

Karyotype analysis of Rheum palmatum

J.Q. Ye, Y.Y. Jia, K. Fan, X.J. Sun and X.M. Wang

School of Pharmacy, Xi'an Jiaotong University, Xi'an, China

Corresponding author: X.M. Wang E-mail: xumeiwang@hotmail.com

Genet. Mol. Res. 13 (4): 9056-9061 (2014) Received October 14, 2013 Accepted January 29, 2014 Published October 31, 2014 DOI http://dx.doi.org/10.4238/2014.October.31.20

ABSTRACT. Rheum palmatum, one of the source plants of the traditional Chinese medicine rhubarb, is anendemic and endangered species. To our knowledge, this is the first report on the chromosome number and karyotype of this species. Sectioning combined with micrography was used to analyze the karyotype. The following results were obtained: R. palmatum had a stable chromosome number 2n = 22; the basic number of chromosomes was 11; karyotype formula is 2n = 22 = 20 metacentric + 2 submetacentric, belonging to Stebbins' 1A type; and karyotype asymmetry index was 55.39%. The present study showed that R. palmatum has a primitive type of karyotype.

Key words: Rheum palmatum; Chromosome number; Karyotype

INTRODUCTION

Rhubarb is a very important traditional Chinese medicine and is widely used as a purgative and anti-inflammatory agent (Chinese Pharmacopoeia Committee, 2010). Three closely related species [Rheum palmatum L., Rheum tanguticum (Maxim. ex Regel) Maxim. ex Balf., and Rheum officinale Baill.] are considered as important sources of rhubarb; all the plants belong to section Palmata A. Los., and are endemic to China (Bao and Grabovskaya-Borodina, 2003). Because of the excessive exploitation of these plants for rhubarb, the wild varieties of these species have been decreasing annually and are now considered endangered. Our field survey suggested that very few natural populations of these species are found in areas such as Chongqing and Guizhou Provinces. Of the three plant species, R. palmatum shows the widest distribution range in China and is mainly found in Hebei, Shanxi, Shaanxi, Gansu, Sichuan, Qinghai, and Tibet Provinces of China. This species can be found at the forest edges on hills, in shrubs, or in the valleys near rivers.

Our research group has investigated the distribution and genetic variation of the section Palmata (Wang et al., 2010, 2012a,b), and many previous studies on this section have focused on components analysis (Zheng and Zhang, 1993; Zhang and Liu, 2004; Ye et al., 2007) and pharmacological properties (Xiao et al., 1984; Yu et al., 2005; Tseng et al., 2006). However, the chromosome number and karyotype of this section are not yet known. Rheum has been reported to have two ploidy levels, 2n = 22 and 4n = 44 (Darlington and Janaki-Ammal, 1945; Darlington and Wylie, 1955). The chromosome number and karyotype have been reported for R. tanguticum (Hu et al., 2007) and were determined for R. officinale in 1928 by Jaretzky. Chromosome number, size, and karyotype are known to be good taxonomic characters, and cytological data are essential in studies focusing on diversification (Stebbins, 1971). Further, polyploidy has long been recognized as a prominent factor responsible for plant diversification and speciation (Stebbins, 1971; Grant, 1981; Levin, 2002). Our previous studies on the distribution pattern (Wang et al., 2010) and morphological characters (unpublished data) of the three species of rhubarb suggest that they might not be different species and might actually form a species complex (R. palmatum). On the other hand, R. tanguticum was initially published by Regel (1874) as a variety of R. palmatum, and Grabovskaya-Borodina indicated that R. tanguticum was a synonym of R. palmatum (Bao and Grabovskaya-Borodina, 2003). Therefore, the present study aimed to expand the current knowledge on the chromosome number and karyotype variation of the source plants of rhubarb and determine whether the karyological data of R. palmatum and R. tanguticum support the recognition of a single species or suggest that they are in fact two species. The results might provide a basis for the future studies on the species differentiation, speciation, and diversification of the section *Palmata*.

MATERIAL AND METHODS

Material

Seeds of *R. palmatum* were collected from Yala in Kangding County, Sichuan Province, China, on September 2, 2010. Voucher (altitude, 3692m; 30°14.782'N, 101°51.833'E; Yu-qu Zhang 1008090218) specimens are deposited in the Herbarium of Shaanxi Normal University (SANU). Somatic chromosomes were studied using samples from the root meristems of germinating seeds.

