



Chromosome number variation and evolution in Neotropical Leguminosae (Mimosoideae) from northeastern Brazil

E.C.X.R. Santos¹, R. Carvalho², E.M. Almeida¹ and L.P. Felix¹

¹Programa de Pós-Graduação em Agronomia,
Departamento de Ciências Biológicas, Universidade Federal da Paraíba,
Areia, PB, Brasil

²Departamento de Biologia, Universidade Federal Rural de Pernambuco,
Recife, PE, Brasil

Corresponding author: L.P. Felix
E-mail: lpfelix@hotmail.com

Genet. Mol. Res. 11 (3): 2451-2475 (2012)

Received September 19, 2011

Accepted May 17, 2012

Published June 27, 2012

DOI <http://dx.doi.org/10.4238/2012.June.27.1>

ABSTRACT. Most members of the subfamily Mimosoideae have pantropical distributions, variable habits, and a basic chromosome number $x = 13$. We examined karyotypic evolution of 27 species of this subfamily occurring principally in northeastern Brazil by examining chromosomes stained with Giemsa. All of the species had semi-reticulated interphase nuclei and early condensing segments in the proximal region of both chromosome arms. The basic number $x = 13$ was the most frequent, with $2n = 2x = 26$ in 19 of the species, followed by $2n = 4x = 52$ and $2n = 6x = 78$. However, the three species of the genus *Calliandra* had the basic number $x = 8$, with $2n = 2x = 16$, while *Mimosa cordistipula* had $2n = 4x = 32$. The karyotypes were relatively symmetrical, although bimodality was accentuated in some species, some with one or two acrocentric pairs. As a whole, our data support earlier hypotheses that the Mimosoideae subfamily has a basic number of $x = 13$ and underwent karyotypic evolution by polyploidy. However, $x = 13$ seems to be a secondary basic number that originated from an

ancestral stock with $x_1 = 7$, in which polyploidy followed by descending dispolyploidy gave rise to the current lineages with $x = 13$. Another lineage, including current representatives of *Calliandra* with $x = 8$, may have arisen by ascending dispolyploidy directly from an ancestral monoploid stock with $x_1 = 7$.

Key words: Mimosoideae; *Calliandra*; Karyotypic evolution; Polyploidy

INTRODUCTION

The subfamily Mimosoideae comprises approximately 40 genera and 2500 species distributed throughout tropical, subtropical, and sub-temperate regions (Judd, 2009). The subfamily is divided into 3 tribes: Mimoseae (which includes the former tribes Parkieae and Mimozygantheae), Acacieae, and Ingeae (Luckow, 2005). Some genera of the subfamily have controversial delimitations, such as *Acacia sl* (one of the largest genera in the subfamily), which is considered polyphyletic; most of the species of northeastern Brazil are included in the genus *Senegalia* (Queiroz, 2009). The subfamily has significant economic importance: many representatives of the genera *Acacia*, *Albizia*, *Stryphnodendron*, *Piptadenia*, *Enterolobium*, *Inga*, *Entada*, *Parkia*, and *Prosopis* are used as ornamental or forage plants, sources of firewood, or for extracting secondary metabolic compounds.

The subfamily Mimosoideae has only been poorly studied in cytological terms, with chromosome numbers reported for approximately 460 species, corresponding to almost 17% of the total number of species in the subfamily. These chromosome numbers vary from $2n = 16$ in some species to $2n = 208$ in *Acacia hebeclada*. The most frequent basic number for the group is $x = 13$, with polyploidy occurring widely (Atchison, 1951; Dahmer et al., 2011).

Although the subfamily is well-represented in Brazil, only a limited number of chromosomal records are available for this group; the data are even more fragmented for the northeastern region of the country. Only 13 species of the genus *Inga* were examined by Mata (2009, unpublished data), and all of them had the basic number $x = 13$, with the evolution of their chromosome numbers being regulated by various cycles of polyploidy.

The subfamily Mimosoideae is particularly well-represented in northeastern Brazil, with a total of 24 genera and 267 species, most of which occur in the “Caatinga” (dryland) Biome (Queiroz, 2009). Despite the fact that this subfamily has a karyotypic evolution pattern that is apparently preferentially regulated by polyploidy, the occurrence of $x = 8$ in the genus *Calliandra* suggests that a dispolyploidy variation occurred at the very start of the diversification of this group, at least in this lineage (Atchison, 1951).

Certain patterns of karyotypic evolution in plants are influenced by environmental factors. Higher polyploidy frequencies, for example, have been observed among plants growing in the arctic tundra (Brochmann et al., 2004), and among orchids growing on inselbergs in Brazil (Felix and Guerra, 2000). However, this relationship has not been observed in the family Leguminosae or in plants growing in the “Caatinga”.

The present study investigated the patterns of karyotypic evolution in different genera of the subfamily Mimosoideae by analyzing the numbers and morphologies of the chromosomes of 27 species belonging to 11 genera of the 3 tribes of Mimosoideae from different Brazilian biomes. Emphasis was placed on the species from the northeastern region of that

country, principally those from the “Caatinga” region and from altitudinal forests (“Brejos de Altitude”), and the chromosomal variations seen in these 2 environments were comparatively evaluated.

MATERIAL AND METHODS

Table 1 presents a list of all the species analyzed, their respective chromosome numbers, previous counts, collection sites, and reference specimens. We used root tips from seeds germinated in Petri dishes or excised from plants cultivated in the experimental gardens of the Plant Cytogenetics Laboratory at the Centro de Ciências Agrárias of the Federal University of Paraíba State. Reference specimens of all analyzed materials were deposited in the Jayme Coelho de Moraes Herbarium (EAN).

Table 1. Species analyzed with respective provenances, biomes, voucher numbers, chromosome numbers, previous counts, and sources.

Taxa	Provenances and biome	Voucher No.	2n	Previous counts	Sources*
Tribe Acacieae					
<i>Senegalia bahiensis</i> (Benth.) Seigler & Ebinger	Sertânea, PE (CA)	J.P. Castro, 206	26	-	-
<i>S. riparia</i> (Kunth.) Britton & Rose	Serraria, PB (BE)	E.C.X.R. Santos, 12	26	-	-
<i>S. tenuifolia</i> (L.) Britton & Rose	Pilões, PB (BE)	E.C.X.R. Santos, 7	26	26	A48
	Pilões, PB (BE)	E.C.X.R. Santos, 10	26	-	-
<i>Vachellia farnesiana</i> (L.) Wight & Arn.	Toritama, PE (CA)	L.P. Felix, S/N	52	52	A48
Tribe Ingeae					
<i>Abarema filamentosa</i> (Beth.) Pittier.	Morro do Chapéu, BA (CA)	J.P. Castro, 118	26	-	-
<i>Calliandra depauperata</i> Benth	Santa Maria da Boa Vista, PE (CA)	J.P. Castro, 24	16	-	-
	Morro do Chapéu, BA (CA)	J.P. Castro, 194	26	-	-
<i>C. leptopoda</i> Benth.	Irecê, BA (CA)	J.P. Castro, 100	16	-	-
<i>C. portoricensis</i> (Jacq.) Benth.	Areia, PB (BE)	E.C.X.R. Santos, 16	16	-	-
<i>Chloroleucon</i> sp	Serraria, PB (BE)	E.C.X.R. Santos, 13	52	-	-
<i>Enterolobium contortisiliquum</i> (Vell.) Moring.	Juarez Távora, PB (CA)	L.P. Felix, S/N	26	26	B74
<i>Inga cinnamomea</i> Benth.	Recife, PE (MA)	L.P. Felix, S/N	26	-	-
Tribe Ingeae					
<i>Inga cinnamomea</i> Benth.	Manaus, AM (AM*)	L.P. Felix, S/N	26	-	-
<i>I. edulis</i> Mart.	Manaus, AM (AM*)	L.P. Felix, S/N	26	26	M09
Tribe Mimoseae					
<i>Anadenanthera colubrina</i> (Vell.) Brenan	Caruaru, PE (CA)	L.P. Felix, 13. 146	26	26	O10
<i>Desmanthus pernambucanus</i> (L.) Thell.	Areia, PB (BE)	E.C.X.R. Santos, 15	26	-	-
	Senhor do Bom Fim, BA (CA)	J.P. Castro, 88	26	-	-
<i>Mimosa arenosa</i> (Willd.) Poir.	Sertânea, PE (CA)	J.P. Castro, 205	26	-	-
	Areia, PB (BE)	E.C.X.R. Santos, 6	26	-	-
<i>M. campicola</i> Harms	Cuitegi, PB (CA)	L.P. Felix, 13. 112	78	26	DN11
	Serraria, PB (BA)	E.C.X.R. Santos, 11	78	-	-
<i>M. cordistipula</i> Benth.	Irecê, BA (CA)	J.P. Castro, 97	32	26	DN11
<i>M. paraibana</i> Barneby	Lagoa Seca, PB (CA)	L.P. Felix, S/N	26	-	-
<i>M. pseudosepiaria</i> Harms	Salgueiro, PE (CA)	J.P. Castro, 7	26	-	-
<i>M. quadrivalvis</i> var. <i>leptocarpa</i> (DC.) Barneby	Areia, PB (BE)	E.C.X.R. Santos, 5	52	-	-
<i>M. sensitiva</i> L.	Serraria, PB (BE)	E.C.X.R. Santos, 9	26	26	DN11
<i>Mimosa setosa</i> var. <i>paludosa</i> (Benth.) Barneby	Paulista, PE (MA)	L.P. Felix, 13.183	26	26, 52	DN11
<i>M. somnians</i> Humb & Bamp. ex Willd.	Areia, PB (BE)	E.M. Almeida, 96	52	26	DN11
<i>Mimosa somnians</i> Humb & Bamp. ex Willd.	Areia, PB (BE)	E.C.X.R. Santos, 3	52	26	-
<i>M. tenuiflora</i> (Willd.) Poir.	Areia, PB (BE)	E.C.X.R. Santos, 2	26	26	DN11
<i>M. ulbrichiana</i> Harms	Areia, PB (BE)	L.P. Felix, 13.134	26	-	-
<i>Parkia pendula</i> (Willd.) Walp.	Goiana, PE (MA)	L.P. Felix, S/N	26	22	BK07
<i>Piptadenia viridiflora</i> (Kunth.) Benth.	Arara, PB (CA)	E.C.X.R. Santos, 14	26	-	-

Sources: AT48 = Atchison, 1948; B74 = Bandel, 1974; BK07 = Barella and Karsburg, 2007; DN11 = Dahmer et al., 2011; M09 = Mata, 2009; O10 = Ortolani et al., 2010. Acronyms for the Brazilian States: AM = Amazonas; BA = Bahia; PB = Paraíba; PE = Pernambuco. Biomes: AM* = Amazônia; BE = Brejo de altitude; CA = Caatinga; MA = Mata Atlântica.

