

# Analysis of the genetic divergence of soybean lines through hierarchical and Tocher optimization methods

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**ABSTRACT.** This study aimed to evaluate the clustering pattern consistency of soybean (*Glycine max*) lines, using seven different clustering methods. Our aim was to evaluate the best method for the identification of promising genotypes to obtain segregating populations. We used 51 generations F5 and F6 soybean lines originating from different hybridizations and backcrosses obtained from the soybean breeding program of Universidade Federal de Uberlândia in addition to three controls (Emgopa 302, BRSGO Luziânia, and MG/BR46 Conquista). We evaluated the following agronomic traits: number of

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days to flowering, number of days to maturity, height of the plant at maturity, insertion height of the first pod, grain yield, and weight of 100 seeds. The data was submitted to analyses of variance followed by average Euclidean distance matrix estimation used as measure of dissimilarity. Subsequently, clusters were formed using the Tocher method and dendrograms were constructed using the hierarchical methods simple connection (nearest neighbor), complete connection (most distant neighbor), Ward, median, average within cluster connection. The nearest neighbor method presented the largest number of genotypes in group I and showed the greatest similarity with the Tocher optimization method. The joint use of these two methodologies allows for differentiation of the most genetically distant genotypes that may constitute the optimal parents in a breeding program.

Key words: *Glycine max*; Genetic diversity; Genetic breeding

# **INTRODUCTION**

Among the main cultivated species, soybean [*Glycine max* (L.) Merrill] is the oleaginous plant with the greatest planted area in the world. It occupies a prominent position in the Brazilian economy, with an estimated production of 94.5 million tons in the 2014/2015 season (CONAB, 2015).

In soybean culture, artificial hybridization has been the used and most common method to increase the intraspecific genetic variability (Baenziger et al., 2006). However, in any breeding program, knowledge of the available germplasm in terms of performance, combination capacity, and genetic dissimilarity is of fundamental importance (Vieira et al., 2005). It has recently seen that modern soybean genotypes present a narrow genetic base complicating the development of superior genotypes (Dellagostin et al., 2011; Hamawaki et al., 2012; Yokomizo et al., 2013). Furthermore, it has been shown that the Brazilian soybean germplasm has maintained constant genetic variability in the last 30 years of expansion and breeding of crops with satisfactory results for many traits of interest (Priolli et al., 2004; Miranda et al., 2007; Wysmierski and Vello, 2013).

Due to the large amount of genes involved in the control of quantitative traits, it is unlikely that modern cultivars contain all the favorable alleles corresponding to all of the loci of agronomic interest in soybean (Mulato et al., 2010). By measuring the genetic divergence of four soybean populations, Hyten et al. (2006) concluded that the low genetic variability observed was likely due to the small number of Asiatic genetic material introduced during the process of domestication and not the selection process itself.

Studies of genetic divergence are important for understanding the genetic variability of populations. Such studies allow for monitoring the germplasm banks and selection of the most efficient crossings. They also generate important information with respect to genetic distance among the genotypes that may be used in the exploitation of the genetic variability in the development of new cultivars (Cruz et al., 2014).

To estimate the genetic diversity, one can, for example, use agronomic, morphological, and molecular traits (Silva et al., 2011). Within the available methods for genetic diversity studies, the average Euclidean distance is used as a measure of dissimilarity, which underlies hierarchical

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clustering methods such as UPGMA (unweighted pair group method with arithmetic mean), nearest neighbor, and the Tocher optimization method (Silva et al., 2011; Azevedo et al., 2013). The Tocher optimization method is normally used concomitantly with the UPGMA hierarchical method, providing correspondence for the allocation of elements in the clusters (Arriel et al., 2006). According to Hamawaki et al. (2012), an agreement between the Tocher optimization method and other hierarchical clustering or multivariate techniques can be used to compare methods.

The choice of analysis method to be utilized depends on the desired precision, ease of analysis, and form of obtaining the data and there is no defined parameter of choice for the study of genetic divergence for a group of genotypes (Cargnelutti Filho et al., 2008; Cruz et al., 2012). The measure of dissimilarity and together with analysis method used should guarantee the breeder security in the selection of parents for the crossings. If there is no agreement between the methods used, the choice of parents depends on the method utilized, which requires selection of the most efficient method (Cargnelutti Filho et al., 2008).

In this study, we evaluated the clustering pattern consistency of soybean lines, starting with a measure of dissimilarity obtained through the average Euclidean distance. Our aim was to identify the best method to indicate promising genotypes for obtaining segregating populations.

