, 140 bp	2DS3, 158 bp	1
2DS4, 133 bp	2DS5, 110 bp	3DL1, 112 bp	3DL2, 135 bp	3DL3, 196 bp	3DS1, 109 bp	2DP1, 168 bp	3DP1, 131 bp	2

Statistical analysis

According to the literature (Hsu et al., 2002), the *KIR* gene detection frequency (F) was measured by counting: $F (\%) = \frac{\text{the positive gene number}}{\text{the total number in the research group}}$. The *KIR* gene frequency was calculated as follows: $GF = 1 - (1 - f)^{1/2}$; genotype frequency was the genotype positive number n_i/N ; the haplotype frequency was the single haplotype positive number $n_j/2N$, where N is the number of study individuals.

RESULTS

KIR gene frequency distribution

The *3DL3*, *2DL4*, *3DP1*, and *3DL2* genes existed in all individuals from the Xinjiang Han population, yielding a gene frequency of 1 (100%). *2DL1*, *2DP1*, and *2DL3* were common, with gene frequencies of 99.46, 99.46, and 98.91%, respectively; followed by *3DL1* and *2DS4*, with frequencies of 91.30 and 89.13%, respectively. The *2DS2* gene frequency was the lowest at 21.74% (Table 2).

KIR genotype frequencies

There were 21 genotypes detected in the Xinjiang Han population. The AJ (2, 2) genotype was relatively common, with a frequency of 42.39%, followed by AH (5, 2), AE (2, 8), and H (2, 4), with frequencies of 17.39, 11.96, and 8.15%, respectively. The R (6, 9), AA (4, 4), AF (1, 2), and T (8, 8) genotypes showed the lowest frequencies, at

0.54%. Of 21 genotypes, 15 were the same as in the 36 genotypes assigned by Hsu et al. (2002). In addition, six new genotypes were found in 11 individuals who could not be classified according to the related literature (labeled HXJ1-6) and thus for which genotype analysis could not be performed based on existing standards. The frequencies for these were low, 2.72, 1.09, and 0.54%, respectively. In contrast, the J (1, 21), B (3, 6), R (6, 9), and AA (4, 4) genotypes identified in five individuals had not been reported in the Han population in current literature; these exhibited frequencies of 1.09, 0.54, 0.54, and 0.54% (Table 3).

KIR gene haplotype frequencies

Nine haplotypes were detected in 184 individuals from the Han population; haplotypes 1, 2, 3, 4, 5, 7, 8, 17, 21. The most common haplotype, 2 (N = 212), was found in 61.3% (212/346) of individuals, followed by haplotype 5 (N = 44), which accounted for 12.7% (44/346). Haplotype 1 (N = 29) accounted for 8.4% (29/346) and haplotype 6 (N = 23) accounted for 6.7% (23/346). Four haplotypes accounted for 89.0%. Haplotypes 7, 10, 11, 12, 13, 14, 15, 16, 18, 19, 20, 22, and 23 were not detected in this study. Haplotypes 1 and 2 belonged to haplotype group A. Eighty-two cases were haplotype A with a frequency of 69.1%, and the ratio of haplotypes A and B was 2.23:1. In addition, six new genotypes were found, for which haplotype analysis could not be performed based on Hsu's standards (Hsu et al., 2002), suggesting that these may be new haplotypes (Table 4).

KIR gene frequencies in the Xinjiang Han population were comparable with other populations, as shown in Table 5.

Table 2. Distribution of *KIR* gene frequencies in the Xinjiang Han population (N = 184).

KIR gene	N	F (%)	GF
3DL3	184	100.0	1
2DS2	40	21.74	0.1154
2DL2	41	22.28	0.1184
2DL3	182	98.91	0.8956
2DP1	183	99.46	0.9265
2DL1	183	99.46	0.9265
3DP1	184	100.0	1
2DL4	184	100.0	1
3DL1	168	91.30	0.7050
3DS1	81	44.02	0.2518
2DL5	89	48.37	0.2815
2DS3	44	23.91	0.1277
2DS5	59	32.07	0.1758
2DS1	84	45.65	0.2628
2DS4	164	89.13	0.6703
3DL2	184	100.0	1

F = gene detection rate; GF = gene frequency.