We undertook an initial survey of the chromosome numbers compiled in established chromosome number indices, in addition to referrals to the original publications, in order to gather basic information about the chromosome number variability within the subfamily as a whole. These data were then organized into a general table of chromosome numbers (Table 2), which was used to elaborate another list demonstrating the haploid chromosome numbers of each genus in decreasing order of their frequency (Table 3) and indicating the most probable basic chromosome numbers. These basic numbers generally corresponded to the most frequent haploid number of the genus or to those that most parsimoniously explained numerical relationships with related genera (Guerra, 2000). Thus, the taxa were organized into tribes according to the taxonomic treatment adopted by Queiroz (2009) (Table 2).

For karyological analyses, the root tips were pretreated with 2 mM 8-hydroxyquinoline for 20 h, fixed in 3:1 Carnoy fixative (absolute ethanol/glacial acetic acid) for 3 to 24 h at room temperature, and stored at -20°C in the fixing agent for subsequent analysis. To prepare the slides, the root tips were hydrolyzed in 5 N HCl for 20 min at room temperature, squashed in 45% acetic acid, frozen in liquid nitrogen (to remove the coverslip), air dried, and stained conventionally with Giemsa (Guerra, 1983). The material was photographed using an Olympus BX41 microscope fitted with an Olympus D-54 digital camera.

RESULTS

All the species analyzed had semi-reticulated interphase nuclei, with condensed chromatin-forming chromocenters with irregular outlines (Figure 1A). Early condensing segments were observed in the proximal regions of the chromosome arms during prophase (Figures 1J, 2F). The chromosome numbers varied from $2n = 16$ in *Calliandra depauperata*, *Calliandra leptopoda*, and *Calliandra portoricensis* to $2n = 78$ in *Mimosa campicola*, while chromosome sizes varied from 0.70 μm in *M. campicola* to 2.57 μm in *Mimosa quadrivalvis* var. *leptocarpa* (Figures 2D,H). The karyotypes were relatively symmetrical, with a predominance of meta- to submetacentric chromosomes; satellites were observed in *Mimosa paraibana* (Figure 2F), *M. quadrivalvis* (Figure 2H), and *Mimosa tenuiflora* (Figure 2L). In at least 2 species, *Piptadenia viridiflora* (Figure 2P) and *Parkia pendula* (Figure 2O), 2 pairs of acrocentric chromosomes lent an accentuated bimodality to the karyotypes. A synthesis of the principal karyotypic characteristics of the species is presented below.

In the tribe Acacieae, *Senegalia bahiensis* (Figure 1B), *Senegalia riparia* (Figure 1C), and *Senegalia tenuifolia* (Figure 1D) had $2n = 26$, while *Vachellia farnesiana* had $2n = 52$ (Figure 1E). The chromosomes were small, with the smallest chromosomes of *V. farnesiana* measuring 0.92 μm , and the largest chromosomes of *S. riparia* measuring 2.36 μm .

In the tribe Ingeae, *Abarema filamentosa* (Figure 1F) and a population of *C. depauperata* from Morro do Chapéu (Figure 1H) were observed to have $2n = 26$ and chromosomes measuring from 1.02 to 1.90 μm . The 2 other species of *Calliandra* (*C. leptopoda* (Figure 1I) and *C. portoricensis*) (Figure 1J), as well as a population of *C. depauperata* (Figure 1H) from Santa Maria da Boa Vista, had karyotypes of $2n = 16$. In the genus *Inga*, *I. edulis* (Figure 1O) and *I. cinnamomea* showed $2n = 26$ (Figure 1N) (the latter species being represented by 2 different populations from the states of Pernambuco and Amazonas). The number $2n = 26$ was also observed in *Enterolobium contortisiliquum* (Figure 1M). On the other hand, the only population of *Chloroleucon* sp (from the municipality of Serraria, Paraíba State) was tetraploid, with $2n = 52$, and had small chromosomes measuring from 0.78 to 1.14 μm (Figure 1L).

Table 2. Chromosome number in subfamily Mimosoideae according to index chromosome numbers and other sources.

Taxa	n	2n	*Source
<i>Acacia</i> Mill.			
<i>A. abyssinica</i> ssp <i>calophylla</i> Brenan		52	G81
<i>A. acinacea</i> Lindl.		26	G81
<i>A. acuminata</i> Benth.		26	G81
<i>A. adinophylla</i> Maslin		26	GJ96
<i>A. arabica</i> Willd.		44, 52, 104	F69
<i>A. armata</i> Heyne	52	26, 28	F69
<i>A. aroma</i> Gillies ex Hook. & Arn.		26, 52	F69, GJ03
<i>A. asak</i> (Forssk.) Willd.			GJ03
<i>A. ataxacantha</i> DC.		52, 104	F69
<i>A. atramentaria</i> Benth.			GJ03
<i>A. auriculiformis</i> A. Cunn. ex Benth.		26	F69, G81, GJ00, GJ03, GJ06
<i>A. baileyana</i> F. Muell.		26	F69, G81, GJ91
<i>A. benthamii</i> Meissn		52	G81, G84
<i>A. berlandieri</i> Benth.		26	F69
<i>A. bivenosa</i> DC.		26	GJ06
<i>A. bonariensis</i> Gill.	26	26, 52	F69
<i>A. botrycephala</i> (Vent.) Desf.		26	G81
<i>A. brachybotrya</i> Benth.		26	G81
<i>A. brachystachya</i> Benth.		52	G81
<i>A. brevispica</i> Harms		26	G81
<i>A. caesia</i> Wight & Arn.		26	G81
<i>A. caffra</i> (Thunb.) Willd.	13	26	G81, GJ03
<i>A. calamiifolia</i> Benth.		26	F69
<i>A. cardiophylla</i> Cunn. ex Benth.		26	G81
<i>A. catacha</i> Willd.		26	G81
<i>A. catechu</i> (L. f.) Brandis		26	G03
<i>A. catechu</i> (L.) Willd.	13	26	F69, M77, G84, GJ91, GJ06
<i>A. caven</i> (Molina) Molina (as <i>A. cavenia</i>)	26	26, 52	F69, GJ03
<i>A. chortophylla</i> Benth.		26	F69
<i>A. chundra</i> (Roxb.) Willd.	13	26	G81
<i>A. concinna</i> DC.		26	F69
<i>A. confusa</i> Merr.	13	26	F69, M73, G84, GJ91
<i>A. constricta</i> Benth.		52	F69
<i>A. cultriformis</i> A. Cunn.		26	F69
<i>A. curviflucta</i> Burkart		26	F69
<i>A. cyanophylla</i> Lindl.		26	F69
<i>A. dealbata</i> Link var. <i>dealbata</i>	13	26	F69, G81, GJ91, GJ94, GJ03, GJ06
<i>A. decora</i> Rehb.		26	G81
<i>A. decurrens</i> Willd.	13	26	F69, GJ91, GJ94, GJ03, GJ06
<i>A. dermatophylla</i> F. Muell.		26	F69

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Acacia detinens</i> Burch.		26	F69
<i>A. dipiera</i> Lindl.		26	G81
<i>A. dodonaeifolia</i> (Pers.) Willd.		26	G81
<i>A. drepanolobium</i> Harms ex Sjoestedt		52	GJ03
<i>A. drummondii</i> Lindl.		26	G81
<i>A. eburnea</i> Willd.	52	52, 104	F69
<i>A. elata</i> Cunn. ex Benth.		26	G81
<i>A. elatior</i> Benth.		52	GJ03
<i>A. elongata</i> Sieb. ex DC.		26	G81
<i>A. falcata</i> Willd.		26	F69
<i>A. falciiformis</i> DC.		26	G81
<i>A. farinosa</i> Lindl.		26	G81
<i>A. farnesiana</i> (L.) Willd.		26, 52, 104	F69, GJ91, GJ03, GJ06
<i>A. furcatispina</i> Burkart	13, 52	26	GJ03
<i>A. furcatispina</i> Burkart (as <i>A. furcata</i>)		26	F69
<i>A. galpinii</i> Burt Davy		40	G81
<i>A. gerrardii</i> Benth.		56	GJ06
<i>A. giraffae</i> Burch.		52	G81
<i>A. glandulifera</i> S. Watson		26	G84
<i>A. glandulicarpa</i> Reeder		26	G81
<i>A. glaucoptera</i> Benth.		26	F69
<i>A. gracilifolia</i> Maiden		26	G81
<i>A. graveolens</i> A. Cunn.		26	F69
<i>A. greggii</i> Benth.		26	GJ03, GJ06
<i>A. guachapele</i> (Kunth) Dugand		26	GJ96
<i>A. harpophylla</i> F. Muell.		26	F69
<i>A. hebeclada</i> DC.		208	G81
<i>A. heterophylla</i> (Lam.) Willd.		52	G81
<i>A. hockii</i> De Wild.		26, 52	G81, G85
<i>A. holosericea</i> Cunn. ex G. Don		52	GJ03
<i>A. horrida</i> Willd.	52	52, 104	F69
<i>A. hova</i> Drake		26	F69
<i>A. implexa</i> Benth.		26	G81, GJ03
<i>A. intsia</i> Willd.	13	26	G81
<i>A. juniperina</i> DC.		26	F69
<i>A. karroo</i> Hayne		52	G81
<i>A. kirikii</i> Oliv. ssp Kirikii		52	G81
<i>A. koa</i> A. Gray		26, 52	F69
<i>A. laeta</i> R. Br.	26	52	F69
<i>A. lenticularis</i> Benth.		26	G84
<i>A. leucocalyx</i> (Britton & Rose) L. Rico		26	GJ96

