

MULTI-OMICS APPROACHES TO MAPPING THE GENE REGULATORY NETWORKS BEHIND ANTHOCYANIN AND NUTRIENT ACCUMULATION IN FRUITS AND VEGETABLES: A COMPREHENSIVE REVIEW

Naval Kishore Meena¹, Dr. V. Madan Mohan Rao², Durgashankar Meena³, Miss Bhawna Panda⁴, Dr. J H Kahodariya⁵, Dr. Shashi Bala⁶, Dr. Shivaraj Kumar Verma⁷, Dr. Sunil Singh⁸, Dr. Ankit Singh⁹, Dr. Saudamini Swain¹⁰, Prince Kumar¹¹

¹Ph.D. Scholar, Department of Horticulture (Fruit Science), Rajasthan College of Agriculture, MPUAT, Udaipur-313001, Rajasthan.

²Head of Department, Department of Botany & Horticulture, Government Degree College, Chintalapudi, Eluru, Dist 534460. (Adikavi Nannayya University, Rajahmahendravaram, Ap).

³Technical Assistant, Department of Fruit Science, Dr BRC Agricultural Research Station, Mandor (Agriculture University, Jodhpur) Pincode 342304.

⁴Assistant Professor, Department of Fruit Science, MGUVV- College of Horticulture and Research Station, Raigarh (Chhattisgarh) Pin 496001. (Mahatma Gandhi Udyanikee Evam Vanikee Vishwavidyalaya, Durg (C.G.).

⁵Deputy Project Director- ATMA, Botad, Department of Agriculture, Farmers Welfare and Co-Operation, Govt. of Gujarat.

⁶Professor, Department of Horticulture, Udai Pratap Autonomous College, Varanasi.

⁷Assistant Professor, Department of Horticulture, Udai Pratap Autonomous College, Varanasi-221002, Uttar Pradesh, India.

⁸Associate Professor, Department of Botany, Udai Pratap Autonomous College, Varanasi.

⁹Assistant Professor, Department of Horticulture, Udai Pratap Autonomous College, Varanasi.

¹⁰Associate Professor and Scientist in Charge, AICRP ON FRUITS, OUAT, Bhubaneswar, Odisha.

¹¹Ph.D Research Scholar, Department of Vegetable and Spice Crops, Uttar Banga Krishi Viswavidyalaya, Pundibari, Cooch behar, WB.

*Corresponding Author email: navalkishor022018@gmail.com

ABSTRACT

Quality of fruit and vegetables is mainly determined by two traits: anthocyanin pigmentation and mineral micronutrient content. Both influence consumer attraction, antioxidant property and food security, but neither is controlled by a few genes independently. Instead, they each rely on complex gene regulatory networks (GRNs) integrating developmental, hormonal and environmental signals at transcriptional, post-transcriptional and epigenetic levels. Many years of gene-by-gene studies have revealed the key structural and regulatory genes, particularly the MYB-bHLH-WD40 (MBW) complex, which control anthocyanin biosynthesis, but these studies rarely linked upstream signalling to chromatin status, transcription, protein levels and metabolite outputs in a single system. That is starting to change now that high-throughput, multi-omics platforms are less expensive and more widely available. Genome-wide association studies (GWAS), bulk and single-cell and spatial transcriptomics, epigenomic profiling (WGBS, ChIP-seq, ATAC-seq), proteomics and metabolomics/ionomics in combination with statistical and machine-learning approaches e.g. weighted gene co-expression network analysis (WGCNA) and Bayesian network inference, allow one now to reconstruct GRNs at a resolution not available even 10 years ago and for an increasing number of plant species. This review brings together current knowledge of the anthocyanin biosynthetic pathway and its MBW-centred transcriptional logic; summarizes the major omics technologies and GRN reconstruction pipelines applied in fruit and vegetable crops; provides comparative case studies for apple, grape, citrus, strawberry, blueberry, tomato, eggplant, pepper, potato, sweet potato, lettuce, radish and other horticultural species; and adds a section on ionomics and iron, zinc and provitamin A biofortification. Epigenetic regulation, the role of environmental and hormonal signals, the application of GRN knowledge for precision breeding (e.g., CRISPR/Cas editing), and the health relevance of anthocyanin-rich commodities are also discussed.

KEYWORDS: anthocyanin biosynthesis; gene regulatory network; multi-omics integration; MYB-bHLH-WD40 complex; transcriptomics; metabolomics; ionomics; biofortification; fruit quality; CRISPR/Cas genome editing

1. INTRODUCTION

Fruits and vegetables contain the highest levels of vitamins, fibre and health-promoting phytochemicals in our diet, and anthocyanins contribute significantly to that value, playing the dual role of defining product colour and serving as the visual attribute influencing nutraceutical quality. These water-soluble flavonoid pigments are present in almost all types of fruits and vegetables such as apples, grapes, oranges, strawberries, blueberries, eggplant, purple cauliflower, radish, potato and sweet potato and their consumption have been linked to a reduction in the risk of CVD, obesity, type 2 diabetes, and various cancers (Khoo *et al.* 2017; Li *et al.* 2017; Manolescu *et al.*, 2019; Noman *et al.*, 2025). Individual genes less frequently control either anthocyanin biosynthesis or nutrient accumulation; instead, both characteristics are a product of GRNs that include transcription factor proteins, signalling kinases, chromatin modifiers, small RNAs and metabolic enzymes

The MBW complex also regulates the majority of the transcriptional regulation at this step. R2R3-MYB transcription factors confer target-gene and tissue specificities, bHLH proteins serve as obligate/facultative MYB partners to enhance MYB binding or elevate transactivation, and WD40-repeat proteins can't bind DNA directly but scaffold the complex and enhance its stability (Xu *et al.*, 2015; Feller *et al.*, 2011; Zhao *et al.*, 2019). A group of activating MYBs-MdMYB10/MdMYB1 in apple, VvMYBA1/VvMYBA2 in grape, FaMYB10 in strawberry, Ruby1 in citrus, and the PAP1/MYB113/MYB114 orthologues in Arabidopsis and relatives appear to mainly switch on late genes such as DFR, ANS and UFGT as opposed to the early ones, which is part of the explanation for why colour phenotypes are so often unlinked to total flavonoid flux (Espley *et al.*, 2007; Kobayashi *et al.*, 2002; Lin-Wang *et al.*, 2010; Butelli *et al.*, 2012). Repressive MYBs like MdMYB6 and FaMYB6-like and TgMYB4 which counteract the activation, whether by directly binding late-gene-promoters or through haplotype competition for the bHLH partners the activators require; the outcome on the whole is a RNA rheostat, fine-tuning the pigment intensity (Naing and Kim, 2018).

The MBW complex does not constitute the entire regulation. Several other regulators, such as WRKY, NAC, bZIPs (predominantly ELONGATED HYPOCOTYL5, HY5), B-box (BBX), RAV, HD-Zip, and SPL transcription factors, have been shown to direct light, temperature, sugar, and hormone signals into the MBW complex, either by binding to the promoters of structural or MYB-genes directly, or by physically associating with components of the MBW complex (Fang *et al.*, 2019; Yao *et al.*, 2017; Zhou *et al.*, 2015; Gou *et al.*, 2011). In strawberry the RAV-family regulator FaRAV1 directly activates FaMYB10 and its targets (Jiang *et al.*, 2023), while in the crosstalk between the HD-Zip protein FaANL2 and the repressor FaMYB6-like, they are antagonists in modulating pigment accumulation in ripening fruit. The regulation of canopy light exposure to fruit-skin colour via light-responsive BBX proteins that are upstream of HY5 and activate MYB10/MYB114 transcription has also been shown in apple and pear (Alabd *et al.*, 2022a, 2022b). A further post resource also plays a role: in apple, phosphorylation by MPK6 of MdHY5 enhances its DNA-binding ability, a sign that these GRNs are indeed active in both the transcriptional and post-translational level (Zhao and Han, 2023).

Table 1. Representative transcription factors and regulatory hubs governing anthocyanin gene regulatory networks across major fruit and vegetable crops.

