

FROM SACRED GROVES TO GLOBAL PERFUMERY: A COMPREHENSIVE REVIEW OF MAGNOLIA CHAMPACA IN ORNAMENTAL HORTICULTURE

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ABSTRACT

Magnolia champaca (L.) Baill, commonly known as Champak or Golden Champa, is a tropical and subtropical flowering tree of immense ornamental, cultural, economic, and pharmaceutical significance. Belonging to the family Magnoliaceae, this species is indigenous to South and Southeast Asia and has been cultivated for centuries for its intensely fragrant flowers, ornamental appeal, timber value, and medicinal properties. In the context of ornamental horticulture and floriculture, *M. champaca* occupies a unique niche as both a landscape tree and a source of commercial loose flowers, garland making and essential oil for the perfume industry. A mature, well-managed tree yields 15–30 kg of fresh flowers per year; intensive commercial plantations in peninsular India achieve 3–8 tonnes per hectare annually. *Champaca* absolute commands a global market price of USD 800–3,000+ per kg, placing it among the most valuable natural floral extracts traded internationally. The principal floral volatile, methyl (E)-cinnamate, constitutes 35–55% of total headspace volatiles in the yellow-flowered form.

Despite its widespread traditional use and considerable economic potential, systematic scientific research on its agronomy, production technology, floral biology, breeding, and post-harvest management remains fragmented and largely underexplored. This comprehensive review synthesizes available scientific literature on *M. champaca* from over 29 references, covering its taxonomic classification, morphological characteristics, ecological requirements, agronomy and commercial cultivation practices, floral biology and reproductive physiology, crop improvement strategies including emerging genomic tools, post-harvest management, value addition prospects, climate change vulnerability, and current industry scenario with future prospects. The review identifies critical knowledge gaps in standardized production protocols, systematic breeding programs, CRISPR-based metabolic engineering, and market development that must be addressed to fully realize the potential of this species in modern ornamental horticulture.

KEYWORDS: Champak, *Magnolia champaca*, floriculture, essential oil, floral biology, ornamental horticulture, post-harvest, crop improvement, climate change, genomics

INTRODUCTION

The global floriculture industry has witnessed exponential growth over the past three decades, driven by rising consumer demand for ornamental plants, cut flowers, fragrances, and herbal products. Within this dynamic industry, tropical flowering trees and shrubs have gained considerable attention due to their unique aesthetic appeal, cultural significance, and multifarious commercial applications. Among these *Magnolia champaca* (L.) Baill. ex Pierre popularly known as Champak, Golden champa, Yellow jade orchid tree, or Joy Perfume tree stands out as one of the most fragrant and economically versatile flowering tree species in the world.

Magnolia champaca belongs to the ancient angiosperm family Magnoliaceae, which represents one of the earliest flowering plant lineages. The species is native to the tropical and subtropical regions of South Asia (India, Nepal, Sri Lanka) and Southeast Asia (Myanmar, Thailand, Indonesia, Malaysia, Philippines, Vietnam, China), where it grows naturally in moist deciduous and semi-evergreen forests at elevations ranging from sea level to approximately 1,600 m. In India alone, it has been revered for over 3,000 years in religious, cultural, and medicinal contexts featured prominently in Sanskrit literature, the Ayurvedic pharmacopoeia, and traditional ceremonies.

From an ornamental horticultural perspective, *M. champaca* offers a compelling combination of attributes: large, glossy evergreen foliage; year-round to seasonally profuse flowering; an intensely rich and distinctive floral fragrance; bright orange to yellowish-white flower coloration; and a stately tree form suitable for avenue planting, parks, gardens, and home landscapes. The flowers are harvested extensively for use in religious offerings, garland making, hair adornment, and as a raw material for the extraction of *Champaca* absolute — one of the most prized natural fragrance ingredients in the global perfume industry.

Despite its immense ethnobotanical heritage and commercial significance, *M. champaca* has not received the same systematic scientific attention as other ornamental floriculture crops such as roses, chrysanthemums, or jasmine. Research on its standardized agronomy, commercial production systems, floral biology, reproductive physiology, post-harvest technology, and genetic improvement is sparse and geographically scattered.

1.1 Scope and Methodology

This review was prepared through a systematic search of the scientific literature using the databases Web of Science, Scopus, PubMed, Google Scholar, and CAB Abstracts. Search terms included 'Magnolia champaca floriculture, essential oil, floral biology, post-harvest and breeding'. The search covered publications from 1980 to 2025, with emphasis on peer-reviewed journal articles, review papers, book chapters, and technical bulletins from national agricultural research institutions. Grey literature including FAO reports, state agricultural university extension bulletins, and CIMAP technical publications was also incorporated where relevant. A total of 125 references *M. champaca* data were absent, particularly in sections on genetics, genomics, and pollination biology.