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Acacia leucophloea</i> (Roxb.) Willd.		52	F69, G81, G106
<i>A. ligulata</i> A. Cunn. ex Benth.	13	26	G106
<i>A. linifolia</i> (Vent.) Willd.		26	G81
<i>A. longifolia</i> (Andrews) Willd.		26	F69, G81
<i>A. macracantha</i> Willd.		26	F69
<i>A. mangium</i> Willd.		26	G100
<i>A. mearnsii</i> De Willd.		26, 52	G103, G106
<i>A. melanoxylon</i> R. Br.		26	F69, G81, G103, G106
<i>A. mellijera</i> (Vahl) Benth.		26	F69, G103
<i>A. modesta</i> Wall.	13	26	F69, G84, G191
<i>A. mollissima</i> Willd.		26, 52	F69, G103, G106
<i>A. montiformis</i> Griseb.		26	F69
<i>A. nigrescens</i> Oliv.		26	G81
<i>A. nilotica</i> (L.) Willd. ex Delile		26	F69, G84, G191, G106
<i>A. nilotica</i> var. <i>adstringens</i> (Schumacher & Thonn.) Chiov.	12, 13, 26, 52	26, 52, 104	F69, G84, G191, G106
<i>A. nilotica</i> var. <i>indica</i> (Benth.) A.F. Hill		52, 104	G103
<i>A. nilotica</i> var. <i>tomentosa</i> (Benth.) A.F. Hill		52	G103
<i>A. notabilis</i> F.V. Muell.		26	G81
<i>A. nubica</i> Benth.		26	G103
<i>A. occidentalis</i> Brandegee		56	G103
<i>A. origina</i> A. Hilde		26	G196
<i>A. pallens</i> Rolfe		26	G191
<i>A. parramattensis</i> Tindale		26	F69
<i>A. pennata</i> Willd.		26	F69, M73, G81
<i>A. penninervis</i> Sieber		26	F69
<i>A. pervillei</i> Benth.		26	G196
<i>A. plurijuga</i> (Standl.) Britton & Rose		26	G196
<i>A. podalyrifolia</i> A. Cunn.		26	F69
<i>A. polyacantha</i> subsp. <i>campylacantha</i> (Hochst. ex A. Rich.) Brenan		26, 52	G81, G103
<i>A. pulchella</i> R. Br.		26	G81
<i>A. purpusii</i> Britton & Rose		26	G196
<i>A. raddiana</i> Savi		78, 104	F69, G106
<i>A. rehmanniana</i> Schinz		52	F69, G81
<i>A. retinodes</i> A. Cunn.		26	F69
<i>A. richii</i> A. Gray		26	F69
<i>A. rigidula</i> Benth.		26	F69
<i>A. rubida</i> A. Cunn.		26	F69
<i>A. robusta</i> Burch.		26, 52	G81, G84, F69
<i>A. rotundifolia</i> Hook.		26	G81

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Acacia salicina</i> Lindl.		26	G106
<i>A. saligna</i> Wendl.		26	F69
<i>A. schaffneri</i> (Wats.) Hermann		26	F69
<i>A. scorpioides</i> Wight	52	52, 104, 208	F69
<i>A. senegal</i> (L.) Willd.	13	26	F69, G84, G103, G106
<i>A. senegal</i> x <i>A. mellifera</i> (M. Vaht) Benth.		39	F69
<i>A. seyal</i> Delile		52	F69
<i>A. seyal</i> Delile var. <i>seyal</i>		104	G103
<i>A. seyal</i> var. <i>fastata</i> (Schweinf.) Oliv.		104	G103
<i>A. sieberiana</i> A. Chev.		52	F69
<i>A. sieberiana</i> DC.		26	G103
<i>A. sieberiana</i> DC. var. <i>woodii</i> (Burr) Davy) Keay & Brennan		104	G81
<i>A. sophorae</i> R. Br.		26	G03
<i>A. sovaleni</i> Maiden		38	G181
<i>A. spirocarpa</i> Hochst.		52	F69
<i>A. spectabilis</i> A. Cunn. ex Benth.		26	G81
<i>A. spinescens</i> Benth.		26	G81
<i>A. stenophylla</i> A. Cunn		26	G81
<i>A. stricta</i> (Andr.) Willd.		26	G81
<i>A. suaveolens</i> (Smith) Willd.		26	G81, G191
<i>A. suberosa</i> A. Cunn. ex Benth.		26	G106
<i>A. suberosa</i> F.V.M.		26	G81
<i>A. suma</i> Buch - Ham. ex Wall.		26	G103, G106
<i>A. suma</i> (Roxb.) Kurz		26	F69
<i>A. sundra</i> (Roxb.) Delile		26	F69
<i>A. tarrensiana</i> (L.) Willd.	26	52	G84
<i>A. tenuifolia</i> Willd.		26	F69
<i>A. tetragonophylla</i> F.V.M.		26	G81
<i>A. texensis</i> Torr et A. Gray		26	F69
<i>A. tomentosa</i> (Michelx) Standl.		26	G196
<i>A. tortilis</i> (Forsk.) Hayne		52, 78, 104	G103, G106
<i>A. tortilis</i> (Forsk.) Hayne ssp. <i>heteracantha</i> (Burch) Brennan		52	G81
<i>A. tortilis</i> (Forsk.) Hayne ssp. <i>raddiana</i> (Savi) Brennan		52, 78, 104	G81, G103
<i>A. tortilis</i> subsp. <i>spirocarpa</i> (Hochst. ex A. Riech.) Brennan		104	G103
<i>A. tortuosa</i> (L.) Willd.		26	F69
<i>A. triptera</i> Benth.		26	G81
<i>A. undulifolia</i> A. Cunn.		26	G81
<i>A. uruphylla</i> Benth.		26	G81

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Acacia verec</i> Guill. & Perr.		26	GJ06
<i>A. vernicosa</i> Standl.		26	F69
<i>A. verticillata</i> Willd.		26, 28	F69
<i>A. vestida</i> Ker.		26	G81
<i>A. victoriae</i> Benth.		26	G81, GJ06
<i>A. villosa</i> (Swartz) Willd.		26	F69
<i>A. visco</i> Lorentz ex Griseb.		26	F69, GJ03
<i>A. xanthophloea</i> Benth.		52	F69
<i>A. xylocarpa</i> A. Cunn.		26	F69
<i>A. zanzibarica</i> (S. Moore) Taub.		52	GJ03
<i>Abarema</i> Pittier			
<i>A. filamentosa</i> (Benth.) Pittier.		26	PS
<i>Adenanthera</i> L.			
<i>A. pavonina</i> L.		24, 26, 28	M73, M74, M77, G81, GJ91
<i>A. microsperma</i> Teijsm. & Binn.	13	24	F69
<i>Albizia</i> Durazz.			
<i>A. adinocephala</i> (Donn. Sm.) Britton & Rose ex Rec.		26	GJ96
<i>A. amara</i> (Roxb.) B. Boivin	13	26	G81, G84
<i>A. chinensis</i> (Osbeck) Merr.	13	26	M73, M74, M77, G81, GJ03
<i>A. concinna</i> Mundk. & Thirum		c. 78	GJ91
<i>A. coreana</i> Nakai		26	M73
<i>A. dinklagei</i> (Harms.) Keyz (ex. <i>Samanea</i>)		26, 56	F69
<i>A. distachya</i> (Vent.) Macbr.		26	F69
<i>A. falcata</i> Baeker ex Merr.		26	GJ91
<i>A. ferruginea</i> (Guill. & Perr.) Benth.		26	F69, GJ91
<i>A. gamblei</i> Prain		26	F69
<i>A. guachapele</i> (Kunth) Dugand		26	GJ96
<i>A. juitibrissin</i> (Willd.) Durass.	13	26, 52	F69, M77, G81, GJ91, GJ06
<i>A. lebbekoides</i> Benth.	13	26	F69, M74, M77, GJ81, G84
<i>A. lebbekoides</i> (DC.) Benth.		26	F69
<i>A. leucocalyx</i> (Britton & Rose) L. Rico		26	GJ96
<i>A. lophantha</i> Benth.	13	26	F69, G84
<i>A. lucida</i> (Roxb.) Benth.	13	26	G84
<i>A. mollis</i> Boiv.	13	26	G84
<i>A. neumannia</i> Heyne		26	F69
<i>A. niopoides</i> (Spruce ex Benth.) Burkart		26	GJ96
<i>A. occidentalis</i> Brandegee		26	GJ96

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Acacia odoratissima</i> (L. f.) Benth.	13	26	F69, M74, M77, G81 e G84
<i>A. plurijuga</i> (Standl.) Britton & Rose		26	GJ96
<i>A. polyphylla</i> Fourn.		104	F69
<i>A. procera</i> (Roxb.) Benth.	13	26	F69, M74, M77, G81, G84, G103
<i>A. purpusii</i> Britton & Rose		26	GJ96
<i>A. saman</i> (Jacq.) Merr. (ex. <i>Samanea</i>)		26	F69, GJ90, GJ96, G103
<i>A. sassa</i> Maobr.		26	F69
<i>A. stipulata</i> Boiv.	13	26	G81, G84
<i>A. tomentosa</i> (Michelx) Standl.		26	GJ96
<i>A. zygia</i> (DC.) Maobr.		26	F69
<i>Anadenanthera</i> Spreng.			
<i>A. colubrina</i> (Vell.) Brenan		26	G85, O10, PS
<i>A. colubrina</i> var. <i>cebil</i> (Griseb.) Alschul		26	G100
<i>A. macrocarpa</i> (Benth.) Brenan		24, 26	G100
<i>Calliandra</i> Benth.			
<i>C. confusa</i> Sprague & Riley		16, 22	G84
<i>C. depauperata</i> Benth.		16, 26	PS
<i>C. eriophylla</i> Benth.		16	GJ90
<i>C. haematocephala</i> Retz.	8	16	G84
<i>C. hematocephala</i> Hassk.		16	F69
<i>C. houstoniana</i> Standley		16	G84
<i>C. inaequilatera</i> Rusby	8	16	F69
<i>C. leptopoda</i> Benth.		16	PS
<i>C. magdalenae</i> (Bert.) Benth.		16	G84
<i>C. physocalyx</i> H. Hern. & M. Sousa		22	GJ91
<i>C. portoricensis</i> (Jacq.) Benth.		16	GJ90, PS
<i>C. pitieri</i> Standl.	22	32	F69, G85
<i>Chloroleucon</i> (Benth.) Britton & Rose			
<i>Chloroleucon</i> sp		52	PS
<i>Desmanthus</i> Willd.			
<i>D. diffusum</i> Hook. & Arn.		22	GJ90
<i>D. illinoensis</i> (Michaux) Mac-Mill.		28	F69, GJ90
<i>D. leptolobus</i> Torr. & A. Gray		28	F69
<i>D. perambucanus</i> (L.) Thell.		26	PS
<i>D. ramosissimum</i> DC.		22	GJ90
<i>Desmanthus velutinus</i> Scheele		28	F69
<i>D. virgatus</i> (L.) Willd.		28	F69, GJ90

Continued on next page

Table 2. Continued.