Species	Key regulatory gene(s)	Regulatory role	Reference
Apple (<i>Malus domestica</i>)	MdMYB10/MdMYB1, MdMYB6, MdBBX21/22, MdHY5	MYB10 activates late genes; MdMYB6 represses via ANS/GSTF12; BBX-HY5 mediates light induction	Espley <i>et al.</i> , 2007; Chagne <i>et al.</i> , 2013
Grape (<i>Vitis vinifera</i>)	VvMYBA1/A2, VvMYBPA1, VvWRKY44/40	MYBA1/A2 control skin-specific UFGT expression; retrotransposon (Gret1) silences white berries	Kobayashi <i>et al.</i> , 2004; Bogs <i>et al.</i> , 2007
Citrus (<i>Citrus</i> spp.)	Ruby1 (<i>CsRuby1</i>)	Cold/light-induced via Tcs1 retrotransposon promoter; activated by CBF/ERF factors	Butelli <i>et al.</i> , 2012; Wang <i>et al.</i> , 2024
Strawberry (<i>Fragaria</i> spp.)	FaMYB10, FaMYB1, FaRAV1, FaMYB6-like	MYB10 is master activator; RAV1 upstream activator; MYB6-like/MYB1 repressors	Medina-Puche <i>et al.</i> , 2014; Jiang <i>et al.</i> , 2023
Blueberry (<i>Vaccinium</i> spp.)	VcMYBL1, VcbHLH1, VcWDL2, MYBPA-type	MBW complex regulated developmentally; MADS-box SQUAMOSA gene contributes	Zhao <i>et al.</i> , 2019; Jaakola <i>et al.</i> , 2010
Tomato (<i>Solanum lycopersicum</i>)	SIAN2/SIAN2-like, SIMYB75	Alternative splicing controls anthocyanin-free phenotype; engineered for fruit-wide pigmentation	Colanero <i>et al.</i> , 2019; Butelli <i>et al.</i> , 2008
Eggplant (<i>Solanum melongena</i>)	MYB113-like, SmWRKY44, COP1	GWAS-identified MYB/COP1 loci; WRKY44 as co-expression hub	Cericola <i>et al.</i> , 2014
Potato (<i>Solanum tuberosum</i>)	StAN1, StbHLH1, StWD40	AN1 is dominant regulatory gene of tuber/leaf anthocyanin	Strygina <i>et al.</i> , 2019
Radish (<i>Raphanus sativus</i>)	RsMYB1	Transposon-induced promoter methylation silences RsMYB1 in red-fleshed radish	Xie <i>et al.</i> , 2014
Lettuce (<i>Lactuca sativa</i>)	ANS-linked GWAS locus	GWAS-validated regulator of leaf anthocyanin used in genomic design breeding	Xie <i>et al.</i> , 2014

3. Multi-Omics Technologies for Gene Regulatory Network Reconstruction

Building a biologically meaningful GRN involves multiple lines of evidence DNA sequence variation, transcript abundance, chromatin state, protein levels, metabolite/ion concentration which must be integrated using appropriate statistical methodologies (Figure 2).

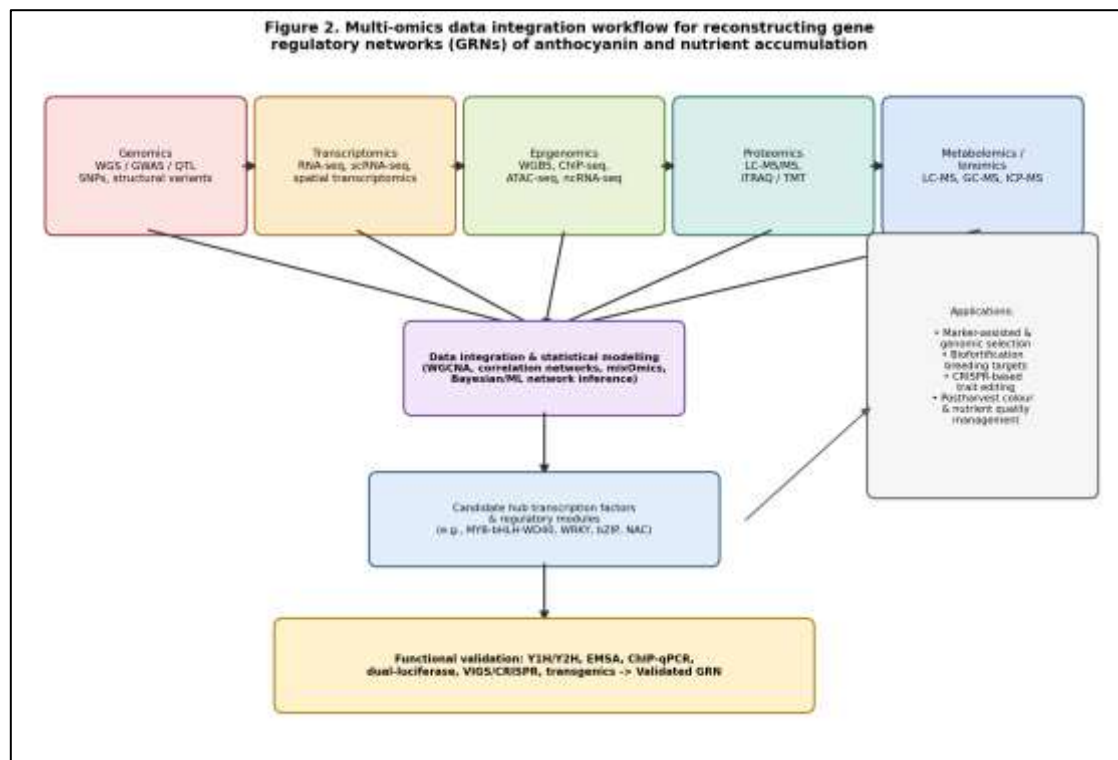


Figure 2. Working process from generation of multi-omics data to statistical integration and identification of hubs for functional validation and subsequent exploitation in breeding for the reconstruction of gene regulatory networks (GRNs) controlling anthocyanin and nutrient accumulation. (Schematic drawing, conceptual framework adapted from Acharjee *et al.*, 2016; Fondi and Lio, 2015; Langfelder and Horvath, 2008)

3.1 Genomics, GWAS and QTL Mapping

GWAS developed based on whole-genome resequencing and SNP arrays have identified a myriad of candidate anthocyanin loci, such as MYB113-like and COP1 in eggplant (Cericola *et al.*, 2014), MaVHAG3 in mulberry, and ANS-related loci uncovered in a wide lettuce panel for genomic-assisted breeding of high-anthocyanin cultivars. Candidate-gene association analyses in grape berries-based investigations in MYB11, MYBCC and MYC-B variants with skin-anthocyanin content (Cardoso *et al.*, 2012). A parallel mineral story, ionomic GWAS in maize, rice and barley has identified QTL for iron, zinc and cadmium accumulation coinciding with transporter and chelator genes, although only a small fraction of these loci currently coincides with functionally validated ionome genes a gap which underscores how much independent validation is still needed (Salt *et al.*, 2008; Baxter *et al.*, 2008).

3.2 Transcriptomics, Single-Cell and Spatial Profiling

Bulk RNA sequencing continues to be the method of choice for identifying differentially expressed genes among developmental stages, tissues and genotypes, and is the basis of much of what is understood of anthocyanin GRNs in apple, grape, citrus, strawberry and other species (Espley *et al.*, 2007; Wang *et al.*, 2024). Single-cell and spatial transcriptomics are beginning to uncover the cell-type heterogeneity that bulk methods inherently average over exposing staged ripening programmes along the tomato fruit's longitudinal axis and cell-specific regulatory states in floral and fruit tissue that were simply beyond the reach of previous techniques (Meir *et al.*, 2021). Single-cell multi-omics is still a new tool in horticultural species, but it will help to better localize which regulatory modules are active in which parts of a fruit or vegetable tissue.

3.3 Epigenomics: DNA Methylation, Histone Marks and Chromatin Accessibility

Whole-genome bisulfite sequencing confirms that hypermethylation at the MYB10/MYB1 promoter regions can fully suppress anthocyanin production, resulting in striped or non-red bud-sport types in both apple and pear, and that exposure to light induces global remodelling and demethylation at the promoters of anthocyanin genes (Bai *et al.*, 2017; El-Sharkawy *et al.*, 2015). The ChIP-seq and ATAC-seq provide a direct read-out of transcription-factor binding to DNA and accessibility of chromatin; in mulberry, for example, tandem analysis of ATAC-seq and RNA-seq at the fruit colour transition stage identified enrichment of AP2/ERF, MYB, NAC, LBD and WRKY motifs in open chromatin and constructed a transcriptional network that connects particular hub transcription factors to pigmentation genes (Buenrostro *et al.*, 2013; Zhang *et al.*, 2008).

3.4 Proteomics, Metabolomics and Ionomics

LC-MS/MS-based proteomics introduces a dimension not attainable by transcriptomics that enables the observation of post-transcriptional and post-translational regulation. Granger-causality analysis of time-course proteomic and metabolomic data in grape berries demonstrated that changes in protein levels can anticipate and foresee flux changes (Wang *et al.*, 2018), a case cited to further support the embedding of proteomic data in GRN

rather than treating it as an add-on. Metabolomic (LC-MS, GC-MS) and ICP-MS-based ionic analyses of mineral content capture the final biochemical phenotype, and support correlation-based retrodiction to upstream transcriptional modules (Alseekh *et al.*, 2021; Fernie *et al.*, 2004; Lahner *et al.*, 2003). The combination of such terminal-phenotype layers with transcriptomic and epigenomic data has been particularly valuable in multi-omics investigations of water-stressed grape berries (Savoi *et al.*, 2017), postharvest kiwifruit ripening, black raspberry ripening (where DNA methylation tracked directly to which anthocyanin species accumulated), and Jaboticaba fruit peel colour development, where WGCNA identified a large co-expression module with six anthocyanin structural genes and 98 transcription factors.