Measurement and analysis of karyotype

The cut roots were first pretreated with 0.2% colchicine for 2h at room temperature, and then pretreated with distilled water at 0°C for 24 h; subsequently, they were fixed with Carnoy's fluid (absolute alcohol:glacial acetic acid, 3:1) at 4°C for at least for 30 min and stored in 70% aqueous ethanol until required. The root tips were hydrolyzed with 1 M hydrochloric acid at 60°C for 16 min, and then rinsed in distilled water, stained, and squashed with carbol fuchsin. The chromosome number was determined from fifty well-spread chromosomal cells from five different root tips. Five cells with equivalent degree of chromosome contraction were used for karyotypic analyses. Photographs of the well-spread chromosomes were enlarged, and homologous chromosomes were paired by their similarity in size. Each chromosome was designated according to the position of the centromere (Levan et al., 1964) as follows: m = metacentric (r = 1.00-1.69); sm = submetacentric (r = 1.70-2.99); st = subtelocentric (r = 3.00-6.99); and t = telocentric (r = 7.00 and above).

Karyotypic analyses were performed according to the method described by Li and Chen (1985) and the classification of Stebbins (1971) was used. Karyotype asymmetry was determined as per Arano (1963). The following parameters were calculated: 1) chromosome relative length; 2) arm ratio; 3) chromosome length ratio; 4) centromeric index; 5) relative index of chromosome length (IRL, the ration of chromosome length to the average length of a haploid set of chromosomes); 6) karyotype asymmetry index (AsK%, the ration of the sum of the long arms of a haploid set of chromosomes to the total length of the haploid set of chromosomes x 100%).

RESULTS

Chromosome number

Fifty cells were observed under a fluorescence microscope (Leica DM5000B, German); of the 50 cells, 48 had 22 chromosomes, whereas 2 had 20 chromosomes. The chromosome number, 2n = 22 was represented by 96% of the total cells (Figure 1). The number of somatic chromosomes of *R. palmatum* can be thought to be 2n = 22. No satellite markers and polyploid chromosomes were observed.

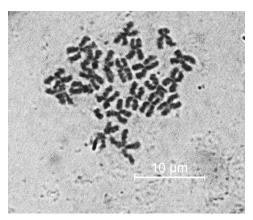


Figure 1. Cytological features of the somatic metaphase spread of Rheum palmatum.

Karyotypes

Somatic chromosome karyotypes were measured in five cells, and the mean karyotype parameters were obtained (Table 1). The individual chromosome length ranged from 1.72 to 2.78 μ m. The total haploid chromatin length (TCL) was 24.41 μ m. The relative chromosome length was between 7.05 and 11.40%. The arm rations ranged from 1.10 to 1.73, with an average of 1.27, and none of the chromosomes had an arm ration of more than 2. The largest chromosome length ratio was 1.62. Karyotypes belonged to the Stebbins 1A category of asymmetry (Stebbins, 1971). The AsK% was 55.39%, and the mean centromeric index was 44.35. The haploid complement of *R. palmatum* consisted of 10 metacentric chromosomes and 1 submetacentric chromosome; therefore, the karyotype formula of *R. palmatum* is 2n = 2x = 22 = 20m + 2sm (Figures 1-3).

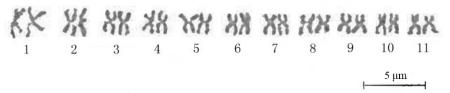


Figure 2. Idiogram of Rheum palmatum chromosomes.

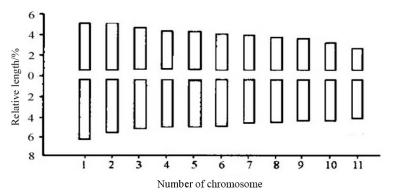


Figure 3. Karyogram of Rheum palmatum chromosomes.