Taxa	n	2n	* Source
<i>Dichrostachys</i> (DC.) Wight & Arn.			
<i>D. glomerata</i> Wight et Arn.		56	F69
<i>D. nyassana</i> Taub.		50	F69
<i>Dinizia</i> Dueke.		26, (-28)	G84
<i>D. excelsa</i> Dueke		28	F69, M77
<i>Dimorphandra</i> Schott			
<i>D. mollis</i> Benth.		26	F69
<i>Ebenopsis</i> Britton & Rose			
<i>E. ebanu</i> (Berland.) Barneby & J.W. Grimes		26	F69
<i>Entada</i> Adams.			
<i>E. abyssinica</i> Seud		28	F69
<i>E. africana</i> Guill. et Perr.		16, 28	F69
<i>E. gigas</i> (L.) Fawcett et Rendle.		28	F69
<i>E. nanni</i> (Oliv.) Tiss.	14	26	F69, GJ90
<i>E. phaseoloides</i> (L.) Merr.		28	GJ90
<i>E. pusaetha</i> DC.	14	~16	GJ90
<i>E. sudanica</i> (Schweinf) Kunth			F69
<i>Enterolobium</i> Mart.			
<i>E. contortisiliquum</i> (Vell.) Morong		26	F69, M77, GJ85, PS
<i>E. cyclocarpum</i> (Jacq.) Griseb	13	26	F69, M77, GJ91
<i>E. gummiiferum</i> (Mart.) Macbr.		26	M77
<i>Faidherbia</i> A. Chev.			
<i>F. albida</i> (Delile) A. Chev. (ex <i>Acacia albida</i> Delile)		26	F69, GJ03
<i>Gagnebina</i> Neek. ex DC.			
<i>G. pterocarpa</i> (Lam.) Baill sp		26	G84
<i>Goldmania</i> Rose ex Micheli			
<i>Harvadia</i> Small		26	G84
<i>H. acattensis</i> (Bentham) Britton & Rose		26	GJ96
<i>H. platyloba</i> (Sprengel) Britton & Rose		26	GJ96
<i>Inga</i> Mill.			
<i>I. bollandii</i> Sprague & Sandwith		26	M09
<i>I. capitata</i> Desv.		52	M09
<i>I. cayennensis</i> Sagot ex Benth.		26, ca. 104	M09
<i>I. cinnamomea</i> Benth.		26	PS
<i>I. cylindrica</i> (Vell.) Mart.		52	M09
<i>I. edulis</i> Mart.		26	F69, GJ85, M09, PS
<i>I. feuillei</i> DC.		26	GJ03
<i>I. ingoides</i> (Rich.) Willd.		26	M09

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Inga laurina</i> (Sw.) Willd.		26, 52	M09
<i>I. marginata</i> Willd.		26	M09
<i>I. striata</i> Benth.		26	M09
<i>I. subnuda</i> Salzm. ex Benth.		26	M09
<i>I. thibaudiana</i> DC.		26	M09
<i>I. vera</i> Willd.		26	M09
<i>I. vera</i> Will. subsp. <i>spuria</i> (Willd.) J. Leon		26	F69, G85
<i>Leucaena</i> Benth.			
<i>L. collinsii</i> Britton & Rose	26, 28	104	F69, GJ91
<i>L. cuspidata</i> Standley	56	52, 56	GJ91, GJ96, GJ03
<i>L. diversifolia</i> (Schldl.) Benth.	52+0-8Bs	52, 56, 104	GJ91, GJ96, GJ94
<i>L. diversifolia</i> subsp. <i>stenocarpa</i> (Urban) Zarate		56	GJ91
<i>L. diversifolia</i> subsp. <i>trichandra</i> Pan & Brewbaker	26+0-8b		GJ91
<i>L. esculenta</i> (Mocino & Sessé ex DC.) Benth.	26	56	GJ91
<i>L. esculenta</i> (Mocino & Sessé ex DC.) Benth. subsp. <i>esculenta</i>		56, 110	GJ96
<i>L. esculenta</i> subsp. <i>paniculata</i> (Britton & Rose) Zarate		~104	F69
<i>L. glabrata</i> Rose		36, 104	F69, G84, G85, GJ90, GJ91, GJ06
<i>L. glauca</i> (L.) Benth.	52		GJ91
<i>L. greggii</i> S. Watson	28		GJ03
<i>L. multicaupitula</i> Schery		52	GJ03
<i>L. pallida</i> Britton & Rose		56	GJ91, GJ03
<i>L. pueblana</i> Britton & Rose	52+0-8B	52, 56?	GJ03
<i>L. pulverulenta</i> (Schlecht.) Benth.	28	56	F69, GJ91
<i>L. retusa</i> Benth. ex A. Gray	28	52	GJ91, GJ03
<i>L. salvadorensis</i> Standley	28		GJ91
<i>L. salvadorensis</i> Standley ex Britton & Rose		56	GJ03
<i>L. shannonii</i> Donn. Sm.		52, 56	GJ03
<i>L. shannonii</i> J.D. Smith	26		GJ91
<i>L. trichodes</i> (Jacq.) Benth.	26	56	GJ91, GJ03
<i>Lysiloma</i> Benth.			
<i>L. bahamensis</i> Benth.		26	F69
<i>L. divaricata</i> Benth.		26	F69
<i>L. latistilqua</i> (L.) Benth.		26	F69
<i>L. tergenina</i> Benth.		26	F69
<i>Mimosa</i> L.			
<i>M. acantholoba</i> (Willd.) Poir. var. <i>acantholoba</i>	26		DN11
<i>M. acutistipula</i> Benth.	52		DN11
<i>M. adenocarpa</i> Benth.	26		DN11

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Mimosa adpressa</i> Hook & Arn	26II	52	GJ03
<i>M. aff. bathyrrhena</i>		26	DN11
<i>M. affinis</i> B.L. Rob.		26	DN11
<i>M. arenosa</i> (Willd.) Poir.		26	PS
<i>M. bimacronata</i> (DC.) Kuntze		26	F69, GJ03, DN11
<i>M. buuncifera</i> Benth.		52	F69, DN11
<i>M. blanchetii</i> Benth.		26	DN11
<i>M. borealis</i> A. Gray		26	DN11
<i>M. brevipedunculata</i> var. <i>hirtula</i> (Burkart) Barneby		52	GJ03
<i>M. caesalpinjifolia</i> Benth.		26	GJ00, DN11
<i>M. campicola</i> Harms		26, 78	DN11, PS
<i>M. camporum</i> Benth.		26	DN11
<i>M. candollei</i> R. Grether		52	DN11
<i>M. capitipes</i> Benth.	13	52	G84
<i>M. cisparanensis</i> Barneby		26	DN11
<i>M. clausenii</i> Benth.		26	M77, DN11
<i>M. cordistipula</i> Benth.		26, 32	DN11, PS
<i>M. cryptothamnus</i> Barneby		26	DN11
<i>M. daleoides</i> Benth.	13II [?] , ca 52	104?	G84, GJ03
<i>M. debilis</i> var. <i>vestita</i> (Benth.) Barneby		26	DN11
<i>M. delicatula</i> Baill.		26	DN11
<i>M. densa</i> Benth.		26	M77
<i>M. depauperata</i> Benth.		26	DN11
<i>M. diplotricha</i> C. Wright ex Sauvalle var. <i>diplotricha</i>	13	26	GJ96, DN11
<i>M. dolens</i> subsp. <i>acerba</i> (Bentham) Barneby		104	GJ96
<i>M. dolens</i> subsp. <i>rigida</i>		104	GJ96
<i>M. dominarum</i> Barneby		26	DN11
<i>M. dysocarpa</i> Benth.		26	DN11
<i>M. echinocaula</i> Benth.		26	DN10
<i>M. emoryana</i> Benth.		52	F69
<i>M. ephedroides</i> Benth.		28	F69
<i>M. flajellaris</i> Benth.		26	GJ06
<i>M. flocculosa</i> Burk.		26	DN11
<i>M. foliolosa</i> Benth.		26	DN11
<i>M. glandulifera</i> Burkart	13	26	GJ96
<i>M. goldmanii</i> Robinson		52	DN11
<i>M. gracilis</i> Benth.		26	DN11
<i>M. hamata</i> Willd.	13	26, 40	DN11
<i>M. heringeri</i> Barneby		26	F69, G81, G84, DN11
<i>M. hexandra</i> Micheli		26	DN11
<i>M. himalayana</i> Gamble	13	26	G81, G84