4. Approaches to Gene Regulatory Network Inference and Functional Validation

Statistical and computational inference that converts correlative multi-omics data into testable regulatory hypotheses (Figure 3). WGCNA is still the most frequently employed approach in multi-omics studies applied to horticulture: genes are clustered into co-expression modules and each module's eigengene is correlated with phenotypic traits to identify candidate hub genes. The strategy has revealed hub regulators in the anthocyanin networks of maize pericarp, blood orange, *Coffea arabica* peel, chrysanthemum and eggplant, among other systems (Langfelder and Horvath, 2008).

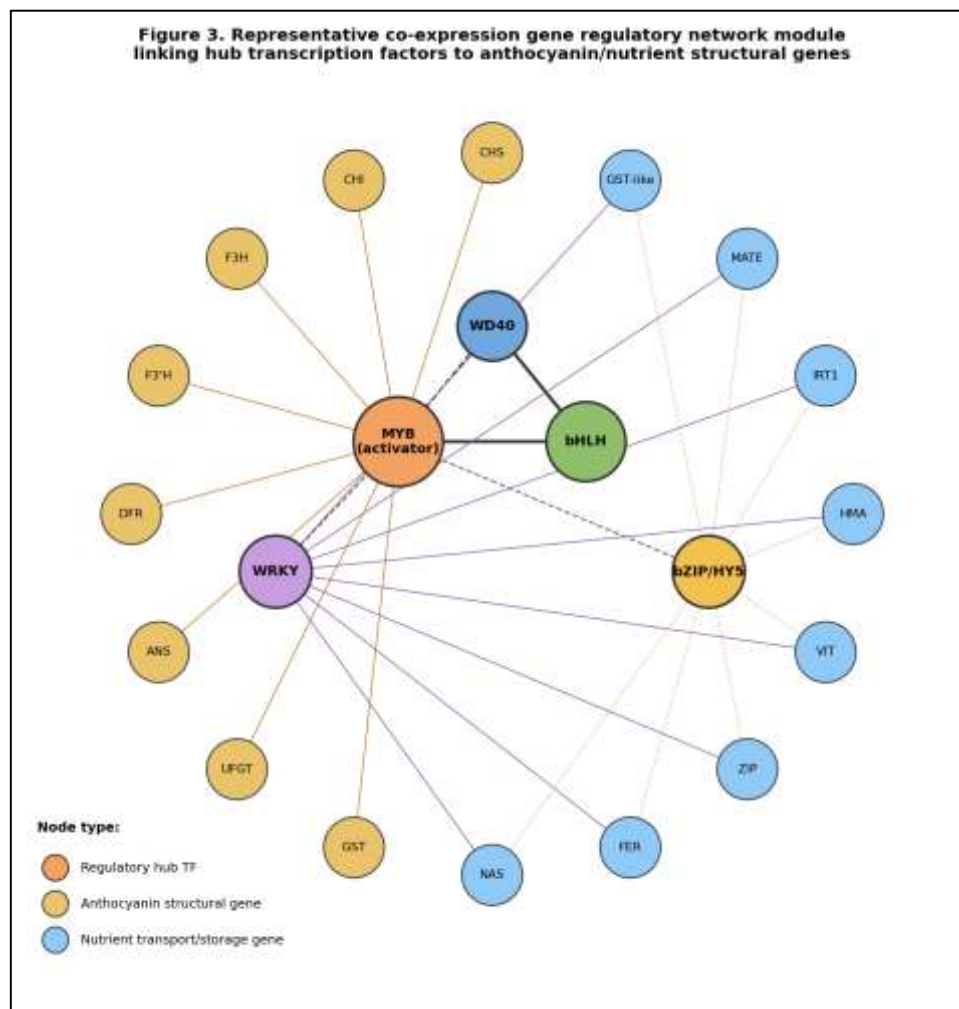


Figure 3. A representative co-expression gene regulatory network module linking hub transcription factors (MYB, bHLH, WD40, WRKY, bZIP/HY5) to anthocyanin structural genes and nutrient transport/storage genes, illustrating the hub-and-spoke topology typical of WGCNA-derived modules. (Original figure, adapted from the WGCNA framework of Langfelder and Horvath, 2008)

Besides WGCNA, inference methodologies that integrate chromatin accessibility superimposing ATAC-seq/ChIP-seq footprints onto expression data further focus the hunt for direct transcription factor-target interactions. Gradient-boosting and deep-learning frameworks developed for mammalian systems, e.g. SCING and maxATAC, are starting to be extended for GRN inference from single-cell and chromatin-accessibility data, and they tend to outperform pure correlation-based methods at comparable scale. However, regardless of the inference method, predictive regulatory edges must still be confirmed experimentally. That validation usually involves yeast-one hybrid (Y1H) and two hybrid (Y2H) assays, EMSA, ChIP-qPCR, dual-luciferase reporter assays, virus induced gene silencing (VIGS), as well as stable or transient overexpression or CRISPR-based knock out performed in the species under investigation or in heterologous systems like tobacco and Arabidopsis, as

shown for functional verification of MdMYB66-MdF3H, FaRAV1-FaMYB10 and AtrANS-anthocyanin pathway interactions.

5. Comparative Case Studies in Fruit Crops

5.1 Pome and Stone Fruits

In apple, transcriptomic data combined with genome resequencing identified a mini-satellite repeat located upstream of MdMYB10 that mediates the gene's own autoregulation and is the molecular basis for red-fleshed phenotypes (Espley *et al.*, 2009; Chagne *et al.*, 2013; Zhang *et al.*, 2019). Both ripening-associated repressors MdMYB6 and MdMYB16 can bind to the promoters of sugar-transport genes including MdTMT1, which provides a connection between anthocyanin accumulation and soluble-sugar metabolism.

5.2 Grape and Small/Berry Fruits

The colour of a grape berry is largely determined by the VvMYBA1/VvMYBA2 locus, which is disrupted in white cultivars by the insertion of the Gret1 retrotransposon (Kobayashi *et al.*, 2004), and the proanthocyanidin and the flavonol branches are controlled independently by VvMYBPA1/PA2 and VvMYBF1 (Bogs *et al.*, 2007). To our knowledge, this is the most comprehensive study combining transcriptomics and targeted metabolomics in water-deficit. The VvMYB15-VvWRKY40 and VvMYBA1-VvWRKY44-VvMYB308 modules provide a striking example of how WRKY proteins can further mediate stress and developmental signals through the core MBW module. Blueberry features a similar MBW complex VcMYBL1, VcbHLH1, VcWDL2 along with a SQUAMOSA MADS-box regulator that controls developmentally regulated anthocyanin accumulation (Zhao *et al.*, 2019; Jaakola *et al.*, 2010). WGCNA-based hub analysis has brought out similar signatures in Aronia melanocarpa (AmMYB10) and mulberry, where a haplotype-resolved genome assembly combined with GWAS pinpointed a vacuolar H⁺-ATPase subunit gene (MaVHAG3) associated with purple pigmentation, and where ATAC-seq/RNA-seq integrative approaches unveiled the chromatin-level network operating during the colour turning stage.

5.3 Strawberry

Tractable transient transformation systems in strawberry have rendered it a model species for the analysis of anthocyanin GRNs. FaMYB10 is the master activator, triggered independently by light and by ABA signaling through the FaNCED1/FaCHLH-mediated ABA biosynthesis and perception (Aharoni *et al.*, 2001; Medina-Puche *et al.*, 2014). FaRAV1 activates FaMYB10 and its biosynthetic-gene promoters directly (Jiang *et al.*, 2023); a complex accessory network modulates it: FaMYB6-like and FaMYB44.2 act as feedback repressors and link anthocyanin accumulation to sucrose metabolism as well by forming complexes with FabHLH3 and binding to the promoters of sucrose-phosphate-synthase; and FaANL2, a HD-Zip protein, represses FaMYB10 in a light- and hormone-dependent way. Homoeolog-specific CRISPR/Cas9-mediated mutagenesis of MYB10-1B in octoploid strawberry has provided such direct causal evidence by producing white-fruited lines with the co-suppression of CHS, DFR and ANS.