2. Taxonomy, Botanical Description, and Genetic Resources

2.1 Taxonomic Classification and Nomenclature

The species now recognized as *Magnolia champaca* has undergone considerable taxonomic revision. It was originally described by Linnaeus as *Michelia champaca* L. in *Species Plantarum* (1753) and was long classified under the genus *Michelia* within the family *Magnoliaceae*. Following extensive molecular phylogenetic analyses, which demonstrated that *Michelia* and several related genera were nested within *Magnolia sensu lato*, the species was reclassified as *Magnolia champaca* (L.) Baill. ex Pierre under the APG III and APG IV systems (Angiosperm Phylogeny Group, 2016). The name *Michelia champaca* continues to appear widely in horticultural, pharmaceutical, and ethnobotanical literature and is treated as a synonym throughout this review.

Table 1. Taxonomic classification of *Magnolia champaca* (L.) Baill. ex Pierre

Taxonomic Rank	Classification
Kingdom	Plantae
Division	Magnoliophyta (Angiosperms)
Class	Magnoliopsida (Dicotyledons)
Order	Magnoliales
Family	Magnoliaceae
Genus	<i>Magnolia</i> L.
Species	<i>M. champaca</i> (L.) Baill. ex Pierre
Synonyms	<i>Michelia champaca</i> L.; <i>Michelia aurantiaca</i> Wall.
Common Names	Champak, Golden Champa, Joy Perfume Tree, Sampangi (Telugu), Shenbagam (Tamil), Champa (Hindi/Sanskrit)

2.2 Morphological Description

Magnolia champaca is a medium to large evergreen or semi-deciduous tree attaining heights of 10–50 m in natural habitats, though in cultivation it is typically maintained at 5–15 m. The bark is grey, smooth to slightly fissured. Leaves are alternate, simple, ovate-lanceolate to elliptic, 10–25 cm long and 5–10 cm wide, with an acuminate apex, cuneate base, and entire margin; the upper surface is deep glossy green, lower surface paler and slightly pubescent.

The flowers are the most striking feature of the species, produced singly in the axils of leaves throughout the year in tropical climates, with peak flowering occurring in summer (March–July in India). Each flower measures 3–5 cm in diameter and consists of 15–21 tepals arranged in three to seven whorls of three; they are creamy-white to golden-yellow or orange in colour, depending on the cultivar, and intensely fragrant. The fragrance is strongest in the early morning hours (5:00–8:00 AM). The gynoecium is an elongated cone of numerous free carpels. Fruits are follicetums bearing 1–3 red-orange seeds with a fleshy aril.

2.3 Intraspecific Variation and Cultivars

Natural and cultivated populations of *M. champaca* show considerable intraspecific variation in flower colour, tepal number, fragrance intensity, tree architecture, and phenology. Two broad color forms are widely recognized: the golden-yellow to orange-flowered form (*forma champaca*) and a white-flowered form (*forma alba*). In horticulture, white *champaca* is preferred for religious use and cut flower production, while the yellow-orange form is the primary source of *champaca* absolute for the perfume industry. Several improved selections have been identified in India, Thailand, and Indonesia, though formal variety registration remains limited.

2.4 Genetic Resources and Germplasm

The genetic diversity of *M. champaca* is harbored across its native range in South and Southeast Asia. In India, significant germplasm accessions are maintained at NBPGR, CIMAP, and several State Agricultural Universities. Molecular characterization using RAPD, ISSR, SSR, and next-generation sequencing approaches has revealed high

genetic diversity within natural populations. The nuclear ribosomal ITS and chloroplast DNA markers have been employed for phylogenetic studies and species delimitation within the *Magnolia–Michelia* complex.

3. Ecological Requirements and Adaptability

3.1 Climatic Requirements

Magnolia champaca is a tropical to subtropical species adapted to warm, humid climates. It thrives optimally under mean annual temperatures of 20–35°C, annual rainfall of 1,200–2,500 mm well-distributed throughout the year, relative humidity of 60–80%, and bright sunshine of 6–8 hours per day. It is predominantly cultivated in the humid tropics of South and Southeast Asia but has naturalized in parts of tropical Africa, the Caribbean, Hawaii, and Queensland, Australia.

Flowering in *M. champaca* is influenced by photoperiod, temperature, and water stress. In seasonal climates, a brief dry spell or mild thermal stress can induce synchronized heavy flowering. Temperature fluctuation between day and night enhances floral fragrance intensity, explaining the superior quality of *champaca* flowers from certain highland growing regions in South India and Indonesia.

3.2 Soil Requirements

The species grows best in deep, well-drained, loamy soils rich in organic matter with a slightly acidic to neutral pH (5.5–7.0). While it can tolerate a range of soil types including sandy loam and red laterite soils, waterlogged or highly compact soils are detrimental. Good soil drainage is essential, as the roots are susceptible to *Phytophthora*-associated root rots under stagnant moisture conditions.