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Mimosa hirsutissima</i> var. <i>barbigera</i> (Benth.) Barneby		26	GJ03
<i>M. hirsutissima</i> Mart. var. <i>hirsutissima</i>	13II	26	DNI1
<i>M. hypoglauca</i> Mart. var. <i>hypoglauca</i>		26	M77
<i>M. imbricata</i> Benth.		52	DNI1
<i>M. incana</i> (Spreng.) Benth.		26	DNI1
<i>M. lacerata</i> Rose		26	G84
<i>M. lasiocarpa</i> Benth.	13	26	M77
<i>M. laticifera</i> Rizz. & Mattos		26	DNI1
<i>M. latispinosa</i> Lam		26	DNI1
<i>M. luisana</i> Brandegee		26	DNI1
<i>M. macrocalyx</i> Micheli		26	GJ03
<i>M. macrosiacha</i> (Benth.) Macbr.	13	26	G84
<i>M. malaccensis</i> (Mart.) Benth.		26	GJ00
<i>M. misera</i> Benth.		26	DNI1
<i>M. monancistrata</i> Benth.		26	DNI1
<i>M. neptunioides</i> Harms		26	DNI1
<i>M. niederleinii</i> Burk		52	DNI1
<i>M. nuda</i> Benth.		52	DNI1
<i>M. nuttallii</i> (DC.) B.L. Turner		26	DNI1
<i>M. obscurigosa</i> Burkart		13II	GJ03
<i>M. oligophylla</i> Micheli		26, 52	GL06
<i>M. oligophylla</i> var. <i>villosula</i> Burkart.		26	GJ03
<i>M. ophiolobocentra</i> Benth.		26	DNI1
<i>M. orthocarpa</i> Benth.		26	DNI1
<i>M. osbitigosa</i> Burkart	13II	26	GJ03
<i>M. papposa</i> Benth.		26	DNI1
<i>M. paratibana</i> Barneby		26	PS
<i>M. paupera</i> Bentham		26	GJ96
<i>M. pigra</i> L.	13, 26II	26	F69, M73, G85, GJ90, GJ03
<i>M. pigra</i> var. <i>delhiscens</i> (Baeneby) Glazier & Mackinder	13II	26, 52	GJ03, DNI1
<i>M. polyantha</i> Benth.		26	DNI1
<i>M. polycarpa</i> Kunth		26	GJ03, GJ06, DNI1
<i>M. polycarpa</i> var. <i>spagazzinii</i> Burkart	13	26	GJ96
<i>M. polydactyla</i> Willd.		26	DNI1
<i>M. polydyma</i> Barneby		26	DNI1
<i>M. pseudoseptaria</i> Harms		26	PS
<i>M. pteridifolia</i> Benth		26	DNI1
<i>M. pudica</i> L.	22, 26, 39II	32, 48, 52, 78	F69, M77, G81, G85, GJ90, GJ06
<i>M. pudica</i> L. var. <i>hispidata</i> Brenan		52	DNI1

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Mimosa pycnocoma</i> Benth.		52	DN11
<i>M. quadrivalvis</i> var. <i>leptocarpa</i> (DC.) Barneby		52	PS
<i>M. radula</i> Benth.		26	DN11
<i>M. revoluta</i> Benth.		26	DN11
<i>M. ramulosa</i> Benth.	52/11		G/06
<i>M. rixosa</i> Mart.	13		G84
<i>M. robusta</i> R. Grether		52	DN11
<i>M. rocae</i> Lorentz & Niederl.		104	G/06
<i>M. rubicautis</i> Lam.		26	M73, G81, G91
<i>M. rubicaulis</i> Lam. subsp. <i>himalayana</i> (Gamble) H. Ohashi	13, 13+0-2B	26	DN11
<i>M. scabrella</i> Benth.		52	DN11
<i>M. schomburgkii</i> Benth.		26	DN11
<i>M. selloi</i> (Benth.) Benth.		26	F69
<i>M. sensitiva</i> L.		26	G/00, DN11, PS
<i>M. sericantha</i> Benth.		26	DN11
<i>M. sepiaria</i> Benth.		26	PG91
<i>M. setosa</i> Benth. var. <i>paludosa</i> (Benth.) Barneby		26, 52	DN11, PS
<i>M. setosa</i> var. <i>urbica</i> Barneby		26	DN11
<i>M. setosissima</i> Taub		26	DN11
<i>M. similis</i> Britton & Rose		26	DN11
<i>M. skimmeri</i> Benth. var. <i>skimmeri</i>		52	DN11
<i>M. sominiensis</i> Willd.		26, 52	G/00, DN11, PS
<i>M. tenuiflora</i> (Willd.) Poir. (as <i>M. hostilis</i>)		26	G/00, DN11, PS
<i>M. tubrichiana</i> Harms		26	PS
<i>M. virgata</i> Barneby		26	DN11
<i>M. warnockii</i> Turner		26	F69
<i>Neptunia</i> Lour.			
<i>N. dimorphanta</i> Domin		28	F69
<i>N. gracilis</i> Benth.		56	F69
<i>N. lutea</i> (Leavenw.) Benth.		28	F69
<i>N. monosperma</i> Benth.		28	F69
<i>N. oleracea</i> Lour.		~52, 54, 56	F69, G81
<i>N. plena</i> (L.) Benth.	28	~72, 78	F69
<i>N. prostrata</i> (Lam) Bail.		56	F69
<i>N. pubescens</i> Benth.		28	F69
<i>N. triquetra</i> (Vahl) Benth.		36	F69
<i>N. uraguensis</i> Hooker & Arn.		26	G/96
<i>Newtonia</i> Bail.			
<i>N. Aubrevillei</i> (Pellegr.) Keay		26	F69
<i>Parappiadenia</i> Brenan			
<i>P. rigida</i> (Benth.) Brenan		26	G84, G/00

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Paraserianthes</i> I.C. Nielsen			
<i>P. lophantha</i> (Willd.) I.C. Nielsen (as <i>Acacia lophantha</i>)		24	F69
<i>Parkia</i> R. Br.			
<i>P. bicolor</i> A. Chev.		24	F69
<i>P. biglandulosa</i> Wight et Arn.		26	F69
<i>P. biglobosa</i> (Jacq.) Benth.		24, 26	F69
<i>P. javanica</i> (Lam.) Merr.		24	GJ90
<i>P. pendula</i> (Willd.) Walp.		22, 26	BK07, PS
<i>Pentaclethra</i> Benth.			
<i>P. macrophylla</i> Benth.	7	26	F69, GJ90
<i>Piptadenia</i> Benth.			
<i>P. macrocarpa</i> Benth.		26	F69
<i>P. obliqua</i> (Pers.) J.F. Macbr.		26	GJ00
<i>P. stipulacea</i> (Benth.) Ducke		26	GJ00
<i>P. viridiflora</i> (Kunth.) Benth.		26	PS
<i>Piptadeniastrum</i> Brenan			
<i>P. africanum</i> (Hook. f.) Brenan		26	F69
<i>Pithecellobium</i> Mart.			
<i>P. angulatum</i> Benth.			
<i>P. brevifolium</i> Benth.	13	26	RM73, RM74
<i>P. candidum</i> (Kunth) Bentham		26	F69
<i>P. dulce</i> Benth.		26	GJ96
<i>P. dulce</i> (Roxb.) Benth.		26	F69
<i>P. guadelupensis</i> Chapm.		26	PG84
<i>P. pallens</i> (Benth.) Standl.		26	F69
<i>P. polycephalum</i> Benth.		52	F69
<i>P. saman</i> (Jacq.) Benth.		26	F69
<i>P. scalare</i>		26	F69
<i>Plathyminia</i> Benth.			
<i>P. foliosa</i> Benth.		26	M77
<i>P. reticulata</i> Benth.		26	M77
<i>Prosopis</i> L.			
<i>P. africana</i> (Guill. & Perr.) Taub.		28	GJ03
<i>P. alba</i> Griseb.	14	28	F69, G81, GJ03
<i>P. algaribilla</i> Griseb.	14	28	G81
<i>P. alpataco</i> Phil.	14	28	F69, G85, G81
<i>P. argentina</i> Burkart		28, 56	F69, G81, G85
<i>P. caldenia</i> Burkart		28, 56	F69, G85
<i>P. campestris</i> Griseb.		28	F69
<i>P. chinensis</i> (Molina) Stuntz		28, 56	F69, G85, GJ03
<i>P. cineraria</i> (L.) Druce		26, 28	GJ91, GJ03, GJ06
<i>P. farcta</i> (Sol. ex Russe!) J.F. Macbr.		28, 56	PG84, GJ00, GJ06
<i>P. ferax</i> Griseb.		28	F69

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Prosopis flexuosa</i> DC.	14	28, 56	F69, G81, G85, GJ03
<i>P. glandulosa</i> Torr.		26, 28	F69, GJ94
<i>P. glandulosa</i> var. <i>torreyana</i> (L.D. Benson) MC Johnston		28	GJ94, GJ03
<i>P. hassleri</i> Harms.		28	G81
<i>P. humilis</i> Hook.		28,	
<i>P. humilis</i> Gill. ex Hook. et Am.		28, 56, ±112	F69, G85
<i>P. insularum</i> (Guill.) Bret. subsp. <i>novoguineensis</i> (Warb.) Brett.		52, 54	F69
<i>P. juliflora</i> DC.	13, 14, 26	28, 52, 56	F69, M73, M77, G81, G84, GJ00, GJ03
<i>P. juliflora</i> var. <i>glandulosa</i>		28, 56, ±112	G85
<i>P. juliflora</i> var. <i>torreyana</i>		28, 56	G85
<i>P. juliflora</i> var. <i>velutina</i>		28, 56	G85
<i>P. koelziana</i> Burkart		28	GJ06
<i>P. kuntzei</i> Harms.		28, 56, ±112	F69, G85
<i>P. laevigata</i> (Willd.) M.C. Johnston		28	G81, GJ03, GJ06
<i>P. nigra</i> (Griseb.) Hieron.	14	28	G81
<i>P. pallida</i> (Humb. & Bonpl. ex Willd.) Kunth		28	GJ03
<i>P. paragnica</i> Speg.		28	G81
<i>P. pubescens</i> Benth.		28, 56	G85
<i>P. reptans</i> Benth.		28, 56, ±112	G85
<i>P. ruscifolia</i> Burkart		28, 56, ±112	F69, G85
<i>P. ruscifolia</i> Griseb.	14	28	F69, G81
<i>P. sericantha</i> Hook		28, 56, ±112	F69, G85
<i>P. siliquastrum</i> (Cav.) DC.		28, 56	F69, G85, GJ03
<i>P. spicigera</i> L.	14	28	G84
<i>P. striata</i> Benth.		26, 56, ~112	F69
<i>P. strombulifera</i>		28	F69
<i>P. tamarugo</i> Phil.		28	G81
<i>P. torquata</i> (Lag.) DC.		28, 56	G85
<i>P. torquata</i> (Lag.) DC.		28	F69
<i>P. velutina</i> Wootton		28	G85
<i>Senegalia</i> Raf.			
<i>S. bahiensis</i> (Benth.) Seigler & Ebinger		26	PS
<i>S. riparia</i> (Kunth) Britton & Rose		26	PS
<i>S. tenuifolia</i> (L.) Britton & Rose		26	A48
<i>Schrankia</i> Willd.			
<i>S. leptocarpa</i> DC.	12	32, 52	M77, G84, GJ90
<i>S. uncinata</i> Willd.		26	G84
<i>Serianthes</i> Benth.			
<i>S. kanehirae</i> Fosberg.		26	G84

Continued on next page

Table 2. Continued.