#### **MATERIAL AND METHODS**

The experiment was initiated on January 19, 2004 in the field of Fazenda Capim Branco of Universidade Federal de Uberlândia, located in the municipality of Uberlândia, Minas Gerais (18°55'23"S, 48°17'19"W, 872 m in altitude).

We evaluated 51 soybean lines from the F5 and F6 generations originating from different hybridizations and backcrosses obtained through the soybean breeding program of the Universidade Federal de Uberlândia. In addition, three controls (Emgopa 302, BRSGO Luziânia and MG/BR46 Conquista) were used. The genotypes were divided in three experiments. Each experiment consisted of 20 genotypes, composed of 17 lines and 3 controls (Table 1).

| Experiment 1      |      | Experiment 2      |      | Experiment 3      |      |  |
|-------------------|------|-------------------|------|-------------------|------|--|
| Genotype          | Code | 1                 | Code | 1                 | Code |  |
| UFU03104364       | 1    | Emgopa 302        | 21   | UFU03311105       | 41   |  |
| UFU03109183       | 2    | UFU03212045       | 22   | Emgopa 302        | 42   |  |
| UFU03112182       | 3    | UFU03204089       | 23   | UFU03306183       | 43   |  |
| UFU03113361       | 4    | UFU03210045       | 24   | UFU03310105       | 44   |  |
| UFU03103299       | 5    | UFU03213041       | 25   | BRSGO Luziânia    | 45   |  |
| UFU03117105       | 6    | UFU03209400       | 26   | UFU03301082       | 46   |  |
| UFU03101082       | 7    | BRSGO Luziânia    | 27   | UFU03302299       | 47   |  |
| Emgopa 302        | 8    | UFU03208400       | 28   | UFU03305187       | 48   |  |
| UFU03114136       | 9    | UFU03203004       | 29   | UFU03304178       | 49   |  |
| UFU03105146       | 10   | UFU03214041       | 30   | UFU03308249       | 50   |  |
| UFU03115105       | 11   | UFU03216204       | 31   | UFU03314062       | 51   |  |
| UFU03110183       | 12   | UFU03207408       | 32   | UFU03315213       | 52   |  |
| BRSGO Luziânia    | 13   | UFU03202004       | 33   | MG/BR46 Conquista | 53   |  |
| UFU03116105       | 14   | UFU03201183       | 34   | UFU03309087       | 54   |  |
| UFU03107178       | 15   | UFU03211062       | 35   | UFU03312089       | 55   |  |
| UFU03111007       | 16   | UFU03205089       | 36   | UFU03307007       | 56   |  |
| UFU03106146       | 17   | MG/BR46 Conquista | 37   | UFU03313089       | 57   |  |
| UFU03108178       | 18   | UFU03215213       | 38   | UFU03317204       | 58   |  |
| MG/BR46 Conquista | 19   | UFU03206232       | 39   | UFU03316204       | 59   |  |
| UFU03102299       | 20   | UFU03217204       | 40   | UFU03303146       | 60   |  |

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In all three experiments, we used a complete randomized block design composed of 20 genotypes and three replications. Each experimental plot consisted of soybean plants planted along a 6-m line, with 0.90 m between lines, totaling 180 plots.

The following agronomic traits were evaluated: number of days to flowering-R1 (NDF), number of days to maturity-R8 (NDM), height of the plant at maturity (HPM), insertion height of the first pod (IHFP), grain yield (YIELD), and weight of 100 seeds (WHS). The soybean development stages (R1 and R8) were identified following the phenological scale of Fehr and Caviness (1977).

The data were analyzed with analysis of variance using the F-test to verify the genetic variability among lines and cultivars using the software SAEG-UFV (SAEG, 2007). The average Euclidean distance matrices between the lines and controls were estimated and used as measures of dissimilarity. Subsequently, clusters were formed using the Tocher method and dendrograms were constructed using the following hierarchical methods simple connection (nearest neighbor), complete connection (most distant neighbor), Ward, median (WPGMA), average within cluster connection, and UPGMA. In the hierarchical clustering methods were established at 60% dissimilarity as criterion for the formation of clusters in the dendrograms.

The coefficient of cophenetic correlation between the matrix of average Euclidean distance and the matrix of cophenetic distance was obtained from the dendrogram of each method. All analyses were conducted in GENES (Cruz, 2013).