3.3 Altitude and Geographic Distribution

In India, *M. champaca* is cultivated extensively in Tamil Nadu, Karnataka, Kerala, Andhra Pradesh, Odisha, West Bengal and Assam spanning altitudes from sea level to about 1,200 m. It is also an important floriculture crop in Thailand, Indonesia, the Philippines, and Vietnam. Its cultivation range is expanding with rising interest from subtropical regions where protected cultivation can mitigate cold stress.

4. Agronomy and Crop Management

4.1 Propagation Methods

4.1.1 Seed Propagation

Seeds are the traditional propagation method for *M. champaca*. Fresh seeds have good germination viability (60–85%) but lose viability rapidly and should be sown within 2–4 weeks of collection. Pre-sowing treatments including soaking in water (24–48 hours) or treatment with gibberellic acid (GA3 at 500–1,000 ppm) enhance germination speed and uniformity. Germination typically occurs in 21–45 days under warm, humid nursery conditions. Seedling trees take 5–8 years to reach commercial flowering age.

4.1.2 Vegetative Propagation

Vegetative propagation is increasingly preferred for commercial cultivation as it ensures genetic uniformity, retention of superior parental traits, and significantly reduced time to first flowering (2–4 years). Air-layering (Gootee/Marcotting) is the most widely practiced method, with success rates of 60–90% when applied during the rainy season on 1–2-year-old pencil-thick branches. Application of IBA (Indole-3-butyric acid) at 5,000–10,000 ppm in lanolin paste to the girdled zone markedly improves rooting success. Semi-hardwood stem cuttings treated with IBA at 5,000–8,000 ppm and maintained under mist propagation benches show 50–80% rooting success. Grafting on seedling rootstocks achieves 40–75% success, while patch and shield budding during periods of active rootstock growth achieves 55–70% success.

Table 2. Comparison of propagation methods for *Magnolia champaca*

Method	Success Rate (%)	Time to Flowering	Key Treatment	Preferred Season
Seed	60–85% germination	5–8 years	GA3 500–1,000 ppm soak	Post-monsoon
Air-layering	60–90	2–4 years	IBA 5,000–10,000 ppm lanolin	Monsoon
Stem cuttings	50–80	2–4 years	IBA 5,000–8,000 ppm + mist bench	Rainy season
Grafting	40–75	2–3 years	Cleft/splice on seedling rootstock	Active growth
Budding	55–70	2–3 years	Patch/shield budding	Active growth
Micropropagation	Variable (protocol-dependent)	1–2 years	BAP/Kinetin + IBA/NAA on MS	Year-round (lab)

			medium	
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4.1.3 Micropropagation

In vitro propagation (tissue culture) offers the most promising avenue for large-scale clonal multiplication of elite genotypes. Protocols have been developed using nodal explants, shoot apices, and immature floral buds on Murashige and Skoog (MS) medium supplemented with cytokinins (BAP, Kinetin) for shoot proliferation and auxins (IBA, NAA) for rooting. Somatic embryogenesis has also been reported from callus cultures. However, commercial-scale micropropagation is yet to be standardized and widely adopted, primarily due to recalcitrance to tissue culture and somaclonal variation during regeneration.

4.2 Field Establishment and Spacing

Site preparation involves deep ploughing (30–45 cm), incorporation of organic matter (20–25 t/ha FYM), and pit digging (60 × 60 × 60 cm) filled with a mixture of topsoil, compost, and basal fertilizer. For intensive commercial flower production, spacings of 4 × 4 m (625 plants/ha) or 3 × 4 m (833 plants/ha) are practiced in South India. For timber-cum-flower production, wider spacings of 6 × 6 m (278 plants/ha) are used. Planting is ideally done at the onset of the monsoon season.

4.3 Nutrient Management

The recommended annual dose per adult tree (5+ years) is approximately N at 300–500 g, P₂O₅ at 200–300 g, and K₂O at 300–400 g, applied in 2–3 split doses. Organic manure application (10–15 kg FYM + 250 g neem cake/tree/year) significantly improves soil biological activity, nutrient use efficiency, and flower aroma quality. Micronutrient deficiencies, particularly zinc, iron, and boron, are commonly observed in acid laterite soils and are corrected through foliar sprays.

4.4 Irrigation Management

Adequate soil moisture is essential during establishment, active shoot growth, and flower bud development stages. Drip irrigation with soil moisture-based scheduling is increasingly adopted in commercial plantations, allowing water savings of 40–60% compared to flood irrigation. Irrigation is typically reduced 2–4 weeks before the expected flowering peak to mildly stress the tree and synchronize flowering.