Taxa	n	2n	*Source
<i>Stryphnodendron</i> Mart.			
<i>S. barbatum</i> Mart.		26	M77
<i>S. confertum</i> Her. & Rizz.		26	M77
<i>S. cristalinae</i> Her.		26	M77
<i>S. platyspicum</i> Rizz & Her.		26	M77
<i>Tetrapleura</i> Benth.			
<i>T. chevalieri</i> Baker f.		26	F69
<i>T. tetraptera</i> Taub.		26	F69
<i>Tachella</i> Wight & Arn.			
<i>T. farnesiana</i> (L.) Wight & Arn.		52	A48, PS
<i>Waiaceodendron</i> Koord.			
<i>W. celebicum</i> Koorders		26	G84
<i>Xylocarpus</i> Benth.			
<i>X. dolabriformis</i> Benth.	12		M74, M77, G81
<i>X. evansii</i> Hutch.			F69
<i>X. xylocarpa</i> Taub.	12	24	M74, M77, G81
<i>X. xylocarpa</i> (Roxb.) Taub. (as <i>X. dolabriformis</i> Benth.)	12		M74, M77, G81
<i>Zapoteca</i> H.M. Hern.			
<i>Z. albae</i> H. Hern.		26	GJ91
<i>Z. caracasana</i> subsp. <i>weberbaueri</i> (Harms) H. Hern.		26	GJ91
<i>Z. formosa</i> (Kunth) H. Hern.		26	GJ90, GJ91
<i>Z. media</i> (M. Martens & Galeotti) H. Hern.		26	GJ90, GJ91
<i>Z. portoricensis</i> (Jacq.) H. Hern. subsp. <i>portoricensis</i>		26	GJ90, GJ91
<i>Z. portoricensis</i> subsp. <i>flavida</i> (Urban) H. Hern.		26	GJ91
<i>Z. tehuana</i> H. Hern.		26	GJ91
<i>Z. tetragona</i> (Willd.) H. Hern.		26	GJ90, GJ91

*Source: A48 = Atchison, 1948; B74 = Bandel, 1974; BK07 = Barella and Karsburg, 2007; DN11 = Dahmer et al., 2011; F69 = Federov, 1969; M09 = Mata, 2009; M73 = Moore, 1973; M74 = Moore, 1974; M77 = Moore, 1977; G81 = Goldblatt, 1981; G84 = Goldblatt, 1984; G85 = Goldblatt, 1985; GJ90 = Goldblatt and Johnson, 1990; GJ91 = Goldblatt and Johnson, 1991; GJ94 = Goldblatt and Johnson, 1994; GJ96 = Goldblatt and Johnson, 1996; GJ00 = Goldblatt and Johnson, 2000; GJ03 = Goldblatt and Johnson, 2003; GJ06 = Goldblatt and Johnson, 2006; O10 = Ortolani et al., 2010; PS = present study.

Table 3. Chromosome numbers and probable base numbers (underlined>) of Mimosoideae (Leguminosae).

Tribes	Genera with the number of species known/analyzed	Chromosome Nos. and most probable base Nos. (underlined>)
Tribe Acacieae	<i>Acacia</i> Mill. (1450/160) <i>Senegalia</i> Raf. (160/3) <i>Vachellia</i> Wight & Am. (81/1)	<u>13</u> , 26, 52 13 26
Tribe Ingeae	<i>Abarema</i> Pittier (44/1) <i>Albizia</i> Durazz. (140/32) <i>Calliandra</i> Benth. (130/12) <i>Chloroleucon</i> (Benth.) Britton & Rose (24/1) <i>Enterolobium</i> Mart. (22/3) <i>Faidherbia</i> A. Chev. (1/1) <i>Harvardia</i> Small. (6/2) <i>Inga</i> Mill. (300/15) <i>Lysiloma</i> Benth. (11/4) <i>Pithecellobium</i> Mart. (110/9) <i>Wallacodendron</i> Koord. (1/1) <i>Zapoteca</i> H.M. Hem. (21/7)	13 <u>13</u> , 26, 34-52 8, 11 13 13 13 13 <u>13</u> , 26, 52 13 <u>13</u> , 26 13 13
Tribe Mimosae	<i>Adenanthera</i> L. (12/2) <i>Anadenanthera</i> Speg. (4/2) <i>Desmanthus</i> Willd. (24/7) <i>Dichrostachys</i> (DC.) Wight and Am. (43/2) <i>Dinizia</i> Ducke (1/1) <i>Dimorphandra</i> Schott (46/1) <i>Entada</i> Adans. (86/7) <i>Gagnebina</i> Neck. ex. DC. (12/1)	12-13, 14 13, 12 14, 11, 13 25-26 13 14, 13 14, 13 13
Tribe Mimoseae	<i>Goldmania</i> Rose ex Micheli (5/1) <i>Leucaena</i> Benth. (22/16) <i>Mimosa</i> L. (530/102) <i>Neptunia</i> Lour. (33/10) <i>Newtonia</i> Baill. (26/1) <i>Parapiptadenia</i> Brenan (6/1) <i>Parkia</i> R. Br. (77/6) <i>Pentaclethra</i> Benth. (11/1) <i>Piptadenia</i> Benth. (133/4) <i>Piptadeniastrium</i> Benth. (3/1) <i>Plathymenia</i> Benth. (3/2) <i>Prosopis</i> L. (111/38) <i>Schrankia</i> Willd. (46/2) <i>Stryphnodendron</i> Mart. (45/4) <i>Tetrapleura</i> Benth. (7/2) <i>Xylia</i> Benth. (16/4)	13 28, 26, 52 <u>13</u> , 26, 52 14, 28 13 13 12-13 13 13 13 13 14, 28, 56 12-13-16-28 13 13 12

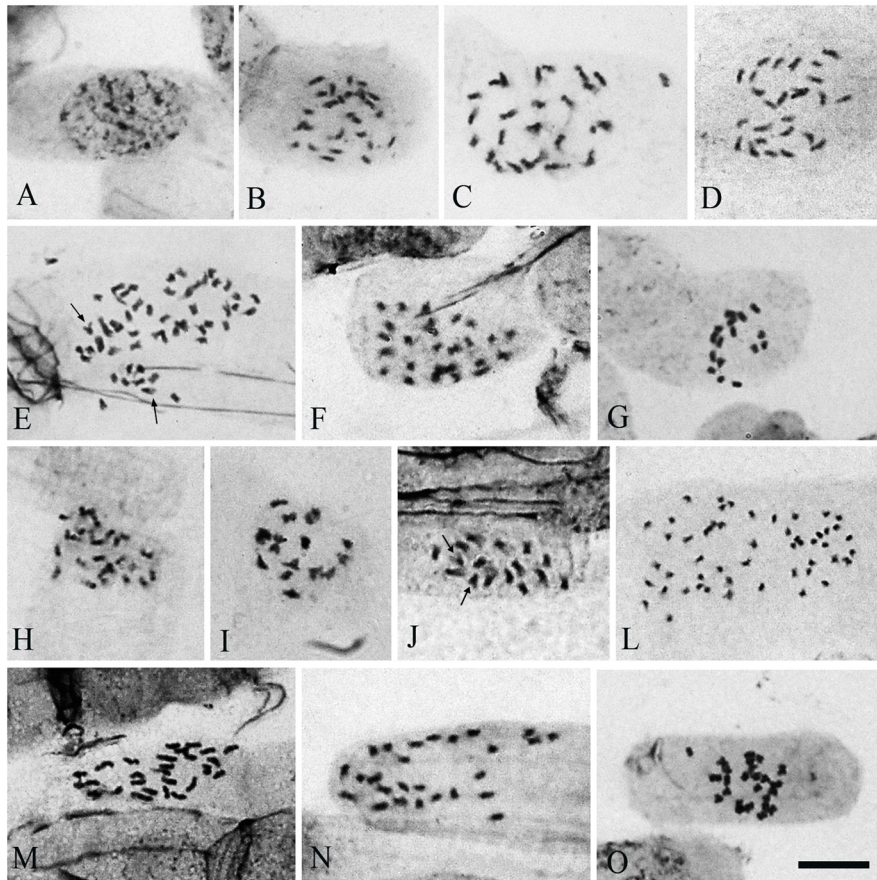


Figure 1. Chromosome complements and interphase nucleus of Leguminosae-Mimosoideae species of Acaciae and Ingeae tribes. **A.** Semi-reticulated interphase nucleus of *Senegalia riparia*; **B-D.** *Senegalia* species with $2n = 26$. *S. bahiensis* (**B**); *S. riparia* (**C**); *S. tenuifolia* (**D**); **E.** *Vachellia farnesiana* ($2n = 52$); **F.** *Abarema filamentosa* ($2n = 26$); **G.** *Calliandra depauperata* ($2n = 16$); **H.** *C. depauperata* ($2n = 26$); **I.** *C. leptopoda*; **J.** *C. portoricensis* ($2n = 16$); **L.** *Chloroleucon* sp ($2n = 52$); **M.** *Enterolobium contortisiliquum* ($2n = 26$); **N.** *Inga cinnamomea* ($2n = 26$); **O.** *I. edulis* ($2n = 26$). Arrows in **E** and **J** indicate acrocentric chromosomes. Bar = 10 μm .