5.4 Citrus

Pigmented citrus-like blood orange results from independent insertions of a Copia retrotransposon upstream of the Ruby1 MYB gene that drive cold-dependant transcriptional activation (Butelli *et al.*, 2012; Huang *et al.*, 2018, 2019). Recent data demonstrate that cold-induced ethylene response factors also directly activate Ruby1 through this retrotransposon-derived promoter, thereby integrating temperature sensing into the citrus anthocyanin GRN (Wang *et al.*, 2024). The fruit specific beta-cyclase 2 gene has also been targeted by CRISPR/Cas9 editing to stack anthocyanin and lycopene pigmentation within a single variety.

5.5 Tomato and Other Solanaceous Fruits

Alternative splicing of the cryptic intron I2 in the R2R3-MYB gene SIAN2-like is the reason why cultivated tomato fruit naturally lacks anthocyanins (Colanero *et al.*, 2019). By introgressing wild Solanum alleles, or ectopically expressing the snapdragon-derived Delila and Rosea1-type MYB/bHLH factors, purple tomato enriched in anthocyanin with enhanced antioxidant capacity was successfully obtained (Butelli *et al.*, 2008; Zhang *et al.*, 2015). Ripe tomato fruit colour is also, independently, determined by a carotenoid mainly lycopene that is regulated by a hierarchical transcriptional network mediated by SIAP2a/SIAP2c, EIN3/EIL and ethylene-responsive factors regulating histone acetylation at lycopene-pathway genes. This is a good reminder that GRNs for fruit colour frequently may show interplay between the flavonoid and carotenoid pathways of secondary metabolism (Duduit *et al.*, 2022).

6. Comparative Case Studies in Vegetable Crops

Vegetable crops are subject to the same MBW-containing regulatory logic described above, but they frequently execute it in root or non-fruit tissues, demanding tissue-specific regulatory adaptations.

Table 2. Multi-omics case studies of anthocyanin gene regulatory networks in representative vegetable and vegetable-fruit crops.

Crop	Tissue studied	Omics approach(es)	Key regulatory finding
Eggplant (Solanum melongena)	Fruit epidermis, vegetative tissue	GWAS (MAGIC population), LD/association mapping	MYB113-like and COP1 loci associated with pigmentation; SmWRKY44 identified as WGCNA hub gene

Crop	Tissue studied	Omics approach(es)	Key regulatory finding
Pepper (<i>Capsicum</i> spp.)	Fruit	GWAS, QTL mapping	A-locus MYB linked with fs10.1 fruit-shape QTL; candidate genes for agronomic/pigment traits
Potato (<i>Solanum tuberosum</i>)	Tuber, leaf	Transcriptomics, metabolomics, TF overexpression	StAN1 (with StbHLH1/StWD40) is master regulator; miR828 associated with skin/flesh colour
Sweet potato (<i>Ipomoea batatas</i>)	Tuberous root	Comparative transcriptomics	IbMYB1/IbbHLH2/IbWRKY activators counteracted by IbMYB27/IbMYBx-ZZ repressors
Radish (<i>Raphanus sativus</i>)	Taproot	Transcriptomics, epigenetics	Transposon-induced RsMYB1 promoter methylation silences red-flesh phenotype
Lettuce (<i>Lactuca sativa</i>)	Leaf	Whole-genome resequencing, GWAS (811 accessions)	ANS-linked QTL validated and deployed in genomic design breeding of high-anthocyanin cultivar
Purple cauliflower/Brassica (<i>Brassica</i> spp.)	Curd/stem tissue	Transcriptomics, biochemical profiling	MYB-dependent activation of anthocyanin pathway in purple tumorous stem mustard
Coffee (<i>Coffea arabica</i>)	Fruit peel	Metabolomics + transcriptomics + WGCNA	Structural genes (3AT, BZ1, lcyE) and MYB/NAC/bHLH TFs co-regulate anthocyanin and carotenoid pigmentation

A common theme in these vegetable systems is the presence of MBW-equivalent activator complexes AN1/StAN1, IbMYB1, RsMYB1, SmMYB113-like which are negatively regulated by tissue or condition-specific repressor MYBs and further influenced by transposable-element insertions and promoter methylation. This suggests that epigenetic silencing is a highly conserved mechanism for regulating anthocyanin GRN output in a manner independent of coding-sequence variation.

7. Multi-Omics Insights into Nutrient (Mineral) Accumulation and Biofortification

The ionome- the full complement of mineral and trace elements found within a plant tissue can now be measured at high throughput using ICP-MS, and can be used for GWAS and QTL mapping of traits related to elemental accumulation similar to how anthocyanin pigmentation traits have been mapped (Lahner *et al.*, 2003; Baxter *et al.*, 2008; Salt *et al.*, 2008). Transport and chelation networks of iron and zinc are controlled by a complex of genes encoding for nicotianamine synthase (NAS), ferritin (FER), ZIP-family zinc transporters, vacuolar iron transporters (VIT), heavy-metal ATPases (HMA), and IRT1-type iron-regulated transporters whose expression is coordinately regulated to control the efficiency with which minerals are absorbed, transported via the xylem/phloem, and deposited into edible tissue (Connorton and Balk, 2019; Connorton *et al.*, 2017; Bashir *et al.*, 2011; Sperotto *et al.*, 2012).

This GRN-level knowledge of transporter networks has nourished agronomic, conventional-breeding and transgenic strategies to biofortification alike. Overexpression of NAS and ferritin genes has also increased the levels of iron and zinc in potato tubers and rice endosperm, and GWAS loci co-locating with known ionome-gene orthologues has fast tracked marker assisted selection for high-iron and high-zinc cultivars in rice, wheat and maize (White and Broadley, 2009; Bouis and Saltzman, 2017). Work in maize with multi-omics approaches has demonstrated that phosphorus fertiliser dramatically alters the whole-plant ionome, with genotype-by-environment interactions having a strong effect on the efficiency of zinc translocation yet another indication that nutrient GRNs, just like anthocyanin GRNs, are plastically responding to the environment conditions rather than behaving as fixed circuits.

Table 3. Representative gene network nodes linking multi-omics evidence to mineral nutrient accumulation and biofortification traits.

Nutrient/trait	Key gene(s)/network node(s)	Omics evidence	Representative reference
Iron accumulation	NAS, FER, IRT1, VIT, HMA	GWAS, ionomics (ICP-MS), transgenics	Connorton and Balk, 2019; Bashir <i>et al.</i> , 2011
Zinc accumulation	ZIP transporters, NAS	Ionomic GWAS, QTL mapping	White and Broadley, 2009; Salt <i>et al.</i> , 2008
Provitamin A (carotenoids)	PSY, PDS, LCYB/LCYE, CYP97 family	Transcriptomics, targeted metabolomics	Duduit <i>et al.</i> , 2022; Zhang <i>et al.</i> , 2015
Anthocyanin-nutrient co-regulation	MYB-HY5-ABA cross-talk nodes	Multi-omics (transcriptome + metabolome + ionome)	Savoi <i>et al.</i> , 2017

8. Epigenetic and Environmental Integration into Anthocyanin and Nutrient GRNs

DNA methylation, histone modification and small RNA-mediated silencing (Figure 2) are now considered to be basic elements rather than a peripheral superstructure of anthocyanin GRNs. Promoters of MYB10/MYB1 are hypermethylated in striped and bud-sport and yellow fruit phenotypes in apple and pear, respectively, while genome-wide dynamic RNA-directed DNA methylation (RdDM) and active demethylation reprogram the methylome in response to light mediating the expression of PyUGT, PyDFR, and other structural genes even the MYB10 promoter itself is not the target of this process (Bai *et al.*, 2017; El-Sharkawy *et al.*, 2015). In black raspberry, system wide methylation changes during ripening have been linked directly to which anthocyanin species accumulate through multi-omics profiling, and in radish a transposon insertion and subsequent promoter methylation of RsMYB1 completely shuts down the anthocyanin GRN, resulting in white-fleshed roots (Xie *et al.*, 2014).

Histone modifications including H3 acetylation and H3K4 trimethylation are associated with transcriptional activation of MdMYB1 alleles in response to light and temperature signals that induce pigmentation, whereas SPL transcription factors regulated by microRNA miR156 destabilize the MBW complex, reducing anthocyanin biosynthesis during floral and fruit maturation (Gou *et al.*, 2011). Taken together, these results suggest that the GRNs controlling the biosynthesis of anthocyanins, and by extension nutrients, are not hardwired. These are circuits that are constantly being reshaped by light quality and duration, ambient and diurnal temperature, water and nutrient status, sugar signalling and developmental stage variables that any multi-omics GRN model seeking to predict breeding outcomes will have to explicitly model.