4.5 Pruning and Canopy Management

Since flowers are borne on current-season shoots, rejuvenation pruning after each major harvest flush promotes the development of new bearing wood. Light to moderate pruning (removal of 20–30% of the canopy) after the summer flush, combined with a fertilizer dose and irrigation, effectively induces a secondary flush of vegetative growth and flowers. Heavy pruning is generally avoided as it can weaken the tree.

4.6 Pest and Disease Management

Key pests include scale insects (*Pulvinaria* sp.), mites (*Oligonychus* sp.), stem borers (*Batocera rufomaculata*), and flower thrips (*Scirtothrips dorsalis*, *Thrips hawaiiensis*). Among diseases, collar rot and root rot caused by *Phytophthora* species, leaf spot diseases (*Pestalotiopsis*, *Colletotrichum*), and powdery mildew have been reported. Integrated pest management (IPM) strategies combining cultural, biological, and judicious chemical interventions are advocated for sustainable production.

5. Floral Biology and Reproductive Physiology

5.1 Flower Morphology and Anatomy

The flowers of *M. champaca* are solitary, axillary, hermaphrodite, and hypogynous. Each flower consists of 15–21 tepals arranged spirally on a receptacle, creamy-white to golden-orange in colour. The tepal surface contains epidermal secretory cells producing characteristic floral volatile compounds. The stamens are numerous (50–200+), arranged spirally. The gynoecium is apocarpous with 10–20 free carpels, each containing 2–4 ovules.

5.2 Anthesis, Pollen Biology, and Fruit Set

Anthesis in *M. champaca* follows a characteristic protogynous (stigma receptive before pollen shedding) sequence, which promotes cross-pollination. The flower opens in stages over 2–3 days: on day 1 (female phase), the stigmas are receptive but anthers are immature; on day 2–3 (male phase), the tepals open fully and anthers dehisce to shed pollen. This temporal separation ensures a high degree of cross-pollination. Pollen grains are monosulcate, measuring 30–45 µm in length, and viability declines rapidly under ambient conditions but can be preserved for 4–6 weeks at –20°C.

Fruit set under natural pollination conditions ranges from 15–35% of flowers, with significant variation between populations and seasons. Hand cross-pollination with compatible pollen donors improves fruit set to 40–65%, confirming partial self-incompatibility. Each mature follicle produces 1–3 seeds with a fleshy red-orange aril, dispersed primarily by birds.

5.3 Floral Fragrance and Volatile Chemistry

The floral fragrance of *M. champaca* is among the most celebrated in the plant kingdom — described as sweet, fruity, tea-like, spicy, and deeply floral. GC-MS analyses have identified over 100 volatile compounds. Major components

include: methyl (E)-cinnamate (the key odor compound, contributing 35–55% of total volatiles in yellow champaca); linalool and its oxides; beta-caryophyllene; alpha-terpineol; indole; methyl benzoate; benzyl acetate; geraniol; and nerol. The fragrance intensity shows a distinct diurnal pattern, peaking in the early morning hours (5:00–8:00 AM). Significant differences in volatile profiles exist between the yellow-flowered (forma champaca) and white-flowered (forma alba) types, the latter being richer in linalool and poorer in methyl cinnamate.

Table 3. Major floral volatile compounds in *Magnolia champaca* (yellow vs. white-flowered forms)

Compound	Chemical Class	Yellow Form (%)	White Form (%)	Odor Descriptor
Methyl (E)-cinnamate	Phenylpropanoid	35–55	5–15	Sweet, balsamic, spicy
Linalool	Monoterpenol	8–18	25–45	Floral, lavender
beta-Caryophyllene	Sesquiterpene	3–8	2–6	Woody, spicy
Methyl benzoate	Benzenoid	2–6	2–5	Fruity, floral
Indole	Nitrogen heterocycle	1–4	2–5	Floral, animalic
alpha-Terpineol	Monoterpenol	1–5	3–8	Lilac, pine
Geraniol + Nerol	Monoterpenol	1–4	4–10	Rose, citrus
Benzyl acetate	Benzenoid ester	1–3	1–4	Jasmine-like

5.4 Pollination Biology

Magnolia champaca is pollinated primarily by beetles (cantharophily) — a primitive pollination syndrome consistent with the ancient phylogenetic position of Magnoliaceae. Scarab beetles (Coleoptera: Scarabaeidae), particularly species of *Protaetia*, are the primary pollinators that visit flowers to feed on tepals, pollen, and floral secretions. Flies, thrips, and occasionally bees also contribute to pollination. The protogynous floral sequence and beetle-trap mechanism, wherein tepals partially enclose the beetles during the male phase, are adaptations maximizing cross-pollination efficiency.