For the tribe Mimosae, a single population of *Anadenanthera colubrina* (Figure 2A) from Caruaru, Pernambuco State, and 2 populations of *Desmanthus pernambucanus* (Figure 2B) from the states of Paraíba and Bahia, showed $2n = 26$. The chromosome numbers for the genus *Mimosa* were more variable, with $2n = 26$ in *M. arenosa* (Figure 2C), *M. paraibana* (Figure 2F), *M. pseudosepiaria* (Figure 2G), *M. sensitiva* (Figure 2I), *M. tenuiflora* (Figure 2L), *M. ulbrichiana* (Figure 2M), and *M. setosa* var. *paludosa* (Figure 2N); *M. cordistipula* had $2n = 32$ (Figure 2E), while 2 populations of *M. campicola* (Figure 2D) had $2n = 78$ and *M. quadrivalvis* var. *leptocarpa* (Figure 2H) and *M. somnians* (Figure 2J) had $2n = 52$. On the other hand, *P. pendula* (Figure 2O) and *P. viridiflora* (Figure 2P) were diploids with $2n = 26$.

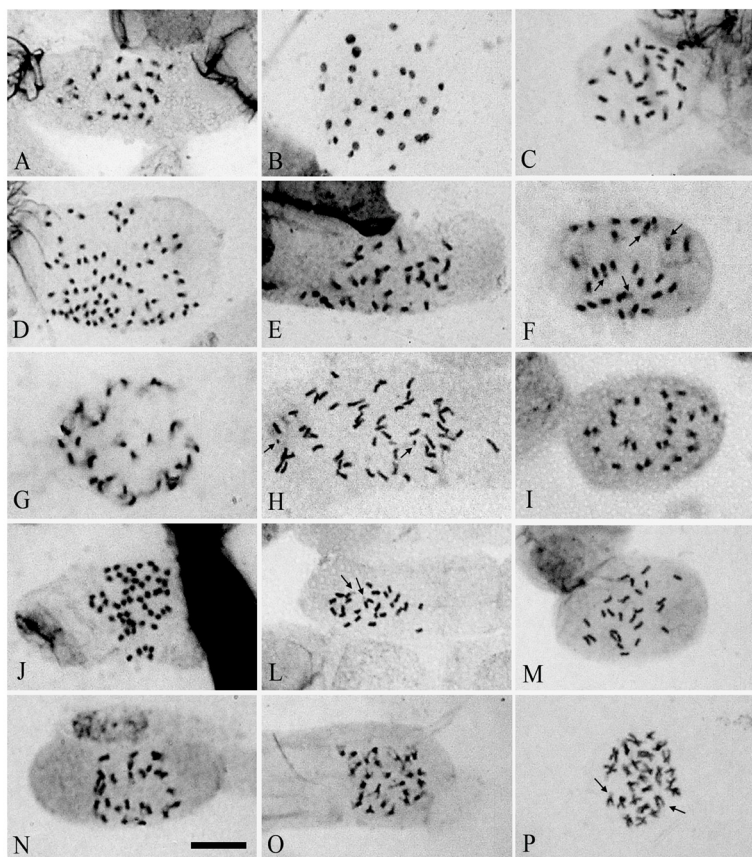


Figure 2. Chromosome complement of Leguminosae-Mimosoideae of the Mimoseae tribe. **A-C.** Species with $2n = 26$. **A.** *Anadenanthera columbrina*; **B.** *Desmathus pernambucanus*; **C.** *Mimosa aremosa*; **D.** *M. campicola* ($2n = 78$); **E.** *M. cordistipula* ($2n = 32$); **F.** *M. paraibana* ($2n = 26$); **G.** *M. pseudosepiaria* ($2n = 26$); **H.** *M. quadrivalvis* var. *leptocarpa* ($2n = 52$); **I.** *M. sensitiva* ($2n = 26$); **J.** *M. somnians* ($2n = 52$); **L.** *M. tenuiflora*; **M.** *M. ulbrichiana*; **N.** *Mimosa* sp. **O.** *Parkia pendula*; **P.** *Piptadenia viridiflora*; all showing $2n = 26$. Arrows in **F**, **H** and **L** indicate satellited chromosomes and in **P** indicate acrocentric chromosomes. Bar = 10 μ m.

DISCUSSION

The subfamily Mimosoideae comprises principally tropical and subtropical representatives of the American and African continents (Queiroz, 2009). Karyological records have been published for approximately 460 species (approximately 17% of the total number of species in the subfamily), and in the Leguminosae family, Mimosoideae is the least studied group in terms of karyological characteristics. Taxa from tropical regions have generally been less intensively studied cytologically than those from temperate regions. In addition to the generally neotropical distributions of the Leguminosae-Mimosoideae, the small sizes of their chromosomes and the apparent stability of their chromosome numbers present unfavorable conditions for more extensive chromosomal analyses. Thus far, no reports have been published on the

chromosome counts for 17 of the 27 species analyzed in the present study; additionally, we acquired the chromosome counts for the genus *Chloroleucon* for the first time. From previously published reports, we confirmed the counts of $2n = 26$ for *S. tenuifolia*, *E. contortisiliquum*, *I. edulis*, *A. colubrina*, *M. cordistipula*, *M. sensitiva*, *M. setosa* var. *paludosa*, and *M. tenuiflora*, and $2n = 52$ for *M. somnians* and *V. farnesiana*. The previous counts of $2n = 22$ for *P. pendula* (Barella and Karsburg, 2007), $2n = 26$ for *M. campicola*, *M. cordistipula*, and *M. somnians* and $2n = 52$ for *M. setosa* var. *paludosa* (Dahmer et al., 2011) were not confirmed. We obtained $2n = 26$ for *P. pendula* and *M. setosa* var. *paludosa*; $2n = 32$, for a population of *M. cordistipula*; $2n = 78$, for 2 populations of *M. campicola*, and $2n = 52$, for 2 other populations of *M. somnians*. The values obtained for the latter 2 groups of populations appear to be related to the occurrence of polyploidy, the karyotypic evolutionary process believed to be most active in the subfamily (Goldblatt, 1981a). Disploidy also seems to have an important role in the evolution of this group, and in some cases, may explain the discrepancies between the samples analyzed in this study and in the subfamily as a whole (see discussion below).

The importance of polyploidy to the evolution of the subfamily Mimosoideae can be appreciated by a study by Dahmer et al. (2011) wherein variations were observed in the chromosome numbers for a sample of 83 principally neotropical species of the genus *Mimosa*. These authors observed that 26% of the species analyzed were polyploids and that at least 3 of them exhibited intraspecific polyploidy. A similar evolutionary pattern of polyploidy was also observed in other South America species of *Mimosa* (Seijo and Fernández, 2001) and *Leucaena* (Boff and Schifino-Wittmann, 2003). A total of 13 species of the genus *Inga* from northeastern Brazil were analyzed, and 4 were found to be polyploids (2 of them showing intraspecific polyploidy; Mata, 2009, unpublished data). The divergence of the counts observed in our study from those previously published confirms the occurrence of intraspecific polyploidy for 3 of the species analyzed. On the other hand, the counts of $2n = 32$ obtained for *M. cordistipula* and $2n = 26$ for *P. pendula* (representing 7.4% of the species analyzed) indicate that dispoloidy may have had a very important role in the karyotypic evolution of the Mimosoideae from northeastern Brazil. On the other hand, analyses of previously observed numerical variations (Table 2) revealed only rare cases of intraspecific dispoloidy in the subfamily (1.46% of the species listed), which may have been a consequence of insufficient population sampling (commonly observed in older cytogenetic analyses) or counting errors (see, for example, Merxmüller, 1970).

Although the majority of the genera of this subfamily clearly show that their karyotypic evolution was regulated by numerous cycles of polyploidy with a basic number $x = 13$, various genera may have diverged from this principal lineage. In *Leucaena*, for example, there are paleopolyploid species with $2n = 52$ and 58, suggesting that there may have been 2 basic numbers-1 originating through dispoloidy from an ancestral stock with $x_2 = 26$ (Boff and Schifino-Wittmann, 2003). Genera, such as *Desmanthus*, *Entada*, and *Prosopis* (Table 2), have the basic number $x = 14$, which apparently resulted from ascending dispoloidy from a principal lineage with $x = 13$. On the other hand, almost all the species of the genus *Calliandra* with known chromosome numbers appear to have been derived from a distinct karyotypic lineage with $x = 8$ (Tables 2 and 3). Interestingly, 1 of the 3 species analyzed in the present study, namely, *C. depauperata*, had $2n = 26$ in 1 of the 2 populations studied—a number that is difficult to explain—as was the case in previously reported counts for the genus, such as $2n = 16, 22$ for *C. confusa* (Goldblatt, 1981a) and $2n = 22$ for *C. physiocalyx* (Hernández and Sousa, 1988).

On the whole, these counts indicate that more than 1 basic number may be involved in the karyotypic diversification of the genus *Calliandra*. The occurrence of $n = 7$ in various species of the genus *Cercis* and $n = 14$ in *Bauhinia* from the tribe Cercideae (which is considered a sister group of the other Leguminosae) (Wojciechowski et al., 2004), suggests a primary basic number of $x_1 = 7$ for the family as a whole (Goldblatt, 1981a). If this was true, *Bauhinia* would have originated by direct polyploidy from the ancestral stock with $x = 7$, while the species of *Calliandra* with $x = 8$ probably represent an old lineage derived from ascendant dispolyploidy. On the other hand, a majority of the genera of Mimosoideae paleopolyploids would have been formed by descending dispolyploidy from an ancestral paleopolyploid with $x = 14$ (Khatoun and Ali, 2006).