9. Genome Editing and Biotechnological Translation of GRN Knowledge

CRISPR/Cas-based genome editing has now become the main strategy for testing GRN hypotheses and for directly engineering traits in horticultural crops. Targeted disruption of FveMYB10 and FveCHS in diploid and octoploid strawberry resulted in predictable reductions in anthocyanin levels, and homeolog-specific editing of MYB10-1B resulted in white-fruited strawberry lines with coordinated down-regulation of CHS, DFR and ANS the first direct causal evidence for the FaMYB10-centred network. In citrus, dual-sgRNA CRISPR/Cas9 editing of the fruit-specific beta-cyclase 2 gene have been exploited to generate varieties that produce both anthocyanin and lycopene pigmentation, a trait that could not be produced through conventional breeding due to sexual incompatibility between the candidate parental genotypes.

In addition to basic loss-of-function editing, CRISPR activation systems such as CRISPR-Act3.0 enable multiplexed and dosage-dependent up-regulation of anthocyanin and other secondary-metabolites pathway genes as demonstrated, e.g., in pear calli providing a gain-of-function approach to biofortification that is complementary to traditional overexpression transgenics. Anthocyanin-marker aided CRISPR (AAC), which integrates pathway activation with a CRISPR/Cas9 cassette as a visible transformation marker, has facilitated the obtainment of transgene-free edited rice plants, a more practical benefit from knowledge on anthocyanin GRN (He *et al.*, 2020). On the nutrition front, transgenic co-expression of the nicotianamine synthase and ferritin genes has been used to accomplish iron and zinc biofortification in potato tubers concurrently, suggesting that GRN-based multigene stacking can surpass single-gene approaches for intricate nutrient traits.

10. Nutritional and Health Significance of Anthocyanin- and Nutrient-Enriched Produce

The case for engineering anthocyanin and nutrient-rich produce is not only based on agronomy or molecular biology, but also supported by a large body of evidence from nutritional and clinical studies. Anthocyanins have antioxidant, anti-inflammatory, and anti-apoptotic properties relevant to the prevention and treatment of cardiovascular disease, obesity, type-2 diabetes, and some cancers, and epidemiological studies have reported an association between higher anthocyanin intake and improved cardiometabolic and cognitive outcomes (Li *et al.*, 2017; Khoo *et al.*, 2017; Manolescu *et al.*, 2019). Clinical and preclinical studies have additionally shown that anthocyanins and their gut-microbial metabolites affect inflammatory markers, lipid profiles, and oxidative-stress biomarkers, and that the bioavailability and metabolism of anthocyanins are influenced by the glycosylation and acylation pattern of the parent anthocyanidin (Noman *et al.*, 2025).

Iron, zinc and provitamin A deficiencies are still among the most prevalent forms of malnutrition globally, affecting disproportionately women in the reproductive age and young children residing in low- and middle-income countries. Iron- and zinc-biofortified crops produced through conventional breeding and transgenic approaches as well as agronomic biofortification have demonstrated population-level composition and efficacy in biofortified crops for iron and zinc is proof that multi-omics-aided mining of GRN hub genes for nutrient transport and accumulation has real, translatable public-health value that surpasses basic science (Bouis and Saltzman, 2017; Vasconcelos *et al.*, 2017).

11. Challenges and Future Perspectives

Several theoretical and technical challenges remain to bridge between the current state of the field and the stage of routine multi-omics GRN reconstruction in horticultural breeding. The first is data heterogeneity: platform-related biases within genomics, transcriptomics, proteomics and metabolomics make it challenging to combine data in a statistically meaningful way, and standardized pipelines tailored for horticultural crops are much less advanced than those for model species or human disease. Approximately 1/3rd of GRNs published are causal as

opposed to correlative WGCNA and similar co-expression approaches do identify statistically associated hub genes, but they cannot distinguish direct regulatory relationships from indirect, co-regulated, or ‘bystander’ ones. This means validation downstream via Y1H/Y2H, ChIP and CRISPR-based perturbation is an unavoidable bottleneck, “rate-limiting step” in their parlance. Third, the single-cell and spatial multi-omics technologies that are truly transformative in model organisms are currently challenging and expensive to implement in the polyploid, heterozygous, and frequently poorly annotated genomes that define most horticultural crops for example the octoploid strawberry and hexaploid sweet potato, as well as many clonally propagated fruit species. A fourth caveat is that the majority of GRN analyses so far have focused on a single omics layer, or at best a double omics layer transcriptome-metabolome dataset. Truly integrative approaches at the level of genomic variation, chromatin accessibility, transcriptome, proteome and metabolome/ionome data integrated in a single statistical framework are still not very common, and precisely such type of integration would be needed to dissect causal hierarchies and cross-talk between the anthocyanin and nutrient-transport networks that were outlined during this review. Fifth, the pathway from validated GRN hub genes to elite cultivars still relies on robust, genotype-independent transformation and regeneration methodologies a bottleneck that remains for many woody perennial fruit crops.

Looking ahead, several trends are expected to drive advancement of the field collectively: the adoption of machine-learning and deep-learning GRN inference methods, which can integrate a variety of heterogeneous, sparse and multimodal omics data; the development of single-cell and spatial multi-omics atlases for staple fruits and vegetable crops; integration of AI-assisted variant-to-function prediction with GWAS and pangenome resources to bridge the gap from statistical association to validated causal genes; and closer linkage of GRN-discovery pipelines with CRISPR/Cas-based functional validation and precision breeding, such that candidate hub genes from multi-omics integration will more rapidly transition into elite breeding lines.

12. CONCLUSION

The use of multi-omics integration has transformed the investigation of anthocyanin and nutrient accumulation in fruits and vegetables from a gene-by-gene approach into a systems-level science that can elucidate the complex interplay between the core MYB-bHLH-WD40 (MBW) complex and its downstream targets, the chromatin and epigenetic states that regulate their activity, and the environmental and hormonal signals that modulate GRN output across development and stress. The comparative case studies presented here from apple, grape, citrus, strawberry, blueberry, tomato, eggplant, potato, sweet potato, lettuce and other crops reveal both a high degree of conservation of the core MBW architecture and significant species- and tissue-specific diversification, driven by the recruitment of accessory transcription factors, transposable-element-mediated promoter evolution, and epigenetic silencing. The same multi-omics and network-inference frameworks are now beginning to produce similarly high-resolution pictures of the transporter and chelator networks involved in iron, zinc, and provitamin A accumulation in the context of ionomics and biofortification. Future investment in integrative, causally validated, and experimentally testable GRN models complemented by CRISPR/Cas-based functional validation and precision breeding shows real potential for systematic improvement of colour, flavour and nutritional quality in horticultural crops on a global scale.

REFERENCES

1. Acharjee A, Kloosterman B, Visser RGF, Maliapaard C (2016). Integration of multi-omics data for prediction of phenotypic traits using random forest. *BMC Bioinformatics*, 17(Suppl 5), 180.
2. Aharoni A, Ric De Vos CH, Wein M, Sun Z, Greco R, Kroon A, Mol JN, O'Connell AP (2001). The strawberry FaMYB1 transcription factor suppresses anthocyanin and flavonol accumulation in transgenic tobacco. *Plant Journal*, 28(3), 319-332.
3. Akagi T, Suzuki Y, Ikegami A, Yonemori K (2010). MBW complexes impinge on anthocyanidin reductase gene regulation for proanthocyanidin biosynthesis in persimmon fruit. *Plant Physiology*, 158(2), 1042-1054.
4. Alabd A, Ahmad M, Zhang X, Gao Y, Peng L, Zhang S, Wu J (2022a). Unravelling the pear fruit anthocyanin biosynthesis regulatory network via the BBX-HY5 module. *Postharvest Biology and Technology*, 194, 112103.
5. Alabd A, Ni J, Xin H, Xu K, Zhang Y, Fu X, Wang Y, Bai S, Teng Y (2022b). BBX proteins in pear are involved in the regulation of anthocyanin biosynthesis. *Plant Physiology and Biochemistry*, 174, 10-22.
6. Allan AC, Hellens RP, Laing WA (2008). MYB transcription factors that colour our fruit. *Trends in Plant Science*, 13(3), 99-102.
7. Alseekh S, Aharoni A, Brotman Y, Contrepolis K, D'Auria J, Ewald J, Fraser PD, Giavalisco P, Hall RD, Heinemann M, *et al* (2021). Mass spectrometry-based metabolomics: a guide for annotation, quantification and best reporting practices. *Nature Methods*, 18(7), 747-756.
8. Azuma A, Yakushiji H, Koshita Y, Kobayashi S (2012). Flavonoid biosynthesis-related genes in grape skin are differentially regulated by temperature and light conditions. *Planta*, 236(4), 1067-1080.
9. Bai S, Tuan PA, Saito T, Honda C, Hatsuyama Y, Ban Y, Moriguchi T (2017). Epigenetic regulation of MdMYB1 is associated with paper bagging-induced red pigmentation of apples. *Planta*, 246(4), 659-671.
10. Bashir K, Ishimaru Y, Shimo H, Nagasaka S, Fujimoto M, Takanashi H, Tsutsumi N, An G, Nakanishi H, Nishizawa NK (2011). The rice mitochondrial iron transporter is essential for plant growth. *Nature Communications*, 2, 322.