5.5 Flower Yield and Seasonality

A mature, well-managed *M. champaca* tree (8–15 years) under commercial cultivation in South India typically yields 15–30 kg of fresh flowers per year, with annual productivity in intensive commercial plantations reported at 3–8 tonnes per hectare. Peak flowering occurs during the hot summer months (March–June) in peninsular India, with a secondary flush in September–November. In truly tropical regions, flowering can be nearly year-round. Synchronized heavy flowering can be induced by imposing a brief irrigation stress (2–4 weeks) during the pre-induction period, a practice widely employed in Tamil Nadu and Kerala.

6. Crop Improvement Strategies

6.1 Germplasm Evaluation and Selection

Systematic evaluation of available germplasm for key horticultural traits is the first step in any crop improvement program. Selection criteria include flower yield per tree, fragrance intensity and quality (assessed by GC-MS profiling), flower color and size, shelf life, duration of the flowering season, precocity, tree architecture, and tolerance to abiotic stresses. Germplasm evaluation programs have been initiated at SAU departments of floriculture in Tamil Nadu, Kerala, and Karnataka, with preliminary identification of high-yielding clones showing 30–50% superior yield over unimproved populations.

6.2 Conventional Breeding Approaches

Controlled hybridization within *M. champaca* populations and between *M. champaca* and related *Magnolia*/*Michelia* species represents a largely unexploited avenue for generating new variation. The hand-pollination protocol involves emasculation during the female phase, followed by application of pollen from the selected male parent during the early male phase. Crossing within *M. champaca* for traits such as white × yellow flower color, compact × standard tree architecture, and high volatile yield × long vase life offers promising prospects. Interspecific hybridization with *M. alba*, *M. doltsopa*, *M. grandiflora*, and *M. figo* has been attempted with variable success; F1 hybrids between *M. champaca* and *M. alba* have been obtained and are reported to combine the intense fragrance of champaca with the larger flower size of *M. alba*.

Table 4. Documented interspecific hybrids involving *Magnolia champaca* and their key traits

Cross	Hybrid / Outcome	Key Traits	Reference
<i>M. champaca</i> × <i>M.</i>	F1 hybrids obtained	Intense fragrance + larger	Liu & Luo, 2009

alba		flowers	
<i>M. champaca</i> × <i>M. doltsopa</i>	Variable success	Cold tolerance (potential)	Flanagan et al., 2016
<i>M. champaca</i> × <i>M. figo</i>	Low success rate	Compact architecture	Zhang et al., 2003
<i>M. champaca</i> × <i>M. grandiflora</i>	Attempted; limited data	Increased flower size	Unpublished / SAU trials

6.3 Polyploidy and Mutation Breeding

Induction of polyploidy through colchicine treatment of shoot apices, germinating seeds, or in vitro shoot cultures offers a strategy for developing improved cultivars with larger flowers, increased fragrance compound production, and enhanced ornamental appeal. Colchicine concentrations of 0.01–0.5% applied for 24–48 hours have been used to induce chromosome doubling. Physical mutagens (gamma radiation at 10–60 Gy) and chemical mutagens (EMS at 0.1–0.5%) have been applied to seeds and vegetative propagules in preliminary studies, generating variants in flower color, tepal number, and growth habit.

6.4 Biotechnological Approaches

6.4.1 Molecular Markers and Genomics

Molecular marker systems including RAPD, ISSR, SSR, and SNP-based markers have been applied to characterize genetic diversity and elucidate phylogenetic relationships within Magnoliaceae. No high-quality reference genome sequence for *M. champaca* has been published to date, though the genome of *Liriodendron chinense* (a related Magnoliid) provides a reference for comparative genomic studies (Chen et al., 2019). Transcriptomic studies (RNA-seq) of *M. champaca* flowers have identified genes involved in floral volatile biosynthesis, including methyl cinnamate biosynthetic pathway enzymes and linalool synthase (Wang et al., 2018). Whole-genome sequencing of *M. champaca* is a high-priority research need that would enable marker-assisted selection, QTL mapping for fragrance and yield traits, and targeted metabolic engineering.

6.4.2 CRISPR/Cas9 and Metabolic Engineering Prospects

The CRISPR/Cas9 system has emerged as a transformative tool for precision crop improvement, and its application in ornamental tree species is now being actively explored. In *M. champaca*, CRISPR-based approaches hold potential for: (i) upregulation of methyl cinnamate biosynthesis by overexpression of phenylalanine ammonia-lyase (PAL) and cinnamate-CoA ligase (CCL) pathway genes; (ii) silencing of competitive metabolic branches that divert carbon flux away from desirable floral volatiles; (iii) modification of flower color through targeted editing of chalcone synthase (CHS) or flavonoid 3',5'-hydroxylase (F3'5'H) genes; and (iv) modification of flowering time genes (FT, SOC1 orthologs) for extended or season-independent flowering. *Agrobacterium tumefaciens*-mediated genetic transformation of *M. champaca* has been demonstrated at proof-of-concept level, but stable heritable transformation remains a challenge due to recalcitrance of Magnoliaceae to tissue culture and low transformation efficiency. Protocols optimized for related *Magnolia* species (particularly *M. officinalis*) may offer transferable methodologies.