### **RESULTS**

CV exp3 (%)

Genetic variability for the traits NDF, HPM, YIELD, and WHS was identified in at least two of the three experiments (Table 2). However, for IHFP, genetic variability was only observed in experiment 2, indicating low genetic variability between soybean lines. For YIELD, we identified genetic variability in all experiments suggesting that this trait can be explored for breeding. The coefficient of variation ranged between 1.35-23.87%, indicating high experimental precision.

| cultivated  | d in Uberlând | ia-Mir | as Gerais, du      | ring the 2004/2 | 2005 season.       |                     |        |             |
|-------------|---------------|--------|--------------------|-----------------|--------------------|---------------------|--------|-------------|
| Experiment  | SV            | d.f    |                    |                 | Ν                  | ٨S                  |        |             |
|             |               |        | NDF                | NDM             | IHFP               | HPM                 | WHS    | YIELD       |
| 1           | Genotypes     | 19     | 19.79*             | 136.20**        | 0.60 <sup>ns</sup> | 144.19**            | 8.18** | 132923.40** |
| 2           | Genotypes     | 19     | 1.67 <sup>ns</sup> | 125.24**        | 0.55**             | 154.82**            | 7.99** | 9605.65**   |
| 3           | Genotypes     | 19     | 31.14**            | 89.46**         | 0.54 <sup>ns</sup> | 83.38 <sup>ns</sup> | 8.84** | 91939.15*   |
| CV exp1 (%) |               |        | 6.16               | 4.68            | 23.87              | 11.33               | 8.21   | 21.66       |
| CV exp2 (%) |               |        | 2.73               | 3.76            | 14.62              | 11.47               | 9.72   | 18.36       |
| CV exp3 (%) |               |        | 1.35               | 2.64            | 21.76              | 15.60               | 7.43   | 19.99       |

Table 2. Summary of analyses of variance in experiments 1-3 with 20 soybean (*Glycine max*) genotypes,

SV: source of variation; d.f.: degrees of freedom; MS: mean square; CV: coefficient of variation. NDF: number of days to flowering; NDM: number of days to maturity; IHFP: insertion height of the first pod; HPM: height of the plant at maturity; WHS: weight of 100 seeds; YIELD: yield. Asterisks (\* and \*\*) indicate significant differences at 5 and 1% probability, respectively, based on an F-test; nsnon-significant difference.

Based on the Tocher clustering method (Table 3) 75, 95, and 70% of the genotypes were grouped in cluster I in experiments 1, 2, and 3, respectively, indicating high a similarity among the evaluated genotypes. The controls (BRSGO Luziânia and MG/BR 46 Conquista) grouped into cluster I in all experiments demonstrating genetic similarity

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among these lines. In experiment 2, the genotypes were grouped into only two clusters. This indicates that the genetic base of this experiment was small, which contributes to the low genetic diversity observed.

**Table 3.** Results from the Tocher optimization method in three experiments with soybean (*Glycine max*) genotypes and three controls, cultivated in Uberlândia-Minas Gerais, during the 2004/2005 season.

| Group | Experiment 1   |
|-------|--|
| Ι     | 3, 5, 13 <sup>(2)</sup> , 4, 12, 14, 2, 16, 1, 20, 11, 6, 15, 18, and 19 <sup>(3)</sup>                                    |
| II    | 10 and 17  |
| III   | 7 and 9  |
| IV    | 8(1)   |
| Group | Experiment 2   |
| I     | 22, 24, 32, 31, 25, 30, 23, 21 <sup>(1)</sup> , 27 <sup>(2)</sup> , 28, 36, 38, 26, 35, 37 <sup>(3)</sup> , 39, 29, and 34 |
| II    | 40   |
| Group | Experiment 3   |
| I     | 58, 60, 56, 59, 57, 46, 47, 53 <sup>(3)</sup> , 54, 49, 45 <sup>(2)</sup> , 41, 44, and 55                                 |
| II    | 50 and 51  |
| III   | 43 and 52  |
| IV    | 48   |
| V     | 42 <sup>(1)</sup>  |

<sup>(1)</sup>Emgopa 302, <sup>(2)</sup>BRSGO Luziânia, and <sup>(3)</sup>MG/BR46 Conquista.

The hierarchical clusters obtained from the dissimilarity matrices demonstrated that the majority of the evaluated genotypes could be found in cluster I in all three experiments (Figures 1 to 18). The genotypes grouped into cluster I in the hierarchical methods were more similar to the results using the Tocher optimization method. In experiments 2 and 3, only the nearest neighbor method presented 100 and 95% similarity, respectively, which cluster I obtained with the Tocher method (Figure 7 and 13). The comparison between the optimization and the hierarchical methods demonstrated that the dendrogram cluster presented the identification of the highest number of clusters and, consequently, improved possibility of identifying divergent parents.