11. Baxter I, Vitek O, Lahner B, Muthukumar B, Borghi M, Morrissey J, Guerinot ML, Salt DE (2008). The leaf ionome as a multivariable system to detect a plant's physiological status. *Proceedings of the National Academy of Sciences USA*, 105(33), 12081-12086.
12. Bogs J, Ebadi A, McDavid D, Robinson SP (2007). Identification of the flavonoid hydroxylases from grapevine and their regulation during fruit development. *Plant Physiology*, 143(3), 1347-1361.
13. Bouis HE, Saltzman A (2017). Improving nutrition through biofortification: a review of evidence from HarvestPlus, 2003 through 2016. *Global Food Security*, 12, 49-58.
14. Buenrostro JD, Giresi PG, Zaba LC, Chang HY, Greenleaf WJ (2013). Transposition of native chromatin for fast and sensitive epigenomic profiling of open chromatin, DNA-binding proteins and nucleosome position. *Nature Methods*, 10(12), 1213-1218.
15. Butelli E, Licciardello C, Zhang Y, Liu J, Mackay S, Bailey P, Reforgiato-Recupero G, Martin C (2012). Retrotransposons control fruit-specific, cold-dependent accumulation of anthocyanins in blood oranges. *Plant Cell*, 24(3), 1242-1255.
16. Butelli E, Titta L, Giorgio M, Mock HP, Matros A, Peterek S, Schijlen EG, Hall RD, Bovy AG, Luo J, Martin C (2008). Enrichment of tomato fruit with health-promoting anthocyanins by expression of select transcription factors. *Nature Biotechnology*, 26(11), 1301-1308.
17. Cardoso S, Lau W, Eiras Dias JE, Fevereiro P, Maniatis N (2012). A candidate-gene association study for berry colour and anthocyanin content in *Vitis vinifera* L. *PLoS ONE*, 7(9), e46021.
18. Castillejo C, Waurich V, Wagner H, Ramos R, Oiza N, Munoz P, Trivino S, Caruana J, Liu Z, Cobo N, *et al* (2020). Allelic variation of MYB10 is the major force controlling natural variation in skin and flesh color in strawberry (*Fragaria spp.*) fruit. *Plant Cell*, 32(11), 3723-3749.
19. Cericola F, Portis E, Lanteri S, Toppino L, Barchi L, Acciarri N, Pulcini L, Sala T, Rotino GL (2014). Linkage disequilibrium and genome-wide association analysis for anthocyanin pigmentation and fruit color in eggplant. *BMC Genomics*, 15, 896.
20. Chagne D, Lin-Wang K, Espley RV, Volz RK, How NM, Rouse S, Brendolise C, Carlisle CM, Kumar S, De Silva N, *et al* (2013). An ancient duplication of apple MYB transcription factors is responsible for novel red fruit-flesh phenotypes. *Plant Physiology*, 161(1), 225-239.
21. Chaim AB, Borovsky Y, De Jong W, Paran I (2003). Linkage of the A locus for the presence of anthocyanin and fs10.1, a major fruit-shape QTL in pepper. *Theoretical and Applied Genetics*, 106(5), 889-894.
22. Chandrasekaran J, Brumin M, Wolf D, Leibman D, Klap C, Pearlsman M, Sherman A, Arazi T, Gal-On A (2016). Development of broad virus resistance in non-transgenic cucumber using CRISPR/Cas9 technology. *Molecular Plant Pathology*, 17(7), 1140-1153.
23. Cheng Y, Tian Y, Guo P, Luo J, Xu C, Zhang Y, Chen G, Xie Q, Hu Z (2024). Novel insights into pigment composition and molecular mechanisms governing flower coloration in rose cultivars exhibiting diverse petal hues. *Plants*, 13(23), 3353.
24. Colanero S, Perata P, Gonzali S (2019). Alternative splicing in the anthocyanin fruit gene encoding an R2R3 MYB transcription factor affects anthocyanin biosynthesis in tomato fruits. *Plant Physiology*, 182(4), 1965-1980.
25. Connorton JM, Balk J (2019). Iron biofortification of staple crops: lessons and challenges in plant genetics. *Plant and Cell Physiology*, 60(7), 1447-1456.
26. Connorton JM, Balk J, Rodriguez-Celma J (2017). Iron homeostasis in plants - a brief overview. *Metallomics*, 9(7), 813-823.
27. Duan S, Wang J, Gao C, Jin C, Li D, Peng D, Du G, Li Y, Chen M (2017). Functional characterization of a heterologously expressed *Brassica napus* WRKY41-1 transcription factor in regulating anthocyanin biosynthesis in *Arabidopsis thaliana*. *Plant Science*, 268, 47-53.
28. Duduit JR, Kosentka PZ, Miller MA, Blanco-Ulate B, Lenucci MS, Panthee DR, Perkins-Veazie P, Liu W (2022). Coordinated transcriptional regulation of the carotenoid biosynthesis contributes to fruit lycopene content in high-lycopene tomato genotypes. *Horticulture Research*, 9, uhac084.
29. El-Sharkawy I, Liang D, Xu K (2015). Transcriptome analysis of an apple (*Malus x domestica*) yellow fruit somatic mutation identifies a gene network module highly associated with anthocyanin and epigenetic regulation. *Journal of Experimental Botany*, 66(22), 7359-7376.
30. Espley RV, Hellens RP, Putterill J, Stevenson DE, Kutty-Amma S, Allan AC (2007). Red colouration in apple fruit is due to the activity of the MYB transcription factor, MdMYB10. *Plant Journal*, 49(3), 414-427.
31. Espley RV, Brendolise C, Chagne D, Kutty-Amma S, Green S, Volz R, Putterill J, Schouten HJ, Gardiner SE, Hellens RP, Allan AC (2009). Multiple repeats of a promoter segment causes transcription factor autoregulation in red apples. *Plant Cell*, 21(1), 168-183.
32. Fang H, Dong Y, Yue X, Hu J, Jiang S, Xu H, Wang Y, Su M, Zhang J, Zhang Z, *et al* (2019). The B-box zinc finger protein MdBBX20 integrates anthocyanin and chlorophyll biosynthesis in apple. *Plant Journal*, 100(6), 1101-1117.
33. Feller A, Machemer K, Braun EL, Grotewold E (2011). Evolutionary and comparative analysis of MYB and bHLH plant transcription factors. *Plant Journal*, 66(1), 94-116.
34. Fernie AR, Trethewey RN, Krotzky AJ, Willmitzer L (2004). Innovation - metabolite profiling: from diagnostics to systems biology. *Nature Reviews Molecular Cell Biology*, 5(9), 763-769.