6.4.3 Transcriptomics and Floral Scent Engineering

RNA-seq-based transcriptome profiling of *M. champaca* flowers at different developmental stages and times of day has identified key biosynthetic genes responsible for diurnal fragrance emission patterns. Differential expression of methyl cinnamate pathway genes peaks at dawn, consistent with the observed fragrance intensity maximum. These transcriptomic datasets provide a roadmap for metabolic engineering targeted at enhancing economically important volatile yields in both the fresh flower and essential oil production contexts.

7. Post-Harvest Management

7.1 Harvesting

Flowers are harvested at the partially-open bud stage in the early morning hours (4:00–7:00 AM), when fragrance content is highest and temperature is low. Fully open flowers have a significantly shorter shelf life (12–24 hours). Flowers are hand-picked individually and collected in clean, ventilated containers — traditionally woven baskets lined with moist cloth — to minimize mechanical damage and prevent heat build-up.

7.2 Post-Harvest Handling and Storage

The major post-harvest challenges are extremely short vase life (12–36 hours at ambient temperature), rapid browning of tepals (due to enzymatic oxidation via polyphenol oxidase), and fragrance loss through volatilization and enzymatic degradation. Under ambient tropical conditions (30–35°C), flowers deteriorate within 12–18 hours of harvest. Multiple treatments have been evaluated to extend vase life, as summarized in Table 5.

Table 5. Comparative post-harvest treatments for extending vase life of *Magnolia champaca* flowers

Treatment	Concentration / Conditions	Vase Life Achieved	Mechanism	Reference
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Ambient (control)	30–35°C	12–18 h	—	Baseline
Cold storage	8–12°C	48–72 h	Reduces respiration & ethylene	Nagalakshmi et al., 2001
8-HQC + Sucrose pulse	200–300 ppm + 2–5% sucrose	36–48 h	Antimicrobial + energy supply	Vaidyanathan & Bhatt, 2006
Silver thiosulfate (STS)	0.2–0.4 mM pulse	48–60 h	Ethylene inhibition	Krishnakumar et al., 1997
Calcium chloride dip	1–2% CaCl ₂ , 30 min	30–42 h	Membrane stabilization	Renuka Devi & Bhimappa, 2016
Modified atmosphere packaging (MAP)	3–5% O ₂ + 5–8% CO ₂	48–72 h	Reduced respiration + oxidation	Nagalakshmi et al., 2001
Perforated PE packaging	25–50 µm PE, 1% perforation	36–48 h	Reduces moisture loss	Renuka Devi & Bhimappa, 2016
Cold + STS + MAP (combined)	10°C + 0.2 mM STS + 4% O ₂	72–96 h	Synergistic multi-mechanism	Estimated; requires validation

7.3 Grading and Packaging

Commercial grading is based on flower size, color intensity, freedom from physical damage, and fragrance intensity. Flowers are typically graded into Extra, Grade A, Grade B, and Market Quality categories. Packaging for short-distance domestic market involves traditional leaf-wrapping or polyethylene bags; for premium export markets, flowers are packaged in ventilated clamshell containers with moist cellulose padding. Cold chain maintenance from farm to consumer includes pre-cooling, refrigerated transport and retail display at 10–15°C is critical for maintaining quality in distant markets.

8. Value Addition and Industrial Applications

8.1 Essential Oil and Absolute Extraction

The most economically significant value-added product from *M. champaca* flowers is Champaca Absolute is a highly concentrated floral extract used in premium perfumery and cosmetics. The absolute is produced by solvent extraction (hexane extraction to produce a concrete, followed by alcohol washing) or by enfleurage. The yield of concrete from *M. champaca* flowers is approximately 0.2–0.4% of fresh flower weight; the absolute yield from the concrete is 40–60%. Champaca absolute commands premium prices of USD 800–3,000+ per kg in the global aromatic ingredient market, making it one of the most valuable natural floral extracts traded globally.

Steam distillation yields a floral water (hydrosol) and a small amount (0.01–0.05%) of essential oil dominated by methyl (E)-cinnamate, linalool, and sesquiterpene hydrocarbons. Supercritical CO₂ extraction (SCCO₂ at 40–50°C and 100–200 bar) is an emerging green extraction technology producing extracts with superior aroma fidelity and no solvent residues, and holds particular promise for premium-grade champaca extract production.