The average within cluster connection proved to reveal the greatest number of clusters, with the formation of 8, 8, and 9 clusters in experiments 1, 2, and 3, respectively. By contrast, the Ward method resulted in the smallest number of clusters, indicating relatively homogeneous distribution of the genotypes. The WPGMA method presented genotype clusterings similar to those of the UPGMA method in all experiments, indicating that these methods are similar to one another.



**Figure 1.** Clustering of 20 soybean (*Glycine max*) genotypes using the simple connection (nearest neighbor) method, from the Euclidean distances obtained based on six agronomic traits from experiment 1.

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**Figure 2.** Clustering of 20 soybean (*Glycine max*) genotypes using the complete connection (most distant neighbor) method, from the Euclidean distances obtained based on six agronomic traits from experiment 1.



Figure 3. Clustering of 20 soybean (*Glycine max*) genotypes using the Ward method, from the Euclidean distances obtained based on six agronomic traits from experiment 1.



Figure 4. Clustering of 20 soybean (*Glycine max*) genotypes using the median method (WPGMA), from the Euclidean distances obtained based on six agronomic traits from experiment 1.



**Figure 5.** Clustering of 20 soybean (*Glycine max*) genotypes using the average within cluster connection method, from the Euclidean distances obtained based on six agronomic traits from experiment 1.

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**Figure 6.** Clustering of 20 soybean (*Glycine max*) genotypes using the average among cluster connection method from the Euclidean distances obtained based on six agronomic traits from experiment 1.



**Figure 7.** Clustering of 20 soybean (*Glycine max*) genotypes using the simple connection (nearest neighbor) method, from the Euclidean distances obtained based on six agronomic traits from experiment 2.



**Figure 8.** Clustering of 20 soybean (*Glycine max*) genotypes using the complete connection (most distant neighbor) method, from the Euclidean distances obtained based on six agronomic traits from experiment 2.



Figure 9. Clustering of 20 soybean (*Glycine max*) genotypes using the Ward method, from the Euclidean distances obtained based on six agronomic traits from experiment 2.

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Figure 10. Clustering of 20 soybean (*Glycine max*) genotypes using the median method (WPGMA), from the Euclidean distances obtained based on six agronomic traits from experiment 2.



**Figure 11.** Clustering of 20 soybean (*Glycine max*) genotypes using the average within cluster connection method, from the Euclidean distances obtained based on six agronomic traits from experiment 2.



**Figure 12.** Clustering of 20 soybean (*Glycine max*) genotypes using the average among cluster connection method from the Euclidean distances obtained based on six agronomic traits from experiment 2.



**Figure 13.** Clustering of 20 soybean (*Glycine max*) genotypes using the simple connection (nearest neighbor) method, from the Euclidean distances obtained based on six agronomic traits from experiment 3.

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**Figure 14.** Clustering of 20 soybean (*Glycine max*) genotypes using the complete connection (most distant neighbor) method, from the Euclidean distances obtained based on six agronomic traits from experiment 3.



Figure 15. Clustering of 20 soybean (*Glycine max*) genotypes using the Ward method, from the Euclidean distances obtained based on six agronomic traits from experiment 3.



Figure 16. Clustering of 20 soybean (*Glycine max*) genotypes using the median method (WPGMA), from the Euclidean distances obtained based on six agronomic traits from experiment 3.



**Figure 17.** Clustering of 20 soybean (*Glycine max*) genotypes using the average within cluster connection method, from the Euclidean distances obtained based on six agronomic traits from experiment 3.

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Figure 18. Clustering of 20 soybean (*Glycine max*) genotypes using the average among cluster connection method from the Euclidean distances obtained based on six agronomic traits from experiment 3.