Table 3. Observed *KIR* locus profiles and the frequencies of *KIR* genes, genotypes, and haplotypes in the Xinjiang Han population (N = 184).

Genotype*	3DL3	2DS2	2DL2	2DL3	2DPI	2DL1	3DP1	2DL4	3DL1	3DS1	2DL5	2DS3	2DS5	2DS1	2DS4	3DL2	N	Frequency	Haplotype
B	+	+	+	+	+	+	+	+	+	+	+	+	-	+	+	1	0.54	3,6	
C	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	5	2.75	5,3	
H	+	+	+	+	+	+	+	+	+	-	-	-	-	-	-	15	8.15	2,4	
J	+	+	+	+	+	+	+	+	+	+	+	-	+	+	+	2	1.09	1,21	
M	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	3	1.63	2,8	
P	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	4	2.17	2,17	
R	+	+	+	+	+	+	+	+	+	+	+	+	+	-	+	1	0.54	1,2	
AA	+	+	+	+	+	+	+	+	+	-	-	-	-	-	+	1	0.54	4,4	
AE	+	-	-	+	+	+	+	+	+	+	+	-	-	-	+	22	11.96	1,6	
AF	+	-	-	+	+	+	+	+	+	+	+	-	-	-	+	1	0.54	1,2	
AH	+	-	-	+	+	+	+	+	+	+	+	-	+	+	+	32	17.39	5,2	
AI	+	-	-	+	+	+	+	+	+	+	+	-	+	+	+	3	1.63	1,5	
AJ	+	-	-	+	+	+	+	+	+	+	-	-	-	-	+	78	42.39	2,2	
Q	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	4	2.17	9,5	
T	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	1	0.54	8,8	
HXJ1	+	-	-	+	+	+	+	+	+	-	-	-	-	-	-	5	2.72		
HXJ2	+	-	-	+	+	+	+	+	+	+	+	-	-	-	-	1	0.54		
HXJ3	+	+	+	+	+	+	+	+	+	+	+	-	-	-	-	2	1.09		
HXJ4	+	+	+	+	+	+	+	+	+	+	+	-	-	-	-	1	0.54		
HXJ5	+	+	+	+	+	+	+	+	+	+	+	-	-	-	-	1	0.54		
HXJ6	+	+	+	+	+	+	+	+	+	+	+	+	+	+	+	1	0.54		

**KIR* genotype nomenclature is according to Uhrberg et al. (2002), the genotypes as yet unreported are named by HXJ1-6. Eleven newly identified *KIR* genotypes could not be assigned to the haplotypes, according to standard method of Hsu et al. (2002)

Table 4. Haplotype profiles and frequencies in the Xinjiang Han population (N = 173).