8.2 Medicinal and Pharmaceutical Applications

Magnolia champaca has a long history of medicinal use in Ayurvedic, Siddha, Unani, and traditional Southeast Asian medical systems. Modern phytochemical investigations have identified alkaloids (magnoflorine, liriodenine, champacine), flavonoids (quercetin, kaempferol, isorhamnetin glycosides), lignans, sesqui- and diterpenoids, and volatile phenylpropanoids (methyl cinnamate, eugenol).

Table 6. Medicinal applications of *Magnolia champaca* plant parts: traditional uses, bioactive compounds, and pharmacological activities

Plant Part	Traditional Use	Bioactive Compounds	Pharmacological Activity
Flowers	Headache, fever, skin disorders	Methyl cinnamate, linalool, flavonoids	Anti-inflammatory, antioxidant, antimicrobial

Bark	Fever, rheumatism, menstrual disorders	Magnoflorine, liriodenine, tannins	Antipyretic, antiprotozoal, uterotonic,
Seeds	Gout, rheumatism, skin diseases	Fixed oil, alkaloids	Anti-arthritic, antibacterial
Leaves	Headache, skin disorders	Volatile oils, flavonoids, lignans	Antifungal, antibacterial
Roots	Kidney stones, urinary complaints	Alkaloids, phenolics	Diuretic, anti-urolithiatic

8.3 Food, Cosmetics, and Agroforestry

Fresh flowers are used to flavor tea (Champaca tea), rice dishes, and sweets in Indian and Southeast Asian cultures. Champaca essential oil has GRAS (Generally Recognized As Safe) status for use as a food flavoring at low concentrations. In the cosmetics sector, *M. champaca* extracts are incorporated into luxury body oils, skin creams, hair care preparations, soaps, and aromatherapy products. The anti-inflammatory and antioxidant properties make them attractive ingredients in anti-aging formulations. Several luxury perfume houses feature champaca as a key ingredient in their compositions. *M. champaca* also yields moderately hard, durable timber (density ~550–650 kg/m³) and serves as an excellent shade tree for coffee, cardamom, and pepper in agroforestry systems, contributing to farm diversification and soil improvement.

9. Climate Change Impacts and Adaptation

9.1 Projected Vulnerabilities

Climate change poses significant and underappreciated threats to *M. champaca* cultivation and essential oil quality. Rising mean temperatures in major production zones of peninsular India (Tamil Nadu, Karnataka, Kerala) projected to increase by 1.5–3.0°C by 2050 under SSP2-4.5 scenarios (IPCC, 2021) are expected to alter flowering phenology, potentially shifting peak flowering earlier by 2–4 weeks and disrupting synchronized market supply. Critically, the diurnal temperature differential that drives fragrance volatile accumulation in highland champaca-growing areas (Nilgiris, Coorg) may be reduced under warming scenarios, potentially diminishing the quality premium these regions command in the champaca absolute market.

Altered monsoon patterns including delayed onset, increased rainfall intensity, and extended dry spells threaten crop establishment and stress-induced flowering synchronization, which currently depends on predictable dry periods. The intensification of extreme weather events (cyclones, flooding) poses direct risks to productive champaca plantations in coastal Karnataka, Kerala, and Odisha. Additionally, projected expansions of the geographic range of thrips (*Scirtothrips dorsalis*) and mite (*Oligonychus* sp.) populations under warmer winters may increase pest pressure and post-harvest losses.

9.2 Adaptation Strategies

Proactive adaptation strategies are essential for sustaining the cultivation and commercial production of *Magnolia champaca* under changing climatic conditions. The development and deployment of heat-tolerant and drought-tolerant genotypes through systematic germplasm screening and marker-assisted breeding approaches could significantly enhance crop resilience. The adoption of precision irrigation systems, particularly drip irrigation integrated with soil moisture sensors, may help mitigate the adverse effects of rainfall variability and water scarcity. Similarly, agroforestry-based intercropping systems can contribute to microclimate regulation by moderating temperature and maintaining relative humidity within production environments. The formulation of climate-smart planting calendars based on downscaled climate projections for major cultivation regions would further support adaptive crop management practices. In addition, the establishment of improved post-harvest infrastructure, including cold storage facilities and refrigerated transportation systems, is necessary to reduce quality deterioration and post-harvest losses under elevated ambient temperatures. Over the long term, breeding efforts focused on developing champaca cultivars with modified flowering responses that are less dependent on specific temperature or rainfall cues may provide greater stability against climate-induced phenological disruptions.

10. Industry Scenario, Market Trends, and Future Prospects

10.1 Domestic Market in India

India is the dominant producer and consumer of *M. champaca* flowers. The bulk of production occurs in Tamil Nadu (Salem, Namakkal, Dharmapuri, Coimbatore, Nilgiris), Kerala (Thrissur, Palakkad, Kozhikode), Karnataka (Hassan, Chikkamagaluru), and Andhra Pradesh. The domestic market is driven by the massive religious and ritual flower market, the garland and ornamental market, the hair flower market in South India, and raw material supply for essential oil extraction. Market prices at farm-gate level range from INR 200–1,500/kg depending on season, quality, and location.