| Chromosome pair | Relative length (%) | Long arm (µm) | Short arm (µm) | Arm ration | Index chromosome length (I.R.L) | Centromeric index (%) | Type |
|-----------------|---------------------|---------------|----------------|------------|------------------------------------|-----------------------|------|
| 1 | 11.40 = 6.23+5.17 | 1.52 | 1.26 | 1.21 | 1.25 | 45.32 | m |
| 2 | 10.87 = 5.70 + 5.17 | 1.39 | 1.26 | 1.10 | 1.19 | 47.55 | m |
| 3 | 9.80 = 5.17 + 4.63 | 1.26 | 1.13 | 1.12 | 1.08 | 47.33 | m |
| 4 | 9.22 = 5.00 + 4.22 | 1.22 | 1.03 | 1.18 | 1.01 | 45.87 | m |
| 5 | 9.18 = 5.00 + 4.18 | 1.22 | 1.02 | 1.20 | 1.01 | 45.55 | m |
| 6 | 9.02 = 4.96 + 4.06 | 1.21 | 0.99 | 1.22 | 0.99 | 45.03 | m |
| 7 | 8.77 = 4.84 + 3.94 | 1.18 | 0.96 | 1.23 | 0.96 | 44.94 | m |
| 8 | 8.57 = 4.80 + 3.77 | 1.17 | 0.92 | 1.27 | 0.94 | 44.06 | m |
| 9 | 8.36 = 4.63 + 3.73 | 1.13 | 0.91 | 1.24 | 0.92 | 45.62 | m |
| 10 | 7.83 = 4.63 + 3.20 | 1.13 | 0.78 | 1.45 | 0.86 | 41.84 | m |
| 11 | 7.05 = 4.47 + 2.58 | 1.09 | 0.63 | 1.73 | 0.78 | 36.60 | sm |
| Total | - | 13.52 | 10.89 | - | - | - | - |
| Mean | - | 1.23 | 0.99 | 1.27 | _ | 44.35 | _ |

J.Q. Ye et al. 9060

DISCUSSION

In general, karyotypes show a transition from symmetry to asymmetry in higher plants (Stebbins, 1971). Primitive or primordial species mostly have symmetric karyotypes, whereas derived, specialized, and advanced plant taxa show asymmetric karyotypes. When the karyotype asymmetry index is close to 50%, the karyotype symmetry is considerably higher, and the karyotype is considered more primitive. The karyotype asymmetry index and karyotype analysis of *R. palmatum* showed an AsK% of 55.39%, which is considerably closer to 50%. Therefore, the karyotype symmetry is high. Further, the karyotypes belonged to Stebbins' 1A category, which also indicates that the karyotypes of *R. palmatum* are symmetric. The karyotype of *R. palmatum* was similar to that of *R. tanguticum* (Hu et al., 2007). Unlike that in *R. officinale*, no polyploids were found in *R. palmatum*. Thus, our findings suggest that *R. palmatum* might be a primitive or primordial species.

Cytological data have been widely used to elucidate the intraspecific and/or interspecific relationships and delimit specific or generic circumscriptions (Stebbins, 1971; Raven, 1975; Hong and Zhang, 1990), and karyotypes are one of the parameters that allow the authentic identification of a specimen. In the present study, besides the similarity in the karyotype, the relative chromosome length, arm ratio, and other parameter values of each chromosome pair were not significantly different (P < 0.05) between R. palmatum and R. tanguticum, confirming that they belong to the same species and have a close relationship. The main morphological characteristics that can be used to distinguish between R. palmatum and R. tanguticum are the degree of leaf blade dissection and shape of the leaf lobes (Bao and Grabovskaya-Borodina, 2003): the leaf blades of R. palmatum are lobed, and the lobed parts are narrowly triangular, whereas the leaf blades of R. tanguticum are dissected, and the lobed parts are narrow, triangular, and lanceolate. However, the transitional morphs between these two species can often be found in the field, and they cannot be easily distinguished morphologically. Further, both the species cannot be discriminated at the chromosomal level. In our previous study on the genetic variation between R. palmatum and R. tanguticum, small molecular variation was detected between the two species (Wang et al., 2012b). Chen et al. (2008) also revealed that R. palmatum and R. tanguticum closely related more than any of other species of rhubarb source. Taken together, these findings indicate that R. palmatum and R. tanguticum might have a common ancestor, and that they might be a single species, i.e., R. palmatum. In other words, karyotypic differentiation has contributed relatively slightly to the interspecific morphological divergence within the source plants of rhubarb. Further chromosomal studies on more populations of the source plants of rhubarb are needed to reveal the diploid differentiation during plant speciation and diversification.