Haplotype	3DL3	2DS2	2DL2	2DL3	2DP1	2DL1	3DP1	2DL4	3DL1	3DS1	2DL5	2DS3	2DS5	2DS1	2DS4	3DL2	N	F (%)
A	1	+	-	-	+	+	+	+	+	-	-	-	-	-	-	+	29	8.4
	2	+	-	-	+	+	+	+	+	-	-	-	-	-	+	+	212	61.3
	3	+	+	+	-	-	-	-	+	+	-	-	-	-	-	+	6	1.7
	4	+	+	+	-	-	-	-	+	+	-	-	-	-	+	+	17	4.9
	5	+	-	-	+	+	+	+	-	+	+	-	+	+	-	+	44	12.7
	6	+	-	-	+	+	+	+	-	+	+	+	-	+	-	+	23	6.7
	7	+	-	-	+	+	+	+	+	+	+	-	-	-	-	+	0	0.0
	8	+	+	+	-	+	+	+	+	-	+	+	-	-	-	+	5	1.5
	9	+	+	+	-	+	+	+	-	+	+	+	-	+	-	+	4	1.2
	10	+	+	+	-	-	-	-	+	+	+	-	-	-	-	+	0	0.0
	11	+	-	-	-	+	-	+	-	-	-	-	-	+	-	+	0	0.0
B	12	+	-	-	+	+	+	-	-	+	-	+	-	-	+	+	0	0.0
	13	+	+	+	-	-	-	-	+	+	-	-	+	+	-	+	0	0.0
	14	+	+	+	-	-	-	-	+	+	-	-	+	+	-	+	0	0.0
	15	+	+	+	-	+	+	+	+	-	-	-	-	-	-	+	0	0.0
	16	+	+	+	-	+	+	+	+	-	-	-	-	-	+	+	0	0.0
	17	+	+	+	-	+	+	+	+	-	+	-	+	+	-	+	4	1.2
	18	+	+	+	-	+	+	+	+	-	+	+	-	+	-	+	0	0.0
	19	+	+	+	-	+	+	+	+	-	+	+	-	-	+	+	0	0.0
	20	+	+	+	-	+	+	+	+	-	+	+	-	-	-	+	0	0.0
	21	+	+	+	-	+	+	+	+	-	+	+	-	-	+	+	2	0.6
	22	+	+	+	-	+	+	+	+	-	+	+	-	+	+	+	0	0.0
	23	+	-	-	-	+	-	+	+	-	+	+	-	-	-	+	0	0.0

*Eleven (5.98%) newly identified KIR genotypes in the 184 samples could not be assigned to the haplotypes according to the standard method of Hsu et al. (2002). F = frequency.

Table 5. Distribution of KIR gene frequencies in the Xinjiang Han population and others (%) (Su et al., 2008).

Different ethnic groups	N	3DL3	2DS2	2DL2	2DL3	2DP1	2DL1	3DP1	2DL4	3DL1	3DS1	2DL5	2DS3	2DS5	2DS1	2DS4	3DL2
Xinjiang Han	184	100.0	21.7	22.3	98.9	99.5	99.5	100.0	100.0	91.3	44.0	48.4	23.9	32.1	45.7	89.1	100.0
Jiangsu Han (He et al., 2009)	269	100.0	19.3	17.8	99.3	99.3	96.3	100.0	100.0	96.3	36.8	39.4	13.8	28.3	36.8	95.9	100.0
Sichuan Han (Wang et al., 2010)	135	100.0	28.2	28.2	97.8	99.3	99.3	100.0	100.0	97.0	35.6	43.0	17.8	28.2	37.0	97.0	100.0
Shanghai Han (Zhang et al., 2003b)	87	100.0	18.4	18.4	97.7	97.7	97.7	100.0	100.0	100.0	34.5	37.9	14.9	24.1	35.6	98.8	100.0
Zhejiang Han (Zhu et al., 2005)	104	100.0	17.3	17.3	99.0	99.0	99.0	100.0	100.0	94.2	32.7	35.6	12.5	23.1	33.6	94.2	100.0
Yunnan Han (Su et al., 2008)	150	100.0	25.3	25.3	99.3	99.3	99.3	100.0	100.0	95.3	32.0	36.7	15.3	26.7	36.0	95.3	100.0
Singapore Chinese	46	100.0	28.3	28.2	100.0	100.0	100.0	100.0	100.0	97.8	30.4	39.1	17.4	21.7	28.3	97.8	100.0
Hong Kong, China	100	100.0	28.0	28.0	98.0	99.0	99.0	100.0	100.0	94.0	39.0	45.0	25.0	26.0	40.0	94.0	100.0
Brazil	70	100.0	53.3	52.2	94.4	96.7	96.7	100.0	100.0	95.6	41.1	58.9	38.9	32.2	37.8	95.6	100.0
Brazil	154	100.0	16.9	14.3	99.4	100.0	99.4	100.0	100.0	94.2	36.4	38.3	16.2	26.2	37.7	94.2	100.0
Japan	41	100.0	14.6	14.6	100.0	100.0	100.0	100.0	100.0	97.6	29.3	39.0	14.6	24.4	34.1	97.6	100.0
Oman	99	100.0	49.5	50.5	87.9	98.0	98.0	100.0	100.0	96.0	29.3	59.6	30.3	39.4	32.3	94.9	100.0
Cuba	70	100.0	52.9	52.9	90.0	97.1	97.1	100.0	100.0	91.4	40.0	55.7	35.7	31.4	38.6	91.4	100.0
The Czech Republic	125	100.0	57.0	59.0	86.0	94.0	95.0	100.0	100.0	94.0	38.0	52.0	36.0	26.0	43.0	92.0	100.0
Caucasians*	465	nt	51.8	44.0	92.7	nt	93.8	100.0	100.0	94.0	41.9	52.0	26.5	33.6	40.8	91.2	100.0
Northern Ireland	154	100.0	47.4	47.4	90.3	98.1	98.1	100.0	100.0	98.1	39.0	50.0	29.9	31.2	38.3	98.1	100.0
South Africa Xhosa	50	100.0	64.0	72.0	64.0	98.0	96.0	100.0	100.0	100.0	4.0	82.0	38.0	62.0	10.0	100.0	100.0