10.2 Global Export Market for Champaca Absolute

The global export market for champaca absolute is concentrated in the premium and luxury perfumery segment. Major importing countries are France, the United States, Germany, the United Kingdom, Japan, and the United Arab Emirates.

India (particularly Kerala and Tamil Nadu) and Indonesia (Java) are the primary producing and exporting countries. Growing consumer preference for natural, sustainable, and traceable fragrance ingredients in fine perfumery is a strong market driver. The increasing demand for certified organic and Fair-Trade aromatic ingredients present a value-addition opportunity for organized champaca producer groups in South India and Indonesia.

10.3 Challenges and Constraints

Several significant challenges constrain the full realization of *Magnolia champaca* commercial potential. One of the major limitations is the lack of improved varieties, as no officially released high-yielding and disease-resistant cultivars are currently available either in India or internationally. Flower harvesting is highly labor-intensive and requires pre-dawn collection, which restricts large-scale commercial expansion. In addition, the fresh flowers possess an ultra-short shelf life, and the inadequacy of cold chain infrastructure results in considerable post-harvest losses ranging from 20–40%. Production is also highly fragmented, being dominated by small-scale farmers with land holdings of only 0.1–1 ha, who often have limited access to technical guidance and organized marketing systems. Another serious issue is the adulteration of essential oil with synthetic methyl cinnamate, which negatively affects market integrity and reduces price realization for genuine products. Furthermore, research investment in this crop remains limited when compared to other commercially important floriculture crops. Climate change has also emerged as a growing concern, causing phenological shifts and quality-related risks, particularly in fragrance-premium highland production zones.

10.4 Future Research Priorities

The prospects for *M. champaca* in ornamental horticulture are highly promising, provided targeted investments are made in:

- (1) Variety development through systematic germplasm evaluation, hybridization, and official cultivar release programs with improved varieties with defined fragrance profiles, yield targets, and growing zone suitability are needed.
- (2) Standardization of location-specific cultivation technology packages covering nursery management, spacing, nutrition, irrigation, and flowering regulation.
- (3) Reference genome sequencing of *M. champaca* to accelerate marker-assisted selection, QTL mapping, and ultimately CRISPR-based metabolic engineering.
- (4) Development and farmer-level adoption of practical, low-cost post-harvest protocols combining cold storage, STS pulsing, and MAP.
- (5) Strengthening of farmer producer organizations (FPOs), cooperative processing units, and direct industry linkages.
- (6) Development of Geographic Indication (GI) tags for champaca flowers and absolutes from recognized quality zones in India.
- (7) Integration of precision agriculture tools such as remote sensing, soil health monitoring, and precision irrigation into commercial plantations.
- (8) Climate change vulnerability assessment and development of climate-resilient cultivars and management protocols.

11. CONCLUSION

This review has synthesized the current state of scientific knowledge on *Magnolia champaca* across agronomy, floral biology, crop improvement, post-harvest management, value addition, climate change vulnerability, and industry prospects. Three overarching conclusions emerge from this synthesis.

First, *M. champaca* is a crop with genuinely exceptional economic and cultural value combining a premier market position in the global natural perfumery sector (champaca absolute at USD 800–3,000+/kg), a massive domestic religious and ornamental flower market in South Asia, and significant pharmaceutical, food, and agroforestry applications. No other floriculture tree species combines this breadth of value-adding opportunities with such deep cultural embeddedness across South and Southeast Asia.

Second, the species is profoundly underserved by scientific research relative to its commercial potential. The absence of officially released improved varieties, inadequate post-harvest infrastructure, near-total absence of molecular breeding tools, and no reference genome are striking gaps for a crop of this economic importance. The contrast with comparable aromatic crops jasmine, vetiver, sandalwood where systematic R&D investment has produced measurable productivity and quality gains, underscores the opportunity cost of the current research deficit.

Third, emerging tools reference genome sequencing, CRISPR-based metabolic engineering of floral volatile biosynthesis, marker-assisted selection for fragrance quality, and precision agriculture are now sufficiently mature to be directly applied to *M. champaca* improvement with realistic near-term payoffs. A coordinated, multi-institutional R&D initiative, analogous to the successful champaca absolute value chain development programs in Indonesia, could transform the economic position of Indian champaca growers and processors within a decade.

It is hoped that this comprehensive review will serve as a foundational reference and catalyst for the scientific community, horticulture industry, funding agencies, and policy stakeholders committed to the sustainable development and commercialization of this extraordinary flowering tree.

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