The coefficient of cophenetic correlation obtained between the matrix of average Euclidean distances and the matrix of cophenetic distances, from the dendrogram of each method ranged between 0.52 and 0.83 and was statistically significant (Table 4). The nearest neighbor, WPGMA, and UPGMA methods presented values above 0.7 for experiments 2 and 3, respectively, suggesting that these methods were consistent.

| soybean ( <i>Glycine max</i> ) genotyp | es.                                   |              |              |  |  |
|--|---------------------------------------|--------------|--------------|--|--|
| Methodologies                          | Coefficient of cophenetic correlation |              |              |  |  |
|  | Experiment 1                          | Experiment 2 | Experiment 3 |  |  |
| Nearest neighbor                       | 0.74*                                 | 0.58*        | 0.79*        |  |  |
| Most distant neighbor                  | 0.72*                                 | 0.67*        | 0.60*        |  |  |
| Ward                                   | 0.70*                                 | 0.59*        | 0.52*        |  |  |
| WPGMA                                  | 0.76*                                 | 0.74*        | 0.77*        |  |  |
| Average within cluster connection      | 0.77*                                 | 0.59*        | 0.64*        |  |  |
| UPGMA                                  | 0.77*                                 | 0.75*        | 0.83*        |  |  |

**Table 4.** Coefficients of cophenetic correlation obtained between the matrix of average Euclidean distances and the matrix of cophenetic distances for the different hierarchical methodologies in three experiments with soybean (*Glycine max*) genotypes.

\*Significant at 1% probability based on a t-test.

#### DISCUSSION

The study of genetic divergence is an important tool to identify parents that contrast the most for crosses seeking to increase the variability through breeding. Knowledge of genetic diversity can be used to produce new cultivars (Veloso et al., 2015). In our study, all investigated traits presented genetic variability in at least two experiments with exception of IHFP. IHFP is a trait important for breeding, with the ideal IHFP value being between 10 and 12 cm to minimize loss during mechanical harvest (Sediyama, 2015). Therefore, this trait is often considered during the selection of soybean lines.

In this study, the identified coefficients of variation were similar with values found in others studies. Selestrino et al. (2014) found coefficient of variation values ranging between 1.37 and 22.27% for NDF, NDM, IHFP, HPM, YIELD, and WHS. According to Carvalho et al. (2003), the maximum acceptable limit for the coefficient of variation in relation to yield and height of the soybean plant is 16 and 12%, respectively. We found that the coefficients of variation for yield were higher than those established by Carvalho et al. (2003) and exceeded the 20% that is required for the inscription of new cultivars in the National Registry of

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Cultivars, the Ministry of Agriculture, Livestock, and Food Supply (MAPA, 1998).

Clustering methods allow for the identification of genotypes genetically that are contrasting for the crosses. The first cluster in a dendrogram is formed by genotypes sharing the greatest similarity (Cruz et al., 2014). We observed some clusters composed of only one or two genotypes (Table 3), and the control cultivar Emgopa 302 proved to be the most divergent among all evaluated genotypes. Selection of divergent parents should be based on the magnitude of the genetic divergence among genotypes, when the aim is to perform crossings resulting in superior progenies in relation to the traits of interest. Vieira et al. (2005) reported that clusters formed by one single individual suggest that those individuals are the most divergent in relation to the rest. According to Abreu et al. (2004), knowledge of genetic divergence allows inferences to be made about the specific combination capacity before carrying out the crossings, resulting in a greater chance of identifying and recovering more promising combinations among the segregating populations.

Our comparative study between the optimization and hierarchical methods demonstrated that the clusters in the dendrograms could be used for identification of the method yielding the highest number of clusters and, consequently, constitute an improvement in the possibility of selecting the most divergent parents. Rotili et al. (2012) suggested that, compared to the Tocher method, the nearest neighbor method can be used with the objective of evaluating the genetic divergence among corn genotypes.

Vogt et al. (2010) also confirmed that the UPGMA and Tocher optimization methods were in agreement in a study of genetic divergence of sunflowers. Furthermore, Bertan et al. (2006) confirmed that the clustering of genotypes using UPGMA resulted in results that were similar to the method of cluster formation among the most divergent genotypes, suggesting optimization method agreement method. Further studies have shown that the UPGMA method is superior to the nearest neighbor and most distant neighbor methods when compared with known information of the genealogy (Sneath and Sokal, 1973; Romesburg, 1984; Dudley, 1994).

Using various clustering methods for processing the data, using different measures of dissimilarity and taking into account the particularities of each method are required for an accurate choice of parents for crossings (Cargnelutti Filho et al., 2008). Through the use of dendrograms, it is possible to evaluate the formation of clusters and, consequently, to select the genetically most distinct genotypes.

The reference information regarding genetic divergence is typically not sufficient for selecting parents for hybridization (Ferreira Júnior et al., 2015). Instead, the reference information should be accompanied by information about the genotype performance in relation to the desirable traits. The ideal strategy is one that combines the results of genetic divergence analyses and the identification of the genotypes carrying the traits that are the target of the breeding program.

#### **Conflicts of interest**

The authors declare no conflict of interest.

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