*Includes 147 Australians, 90 Irish, 130 Britons, 52 Americans, and another group of 40 Australians (Su et al., 2008).

DISCUSSION

KIR genes are genetically polymorphic, and are located on human chromosome 19q13.4, spanning approximately 150 kb. They are arranged head-to-tail and clustered; their

polymorphisms are encoded by the multi-gene family clusters, with diverse structure and function. The KIR locus contains multiple alleles, which can be expressed by the different KIR genes and vary in number and type between different ethnic groups and different areas. Different individuals have different numbers and types of KIR genes, and therefore show different haplotypes. At present, many laboratories are dedicated to KIR genotyping and haplotype composition studies, and KIR distribution profiles have been reported in more than 140 groups of the world's populations (Hsu et al., 2002; Rajalingam et al., 2002; Uhrberg et al., 2002; Jiang et al., 2005; Zhu et al., 2005; Velickovic et al., 2006; Middleton et al., 2008; Pavlova et al., 2008; Su et al., 2008; He et al., 2009; Dai et al., 2010; Wang et al., 2010).

Our study used the SSP-PCR method to interrogate the KIR gene locus in 184 individuals in the Xinjiang Han population who were the recipients or donors of a kidney transplant. We compared the test results with the other populations reported in the literature and obtained the KIR gene distribution characteristics of the Xinjiang Han population. The detection rates of 3DL3, 2DL4, 3DP1, and 3DL2 were 100% in each group, the rates of 2DL1, 2DP1, 2DL3, and 3DL1 showed a high gene frequency of >90%; these results were consistent with the data reported from the rest of the world populations. 2DS2 and 2DL2 showed lower frequencies, similar to the published data from the Han population in Jiangsu, Shanghai, Zhejiang, and Yunnan. These were, however, significantly higher than those in the South Korean and Japan populations, and significantly lower than those in Sichuan, Hong Kong, Singaporean, Singapore, Brazil, Oman, Cuba, the Czech Republic, Caucasian, Northern Ireland, and South African Xhosa populations. The 3DS1 gene frequency of 44.0% and the 2DS1 gene frequency of 45.7% were higher than in all comparable populations. The 2DS3 gene frequency and the 2DL5 gene frequency were 23.9 and 48.4%, respectively, which were higher than in the Han, Japanese, South Korean, and Caucasian populations, but significantly lower than in the Brazilian, Czech Republic, Oman, Cuban, and South African Xhosa populations. The 2DS5 gene frequency was 32.1% and significantly higher than the Han in Asia, South Korean, Singapore, and Japanese and Caucasian populations, but significantly lower than in the South African Xhosa and Oman populations. The results of this study demonstrated that the Xinjiang Han population has a unique distribution characteristic of KIR genes compared to the Han population in the other areas of China and elsewhere in Asia, as well as with the populations in other countries. This reflected that genetic background plays a large role in determining the KIR polymorphic distributions in different populations in different areas.