Polyploidy constitutes one of the principal mechanisms of speciation in plants, and it is estimated that more than 70% of all angiosperms are polyploids (Stebbins, 1971; Leitch and Bennett, 1997). Polyploids have been reported to have good ability to adapt to unstable environments and are most common in ecosystems that experience severe climatic stress, such as those in the arctic or glacial regions (Brochmann et al., 2004; Parisod et al., 2010) or in disturbed environments (Stebbins, 1971). The “Caatinga” biome covers about 70% of northeastern Brazil and is characterized by extreme water-related stress conditions and generally thin, rocky, and eroded soils. Some taxonomic groups, such as orchids of the genera *Oncidium* and *Epidendrum* (Felix and Guerra, 2000, 2010), and *Hoffmannseggella* (Yamagishi-Costa and Forni-Martins, 2009) have a strong tendency to form polyploids in environments that experience extreme conditions of abiotic stress. Other taxonomic groups, however, such as the families Convolvulaceae (Pitrez et al., 2008) and Amaryllidaceae (Felix et al., 2011), show no apparent correlation between polyploidy and environmental stress. The evolution of plant groups by the duplication of their genomes is generally rarer in tropical regions than in temperate zones, where environmental fluctuations are more frequent and are conducive to the formation of non-reduced gametes (Leitch and Bennett, 1997). This might explain the low frequency of reported polyploid species for the subfamily Mimosoideae and for other plant families that occur in tropical regions. The present study revealed that polyploid species occur in both “Caatinga” environments and altitudinal forests (a relatively stable meso-environment). Similarly, Dahmer et al. (2011) reported that some specimens from “Cerrado” (savanna), Atlantic forest, and “Caatinga” regions demonstrated no apparent correlation between polyploidy and high-stress environments. A correlation between polyploidy and abiotic stress has been demonstrated only in rare cases, and therefore, this phenomenon cannot be considered generic (reviewed by Parisod et al., 2010).

One of the explanations offered for the predominance of polyploidy, particularly allopolyploidy, as a predominant evolutionary process in angiosperms is the acquisition of evolutionary advantages associated with the formidable reorganization of the genome induced by polysomic inheritance. In these cases, polyploid plants that were established in disturbed environments could more easily colonize new and different habitats (Brochmann et al., 2004; Parisod et al., 2010). However, this process appears to have been stochastic for the subfamily Mimosoideae, recurring in diverse lineages and equally favoring polyploid populations in both stable and mesic environments (such as “Brejos de Altitude”) and in unstable environments subject to climatic stress (such as the dryland “Caatinga”). Most genera of Mimosoideae that were formed from ancient paleopolyploid species with $x = 13$ were apparently included in this process. Within the subfamily, the genus *Calliandra*, with $x = 8$, was the only lineage that

maintained a diploid level, and together with the genus *Cercis* (subfamily Caesalpinioideae), it represents one of the oldest diploid lineages related to an ancestral stock with $x = 7$.

ACKNOWLEDGMENTS

Research supported by CNPq.

REFERENCES

- Atchison E (1948). Studies in the *Leguminosae*. II. Cytogeography of *Acacia* (Tourn). *L. Am. J. Bot.* 35: 651-655.
- Atchison E (1951). Studies in the *Leguminosae* VI. Chromosome number among tropical woody species. *Am. J. Bot.* 38: 538-554.
- Bandel G (1974). Chromosome numbers and evolution in the Leguminosae. *Caryologia* 27: 17-32.
- Barella APW and Karsburg IV (2007). Caracterização morfológica dos cromossomos mitóticos de *Parkia pendula* (Willd.) Benth ex Walp. e *Dinizia excelsa* Ducke (Fabaceae, Mimosoideae). *Rev. Cienc. Agr.-Ambient.* 5: 85-93.
- Boff T and Schifino-Wittmann MT (2003). Segmental allopolyploidy and paleopolyploidy in species of *Leucaena* benth: evidence from meiotic behaviour analysis. *Hereditas* 138: 27-35.
- Brochmann C, Brysting AK, Alsos IG, Borgen L, et al. (2004). Polyploidy in arctic plants. *Biol. J. Linn. Soc.* 82: 521-536.
- Dahmer N, Simon MF, Schifino-Wittmann MT, Hughes CE, et al. (2011). Chromosome numbers in the genus *Mimosa* L.: cytotoxic and evolutionary implications. *Plant Syst. Evol.* 3: 211-220.
- Federov AMA (1969). Chromosome Numbers of Flowering Plants. Komarov Botanical Institute, Leningrad.
- Felix LP and Guerra M (2000). Cytogenetics and cytotoxicity of some Brazilian species of cymbidoid orchids. *Genet. Mol. Biol.* 23: 957-978.
- Felix LP and Guerra M (2010). Variation in chromosome number and basic number of subfamily Epidendroideae (Orchidaceae). *Bot. J. Linn. Soc.* 163: 234-278.
- Felix WJP, Felix LP, Melo NF, Oliveira MBM, et al. (2011). Karyotype variability in species of the genus *Zephyranthes* Herb. (Amaryllidaceae-Hippeastreae). *Plant. Syst. Evol.* 294: 263-271.
- Goldblatt P (1981a). Cytology and the Phylogeny of the Leguminosae. In: Advances in Legume Systematics, Part 1 (Polhill RM and Raven PM, eds.). Royal Botanic Gardens, Kew, 427-463.
- Goldblatt P (1981b). Index to Plant Chromosome Numbers 1975-1978. Missouri Bot. Gard., St Louis.
- Goldblatt P (1984). Index to Plant Chromosome Numbers 1979-1981. Missouri Bot. Gard., St Louis.
- Goldblatt P (1985). Index to Plant Chromosome Numbers 1982-1983. Missouri Bot. Gard., St Louis.
- Goldblatt P and Johnson DE (1990). Index to Plant Chromosome Numbers 1986-1987. Missouri Bot. Gard., St Louis.
- Goldblatt P and Johnson DE (1991). Index to Plant Chromosome Numbers 1988-1989. Missouri Bot. Gard., St Louis.
- Goldblatt P and Johnson DE (1994). Index to Plant Chromosome Numbers 1990-1991. Missouri Bot. Gard., St Louis.
- Goldblatt P and Johnson DE (1996). Index to Plant Chromosome Numbers 1992-1993. Missouri Bot. Gard., St Louis.
- Goldblatt P and Johnson DE (2000). Index to Plant Chromosome Numbers 1984-1985. Missouri Bot. Gard., St Louis.
- Goldblatt P and Johnson DE (2003). Index to Plant Chromosome Numbers 1998-2000. Missouri Bot. Gard., St Louis.
- Goldblatt P and Johnson DE (2006). Index to Plant Chromosome Numbers 2001-2003. Missouri Bot. Gard., St Louis.
- Guerra M (1983). O uso do Giemsa na citogenética vegetal - comparação entre a coloração simples e o bandeamento. *Cienc. Cult.* 35: 190-193.
- Guerra M (2000). Chromosome Number Variation and Evolution in Monocots. In: Monocots: Systematics and Evolution (Wilson KL and Morrison DA, eds.). CSIRO, Melbourne, 127-136.
- Hernández HM and Sousa M (1988). Two new species of *Calliandra* (Leguminosae: Mimosoideae) from southern Mexico. *Syst. Bot.* 13: 519-524.
- Judd WS, Campbell CS, Kellogg EA, Stevens PF, et al. (2009). Sistemática Vegetal: Um Enfoque Filogenético. Artmed, Porto Alegre.
- Khatoon S and Ali SI (2006). Chromosome Numbers and Polyploidy in the Legumes of Pakistan. *Pak. J. Bot.* 38: 935-945.
- Leitch IJ and Bennett MD (1997). Polyploidy in angiosperms. *Trends Plant Sci.* 12: 470-476.
- Luckow M (2005). Mimosae. In: Legumes of the World (Lewis GP, Schrine BD, Mackinder BA and Lock M, eds.). Royal Botanic Gardens, Kew.
- Mata MF (2009). O Gênero *Inga* (Leguminosae, Mimosoideae) no Nordeste do Brasil: Citogenética, Taxonomia e Tecnologia de Sementes. Doctoral thesis, Universidade Federal da Paraíba, UFPB, Areia.
- Merxmüller H (1970). Provocation of biosystematics. *Taxon* 19: 140-145.

- Moore RJ (1973). Index to plant chromosome number 1967-1971. *Regnum Veg.* 90: 1-539.
- Moore RJ (1974). Index to plant chromosome number 1972. *Regnum Veg.* 91: 1-108.
- Moore RJ (1977). Index to plant chromosome number 1973-1974. *Regnum Veg.* 91: 1-257.
- Ortolani FA, Melloni MNG, Mariotto CFG and Moro JR (2010). Caracterização citogenética em *Anadenanthera colubrina* (Vell.) Brenan (Mimosoideae) e *Guazuma ulmifolia* Lam. (Sterculiaceae). *Acta Bot. Bras.* 24: 299-303.
- Parisod C, Holderegger R and Brochmann C (2010). Evolutionary consequences of autopolyploidy. *New Phytol.* 186: 5-17.
- Pitrez SR, Andrade LA, Alves LIF and Felix LP (2008). Karyology of some Convolvulaceae species occurring in NE Brazil inselbergs. *Plant Syst. Evol.* 276: 235-241.
- Queiroz LP (2009). Leguminosas da Caatinga. Universidade Estadual de Feira de Santana, Feira de Santana.
- Seijo G and Fernández A (2001). Chromosome numbers of some southernmost species of *Mimosa* L. (Leguminosae). *Cytologia* 66: 19-23.
- Stebbins GL (1971). Chromosomal Evolution in Higher Plants. Edward Arnold, London.
- Wojciechowski MF, Lavin M and Sanderson MJ (2004). A phylogeny of legumes (Leguminosae) based on analysis of the plastid matK gene resolves many well-supported subclades within the family. *Am. J. Bot.* 91: 1846-1862.
- Yamagishi-Costa J and Forni-Martins ER (2009). Hybridization and polyploidy cytogenetic indications for *Hoffmannseggella* (Orchidaceae) species evolution. *Int. J. Bot.* 5: 93-99.