ACKNOWLEDGMENTS

Research supported by the National Natural Science Foundation of China (#31470401).

REFERENCES

Arano H (1963). Cytological studies in subfamily carduoideae (Compositae) of Japan IX. *Bot. Mag.* 76: 32-39. Bao BJ and Grabovskaya-Borodina AE (2003). *Rheum*. In: Flora of China (Li AR and Bao BJ, eds.). Science Press, Beijing and Missouri Botanical Garden, St. Louis 5: 341-350.

Chen DX, Li LY, Zhong GY, Qin SY, et al. (2008). Study on genetic relationship of official Rheum by SRAP. Zhongguo

Zhong Yao Za Zhi 33: 2309-2312.

Chinese Pharmacopoeia Committee (2010). Pharmacopoeia of the People's Republic of China. China Medical Science and Technology Press, Beijing.

Darlington CD and Janaki-Ammal EK (1945). Chromosome Atlas of Cultivated Plants. Allen and Unwin, London.

Darlington CD and Wylie AP (1955). Chromosome Atlas of Flowering Plants. 2nd edn. Allen and Unwin, London.

Grant V (1981). Plant Speciation. 2nd edn. Columbia University Press, New York.

Hong DY and Zhang SZ (1990). Observations on chromosomes of some plants from western Sichuan. Cathaya 2: 191-197.

Hu YP, Xie XL, Feng HS, Zhao XD, et al. (2007). Karyotype analysis of Rheum tanguticum chromosome. Chin. Pharmaceut. J. 42: 42.-258.

Levan A, Fredga K and Sandberg AA (1964). Nomenclature for centromeric position on chromosomes. *Hereditas* 52: 201-220.

Levin DA (2002). The Role of Chromosomal Change in Plant Evolution. Oxford University Press, Oxford.

Li MX and Chen RY (1985). A suggestion on the standardization of karyotype analysis in plants. *J. Wuhan Bot. Res.* 3: 297-302

Raven PH (1975). The bases of angiosperm phylogeny: cytology. Ann. Mo. Bot. Gard. 62: 724-764.

Regel (1874). Rheum palmatum L. var. tanguticum Maxim. Gartenflora 23: 305-306.

Stebbins GL (1971). Chromosomal Evolution in Higher Plants. Edward Arnold, London.

Tseng SH, Lee HH, Chen LG, Wu CH, et al. (2006). Effects of three purgative decoctions on inflammatory mediators. *J. Ethnopharmacol.* 105: 118-124.

Wang XM, Hou XQ, Zhang YQ and Li Y (2010). Distribution pattern of genuine species of rhubarb as traditional Chinese medicine. *J. Med. Plants Res.* 4: 1865-1876.

Wang XM, Hou XQ, Zhang YQ, Yang R, et al. (2012a). Genetic diversity of the endemic and medicinally important plant *Rheum officinale* as revealed by Inter-Simpe Sequence Repeat (ISSR) markers. *Int. J. Mol. Sci.* 13: 3900-3915.

Wang XM, Yang R, Feng SF, Hou XQ, et al. (2012b). Genetic variation in *Rheum palmatum* and *Rheum tanguticum* (Polygonaceae), two medicinally and endemic species in China using ISSR markers. *PLoS One* 7: e51667.

Xiao PG, He LY and Wang LW (1984). Ethnopharmacologic study of Chinese rhubarb. J. Ethnopharmacol. 10: 275-293.

Ye M, Han J, Chen H, Zheng J, et al. (2007). Analysis of phenolic compounds in rhubarbs using liquid chromatography coupled with electrospray ionization mass spectrometry. *J. Am. Soc. Mass Spectrom.* 18: 82-91.

Yu M, Luo YL, Zheng JW, Ding YH, et al. (2005). Effects of rhubarb on isolated gastric muscle strips of guinea pigs. World J. Gastroenterol. 11: 2670-2673.

Zhang HX and Liu MC (2004). Separation procedures for the pharmacologically active components of rhubarb. J. Chromatogr. B Analyt. Technol. Biomed. Life Sci. 812: 175-181.

Zheng JH and Zhang ZG (1993). Retrospect and prospect of studies on Rheum L. J. Beijing Med. Univ. 25 (Suppl): 1-2.