There were 21 KIR genotypes detected in the Xinjiang Han population, fewer than the 26 genotypes detected in the Zhejiang Han population (Zhu et al., 2005), the 30 genotypes in the Yunnan Han population (Su et al., 2008), the 34 genotypes in the Jiangsu Han population (Dai et al., 2010), and the 31 genotypes in the Sichuan Han population (Wang et al., 2010), but more than the 18 genotypes in the Shanghai Han population (Zhang et al., 2003b). Among the 21 kinds of KIR genotypes, the AJ (2, 2) genotype was the most common; AF (1, 2), R (6, 9), A (4, 4), AF (1, 2), and T (8, 8) were less common. This profile was different than that of the AJ and AF dominating genotypes found in the Zhejiang, Shanghai, Jiangsu, and Yunnan populations (Zhu et al., 2005; Su et al., 2008; He et al., 2009), while the most common genotype in the Caucasian population was AG (1, 1). In addition, we found 21 genotypes in 184 Han individuals, six (from 11 individuals) of which had not been reported and could not be named in accordance with the relevant literature (Hsu et al., 2002), and were tentatively named HXJ1-6. These six new genotypes were, however, identified in the other populations in China, with IDs of 75, 64, 68, 11, 7, and 6. For the six genotypes, the Yunnan population IDs were

6 and 8; Zhejiang population IDs were 11, 6, 68, 75; the Shanghai population ID was 7; and the Sichuan population ID was the same as us. The ID of 75 was the most common genotype, accounting for 2.72% in five individuals.

KIR haplotypes could be classified into two groups: haplotypes A and B. Haplotype A contained only a single activated receptor gene 2DS4, but haplotype B had more activated receptor genes. Nine haplotypes were detected in 184 Han individuals. The most common haplotype, 2, was found in 61.3% of the subjects, followed by haplotypes 5 and 1, which accounted for 12.7 and 8.4%, respectively. Haplotypes 1 and 2 belonged to haplotype A. Haplotype A was the primary phenotype, with frequencies of 69.1% in the Xinjiang Han population. This was similar to that observed in published data from Han populations in Yunnan, Shanghai, Zhejiang, and Sichuan, with frequencies of 69.4, 74.8, 74.4, and 68.6%. These results showed that genetic background was the major determinant for the haplotype grouping classification, regardless of where individuals from the Han population resided. It has been reported that haplotype 2 was the common haplotype in the Han populations in Yunnan (Su et al., 2008) and Shanghai (Zhang et al., 2003b), but that haplotype 1 was the common haplotype in the Caucasian population. The ratio of haplotype A was higher. It was consistent with Yunnan, Zhejiang, Shanghai, Japanese, and Korean populations. However, the major advantage was haplotype B of Australian aboriginal population, it had a difference with which the ratio of haplotype A and haplotype B of Caucasian population kept relative balance.

In this study, six new genotypes were identified for which haplotype analysis could not be performed based on Hsu's standards, suggesting that they may be new haplotypes. Zhang et al. (2003b) considered that certain special genotypes might have more than one haplotype combination. For instance, genotype A might be the combination of 1 and 10, or might also be 3 and 7; genotype C might be the combination of 5 and 3, or might also be 3 and 7; genotype C might be the combination of 5 and 3, or might also be 1 and 14. The priority in these settings was the common haplotype combination, and then was confirmed by expanding the genealogy or DNA graph.

In conclusion, the genetic polymorphism profile of the KIR locus in the Xinjiang Han population showed common characteristics of the general Han population, as well as some unique characteristics, but showed significant differences from African and Caucasian populations. China has a large population and extensive land, and represents a multi-ethnic country, with long-term coexistence of multiple populations. The KIR gene and haplotype distributions showed unique properties. At present, the theories and studies on this phenomenon are still inadequate. In order to elucidate the nature of the Chinese KIR locus, multi-regional cooperation, large sample sizes, and studies of underlying KIR structure and function should be further pursued. This study provided the basic information for further studies of KIR genes in human genetics, transplantation immunology, and genetic disease.

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