35. Fischer TC, Mirbeth B, Rentsch J, Sutter C, Ring L, Flachowsky H, Habegger R, Hoffmann T, Hanke MV, Schwab W (2014). Premature and ectopic anthocyanin formation by silencing of anthocyanidin reductase in strawberry (*Fragaria x ananassa*). *New Phytologist*, 201(2), 440-451.
36. Fondi M, Lio P (2015). Multi-omics and metabolic modelling pipelines: challenges and tools for systems microbiology. *Microbiological Research*, 171, 52-64.
37. Frazee AC, Perte G, Jaffe AE, Langmead B, Salzberg SL, Leek JT (2015). Ballgown bridges the gap between transcriptome assembly and expression analysis. *Nature Biotechnology*, 33(3), 243-246.
38. Gao J, Wang G, Ma S, Xie X, Wu X, Zhang X, Wu Y, Zhao P, Xia Q (2015). CRISPR/Cas9-mediated targeted mutagenesis in *Nicotiana tabacum*. *Plant Molecular Biology*, 87(1-2), 99-110.
39. Gou JY, Felippes FF, Liu CJ, Weigel D, Wang JW (2011). Negative regulation of anthocyanin biosynthesis in *Arabidopsis* by a miR156-targeted SPL transcription factor. *Plant Cell*, 23(4), 1512-1522.
40. Grotewold E (2006). The genetics and biochemistry of floral pigments. *Annual Review of Plant Biology*, 57, 761-780.
41. Han K, Jeong HJ, Yang HB, Kang SM, Kwon JK, Kim S, Choi D, Kang BC (2016). An ultra-high-density bin map facilitates high-throughput QTL mapping of horticultural traits in pepper (*Capsicum annuum*). *DNA Research*, 23(2), 81-91.
42. Han YY, Ming F, Wang JW, Ye JQ, Shen L, Wang W, Zhao Y (2007). Molecular evolution and functional specialization of chalcone synthase superfamily from *Phalaenopsis* orchid. *Genetica*, 128(1-3), 429-438.
43. He Y, Zhu M, Wu J, Ouyang L, Wang R, Sun H, Yan L, Wang L, Xu M, Zhan H, Zhao Y (2020). Repurposing of anthocyanin biosynthesis for plant transformation and genome editing. *Frontiers in Genome Editing*, 2, 607982.
44. Holton TA, Cornish EC (1995). Genetics and biochemistry of anthocyanin biosynthesis. *Plant Cell*, 7(7), 1071-1083.
45. Huang D, Wang X, Tang Z, Yuan Y, Xu Y, He J, Jiang X, Peng SA, Li L, Butelli E, *et al* (2018). Subfunctionalization of the Ruby2-Ruby1 gene cluster during the domestication of citrus. *Nature Plants*, 4(12), 930-941.
46. Huang H, Xie S, Xiao Q, Wei B, Zheng L, Wang Y, Cao Y, Zhang X, Xie Q, Hu Z (2016). Sucrose and ABA regulate anthocyanin accumulation in radish taproot. *Frontiers in Plant Science*, 7, 1054.
47. Huang D, Yuan Y, Tang Z, Huang Y, Kang C, Deng X, Xu Q (2019). Retrotransposon promoter of Ruby1 controls both light- and cold-induced accumulation of anthocyanins in blood orange. *Plant, Cell & Environment*, 42(11), 3092-3104.
48. Jaakola L, Poole M, Jones MO, Kamarainen-Karppinen T, Koskimäki JJ, Hohtola A, Haggman H, Fraser PD, Manning K, King GJ, *et al* (2010). A SQUAMOSA MADS box gene involved in the regulation of anthocyanin accumulation in bilberry fruits. *Plant Physiology*, 153(4), 1619-1629.
49. Jaakola L (2013). New insights into the regulation of anthocyanin biosynthesis in fruits. *Trends in Plant Science*, 18(9), 477-483.
50. Jiang L, Yue M, Liu Y, Zhang N, Lin Y, Zhang Y, Wang Y, Li M, Luo Y, Zhang Y, *et al* (2023). A novel R2R3-MYB transcription factor FaMYB5 positively regulates anthocyanin and proanthocyanidin biosynthesis in cultivated strawberries (*Fragaria x ananassa*). *Plant Biotechnology Journal*, 21(6), 1140-1158.
51. Jiu S, Guan L, Leng X, Zhang K, Haider MS, Yu X, Zhu X, Zhang C, Fang J (2021). The identification and functional analysis of GRAS gene family in grapevine (*Vitis vinifera* L.). *Plant Signaling & Behavior*, 16(2), 1849490.
52. Kadomura-Ishikawa Y, Miyawaki K, Takahashi A, Masuda T, Noji S (2015). RNAi-mediated silencing and overexpression of the FaF3H gene and its effect on flavonoid biosynthesis in strawberry (*Fragaria x ananassa*) fruit. *Plant Biotechnology*, 32(1), 1-9.
53. Kadomura-Ishikawa Y, Miyawaki K, Noji S, Takahashi A (2013). Phototropin 2 is involved in blue light-induced anthocyanin accumulation in *Fragaria x ananassa* fruits. *Journal of Plant Research*, 126(6), 847-857.
54. Karppinen K, Lafferty DJ, Albert NW, Mikkola N, McGhie T, Allan AC, Espley RV, Jaakola L (2021). MYBA and MYBPA transcription factors co-regulate anthocyanin biosynthesis in blue-coloured berries. *New Phytologist*, 232(4), 1350-1367.
55. Khoo HE, Azlan A, Tang ST, Lim SM (2017). Anthocyanidins and anthocyanins: colored pigments as food, pharmaceutical ingredients and the potential health benefits. *Food & Nutrition Research*, 61(1), 1361779.
56. Klimek-Chodacka M, Oleszkiewicz T, Lowder LG, Qi Y, Baranski R (2018). Efficient CRISPR/Cas9-based genome editing in carrot cells. *Plant Cell Reports*, 37(4), 575-586.
57. Kobayashi S, Goto-Yamamoto N, Hirochika H (2004). Retrotransposon-induced mutations in grape skin color. *Science*, 304(5673), 982.
58. Kobayashi S, Ishimaru M, Hiraoka K, Honda C (2002). Myb-related genes of the Kyoho grape (*Vitis labruscana*) regulate anthocyanin biosynthesis. *Planta*, 215(6), 924-933.
59. Koes R, Verweij W, Quattrocchio F (2005). Flavonoids: a colorful model for the regulation and evolution of biochemical pathways. *Trends in Plant Science*, 10(5), 236-242.
60. Lahner B, Gong J, Mahmoudian M, Smith EL, Abid KB, Rogers EE, Guerinot ML, Harper JF, Ward JM, McIntyre L, *et al* (2003). Genomic scale profiling of nutrient and trace elements in *Arabidopsis thaliana*. *Nature Biotechnology*, 21(10), 1215-1221.

61. Lai B, Du LN, Liu R, Hu B, Su WB, Qin YH, Zhao JT, Wang HC, Hu GB (2016). Two LcbHLH transcription factors interacting with LcMYB1 in regulating late structural genes of anthocyanin biosynthesis in *Nicotiana tabacum* and *Litchi chinensis* during anthocyanin accumulation. *Frontiers in Plant Science*, 7, 166.
62. Langfelder P, Horvath S (2008). WGCNA: an R package for weighted correlation network analysis. *BMC Bioinformatics*, 9, 559.
63. Li D, Wang P, Luo Y, Zhao M, Chen F (2017). Health benefits of anthocyanins and molecular mechanisms: update from recent decade. *Critical Reviews in Food Science and Nutrition*, 57(8), 1729-1741.
64. Lin Y, Jiang L, Chen Q, Li Y, Zhang Y, Luo Y, Zhang Y, Sun B, Wang X, Tang H (2018). Comparative transcriptome profiling analysis of red- and white-fleshed strawberry (*Fragaria x ananassa*) provides new insight into the regulation of the anthocyanin pathway. *Plant and Cell Physiology*, 59(9), 1844-1859.
65. Lin-Wang K, Bolitho K, Grafton K, Kortstee A, Karunairetnam S, McGhie TK, Espley RV, Hellens RP, Allan AC (2010). An R2R3 MYB transcription factor associated with regulation of the anthocyanin biosynthetic pathway in Rosaceae. *BMC Plant Biology*, 10, 50.
66. Liu Y, Tikunov Y, Schouten RE, Marcelis LFM, Visser RGF, Bovy A (2018). Anthocyanin biosynthesis and degradation mechanisms in Solanaceous vegetables: a review. *Frontiers in Chemistry*, 6, 52.
67. Ma X, Chen X, Xu B, Cheng Y, Wang X, Yang M, Zhao Q, He J, He L, Zhang Y (2020). Single-cell RNA sequencing analyses of cell heterogeneity and formation of Nudt13-marked germ cells in mouse testes. *Cell Proliferation*, 53(9), e12888.
68. Manolescu BN, Oprea E, Mititelu M, Ruta L, Farcasanu I (2019). Dietary anthocyanins and stroke: a review of pharmacokinetic and pharmacodynamic studies. *Nutrients*, 11(7), 1479.
69. Medina-Puche L, Cumplido-Laso G, Amil-Ruiz F, Hoffmann T, Ring L, Rodriguez-Franco A, Caballero JL, Schwab W, Munoz-Blanco J, Blanco-Portales R (2014). MYB10 plays a major role in the regulation of flavonoid/phenylpropanoid metabolism during ripening of *Fragaria x ananassa* fruits. *Journal of Experimental Botany*, 65(2), 401-417.
70. Meir S, Kochanek B, Cherry-Rozenberg S, Rosenberger I, Fait A, Rogachev I (2021). Transcriptomic and metabolomic dynamics of the shoot apical meristem during floral transition in tomato. *Plant Journal*, 106(6), 1547-1564.
71. Ming R, Zhang T, Wu Y, Xu J, Liu Z, Hu J (2022). CRISPR-Act3.0 for highly efficient multiplexed gene activation in plants. *Nature Plants*, 8(9), 1035-1051.
72. Naing AH, Kim CK (2018). Roles of R2R3-MYB transcription factors in transcriptional regulation of anthocyanin biosynthesis in horticultural plants. *Plant Molecular Biology*, 98(1-2), 1-18.
73. Noman AA, Amin MZ, Uddin MJ, Sarker MMR (2025). Nutraceutical potential of anthocyanins: a comprehensive treatise. *Food Science & Nutrition*, 13(5), e70164.
74. Passeri V, Koes R, Quattrocchio FM (2016). New challenges for the design of high value plant products: stabilization of anthocyanins in plant vacuoles. *Frontiers in Plant Science*, 7, 153.
75. Pecker I, Gabbay R, Cunningham FX, Hirschberg J (1996). Cloning and characterization of the cDNA for p-carotene desaturase from tomato reveals a novel type of ribosome-binding site. *Plant Molecular Biology*, 30(4), 807-819.
76. Perteu M, Perteu GM, Antonescu CM, Chang TC, Mendell JT, Salzberg SL (2015). StringTie enables improved reconstruction of a transcriptome from RNA-seq reads. *Nature Biotechnology*, 33(3), 290-295.
77. Petroni K, Tonelli C (2011). Recent advances on the regulation of anthocyanin synthesis in reproductive organs. *Plant Science*, 181(3), 219-229.
78. Rapisarda P, Fanella F, Maccarone E (2000). Reliability of analytical methods for determining anthocyanins in blood orange juices. *Journal of Agricultural and Food Chemistry*, 48(6), 2249-2252.
79. Ravaglia D, Espley RV, Henry-Kirk RA, andreotti C, Ziosi V, Hellens RP, Costa G, Allan AC (2013). Transcriptional regulation of flavonoid biosynthesis in nectarine (*Prunus persica*) by a set of R2R3 MYB transcription factors. *BMC Plant Biology*, 13, 68.
80. Roller A, Baumgartner S, Windhausen P, Muenchhoff A, Weigand E, Beckmann M, Krishnamoorthy P, Bienert G, Brueck H (2025). Genetic variation and quantitative trait loci analysis of the maize ionome in response to phosphorus fertilisation. *Plant, Cell & Environment*, 48(9), 6521-6538.
81. Sakuma Y, Liu Q, Dubouzet JG, Abe H, Shinozaki K, Yamaguchi-Shinozaki K (2002). DNA-binding specificity of the ERF/AP2 domain of Arabidopsis DREBs, transcription factors involved in dehydration- and cold-inducible gene expression. *Biochemical and Biophysical Research Communications*, 290(3), 998-1009.
82. Salt DE, Baxter I, Lahner B (2008). Ionomics and the study of the plant ionome. *Annual Review of Plant Biology*, 59, 709-733.
83. Savoi S, Wong DCJ, Degu A, Herrera JC, Bucchetti B, Peterlunger E, Fait A, Mattivi F, Castellarin SD (2017). Multi-omics and integrated network analyses reveal new insights into the systems relationships between metabolites, structural genes and transcriptional regulators in developing grape berries (*Vitis vinifera* L.) exposed to water deficit. *Frontiers in Plant Science*, 8, 1124.
84. Shoji T, Kajikawa M, Hashimoto T (2013). Clustered transcription factor genes regulate nicotine biosynthesis in tobacco. *Plant Cell*, 25(10), 3897-3908.
85. Sperotto RA, Ricachenevsky FK, Waldow VA, Fett JP (2012). Iron biofortification in rice: it's a long way to the top. *Plant Science*, 190, 24-39.

86. Strygina KV, Kutuzova SN, Belousov AA, Khlestkina EK (2019). Identification and characterization of regulatory network components for anthocyanin synthesis in barley aleurone. *BMC Plant Biology*, 19(Suppl 1), 1-9.
87. Telias A, Wang K, Stevenson DE, Cooney JM, Hellens RP, Allan AC (2011). Apple skin patterning is associated with differential expression of MYB10. *BMC Plant Biology*, 11, 93.
88. Umemura H, Otagaki S, Wada M, Kondo S, Matsumoto S (2013). Expression analysis of the MdMYB110a gene, which encodes an R2R3 MYB transcription factor, from the red flesh sport of 'Fuji' apple. *Plant Cell Reports*, 32(12), 1893-1901.
89. Vasconcelos MW, Gruissem W, Bhullar NK (2017). Iron biofortification in the 21st century: setting realistic targets, overcoming obstacles and new strategies for healthy nutrition. *Current Opinion in Biotechnology*, 44, 8-15.
90. Wang X, Yuan Y, Wang L, Xu Q, Huang D (2024). The R2R3 MYB Ruby1 is activated by two cold-responsive ethylene response factors, via the retrotransposon in its promoter, to positively regulate anthocyanin biosynthesis in citrus. *Plant Journal*, 118(6), 1861-1878.
91. Wang N, Jiang S, Zhang Z, Fang H, Xu H, Wang Y, Chen X (2019). *Malus sieversii*: the origin, flavonoid synthesis mechanism and breeding of red-skinned and red-fleshed apples. *Horticulture Research*, 5, 70.
92. Wang Y, Wang Y, Song Z, Zhang H (2016). Repression of MYBL2 by both microRNA858a and MYBL2 itself upregulates flavonoid biosynthesis genes and anthocyanin accumulation in *Arabidopsis*. *Plant Cell*, 28(11), 2866-2883.
93. White PJ, Broadley MR (2009). Biofortification of crops with seven mineral elements often lacking in human diets - iron, zinc, copper, calcium, magnesium, selenium and iodine. *New Phytologist*, 182(1), 49-84.
94. Winkel-Shirley B (2001). Flavonoid biosynthesis: a colorful model for genetics, biochemistry, cell biology and biotechnology. *Plant Physiology*, 126(2), 485-493.
95. Xie Q, Hu Z, Zhang Y, Tian S, Wang Z, Zhao Z, Yang Y, Chen G (2014). Accumulation and molecular regulation of anthocyanin in purple tumorous stem mustard (*Brassica juncea* var. *tumida* Tsen et Lee). *Journal of Agricultural and Food Chemistry*, 62(29), 7813-7821.
96. Xing S, Chen K, Zhu H, Zhang R, Zhang H, Li B, Gao C (2020). Fine-tuning sugar content in strawberry. *Genome Biology*, 21(1), 230.
97. Xu W, Dubos C, Lepiniec L (2015). Transcriptional control of flavonoid biosynthesis by MYB-bHLH-WDR complexes. *Trends in Plant Science*, 20(3), 176-185.
98. Yao G, Ming M, Allan AC, Gu C, Li L, Wu X, Wang R, Chang Y, Qi K, Zhang S, Wu J (2017). Map-based cloning of the pear gene MYB114 identifies an interaction with other transcription factors to coordinately regulate fruit anthocyanin biosynthesis. *Plant Journal*, 92(3), 437-451.
99. Zhang Y, Liu Z, Liu R, Hao H, Bi Y (2013). Ultraviolet-B irradiation and temperature affect the fruit anthocyanin content and skin color of prevernal 'Hongfeng' peach. *Journal of Integrative Agriculture*, 12(11), 2058-2065.
100. Zhang Y, Butelli E, Alseekh S, Tohge T, Rallapalli G, Luo J, Kwar PG, Hill L, Santino A, Fernie AR, Martin C (2015). Multi-level engineering facilitates the production of phenylpropanoid compounds in tomato. *Nature Communications*, 6, 8635.
101. Zhang L, Hu J, Han X, Li J, Gao Y, Richards CM, Fang H, Zhao Y, Wang D, Wang Y, *et al* (2019). A high-quality apple genome assembly reveals the association of a retrotransposon and red fruit colour. *Nature Communications*, 10, 1494.
102. Zhang Y, Liu T, Meyer CA, Eeckhoutte J, Johnson DS, Bernstein BE, Nusbaum C, Myers RM, Brown M, Li W, Liu XS (2008). Model-based analysis of ChIP-Seq (MACS). *Genome Biology*, 9(9), R137.
103. Zhang A, Yang H, Ji S, Tian C, Chen N, Gong H, Li J (2022). Metabolome and transcriptome analyses of anthocyanin accumulation mechanisms reveal metabolite variations and key candidate genes involved in the pigmentation of *Prunus tomentosa* Thunb. cherry fruit. *Frontiers in Plant Science*, 13, 938908.
104. Zhao Y, Han Y (2023). Colorful hues: insight into the mechanisms of anthocyanin pigmentation in fruit. *Plant Physiology*, 192(3), 1718-1732.
105. Zhao M, Li J, Zhu L, Chang P, Li L, Zhang L (2019). Identification and characterization of MYB-bHLH-WD40 regulatory complex members controlling anthocyanidin biosynthesis in blueberry fruits development. *Genes*, 10(7), 496.
106. Zhou H, Lin-Wang K, Wang H, Gu C, Dare AP, Espley RV, He H, Allan AC, Han Y (2015). Molecular genetics of blood-fleshed peach reveals activation of anthocyanin biosynthesis by NAC transcription factors. *Plant Journal*, 82(1), 105-121.
107. Zoratti L, Karppinen K, Luengo Escobar A, Haggman H, Jaakola L (2014). Light-controlled flavonoid biosynthesis in fruits. *Frontiers in Plant Science*, 5, 534.